Lower expression of ER-α36 is associated with the development of endometrial hyperplasia in PCOS patients

Sheng-Li Lin1,2,3,*, Bin-Bin Tu1,2,3,*,#, Xiao-Guo Du1,2,3, Li-Ying Yan1,2,3 and Jie Qiao1,2,3

1Reproductive Medical Centre, Department of Obstetrics and Gynecology, Peking University Third Hospital, 2Key Laboratory of Assisted Reproduction, Ministry of Education and 3Beijing Key Laboratory of Reproductive Endocrinology and Assisted Reproduction, Beijing, China, #Present address: Department of Gynecological Oncology, Cancer Institute (Hospital), Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China, *Sheng-Li Lin, Bin-Bin Tu contributed equally to this work.

Summary. Objective: To explore the expression change of ER-α36 in endometria of PCOS patients. Methods: Proliferative endometria were collected and divided into three groups: CE group (n=30), proliferative endometria from control women; PCOSE group (n=30), proliferative endometria from PCOS patients; HPCOSE group (n=15), hyperplastic endometria from PCOS patients. The cellular localization of ER-α36 and ER-α66 was analyzed by immunohistochemistry, and expression of ER-α36 and ER-α66 was determined by immunohistochemistry and Real-Time Quantitative PCR. The correlation between serum hormone concentration and expression of ER-α36 and ER-α66 was analyzed. Results: ER-α36 was localized on the cell surface, cytoplasm and nucleus of glandular epithelial cells of both CE and PCOSE groups, mainly on the cell surface, and ER-α66 was localized on the nucleus. The protein and mRNA expression of ER-α36 was decreased from CE, PCOSE to HPCOSE group, while expression of ER-α66 showed no obvious difference among groups. The expression ratio of ER-α36 to ER-α66 of CE, PCOSE and HPCOSE group showed a decreasing tendency. The expression of ER-α36 and its expressive ratio to ER-α66 was negatively correlated with serum concentration of LH, LH/FSH, testosterone and androstenedione. Conclusions: Lower expression of ER-α36 is associated with the development of endometrial hyperplasia in PCOS patients.

Key words: Polycystic ovary syndrome, Estrogen receptor-α36, Endometrial hyperplasia

Introduction

Polycystic ovary syndrome (PCOS) is one of the most common endocrine disorders in women of reproductive age, which is characterized by oligomenorrhea or amenorrhea, hyperandrogenemia or hyperandrogenism and polycystic ovaries (Carmina, 2004). The prevalence of PCOS among women of reproductive age is 6.7% (Chang, 2004). The association between PCOS and endometrial hyperplasia has been reported for many years (Coulam et al., 1983; Dahlgren et al., 1991, 1992; Escobedo et al., 1991; Wild et al., 2000; Schmeler et al., 2005; Navaratnarajah et al., 2008), indicating higher incidence of endometrial hyperplasia in PCOS patients.

However, the molecular mechanism underlying higher prevalence of endometrial hyperplasia in PCOS patients is not totally understood. The multi-factorial and heterogenous natures of PCOS make it difficult to explore the risk factors associated with development of endometrial hyperplasia in PCOS patients. It may be attributed to persistent stimulation of endometrial tissue by estrogen (mainly estrone) without progesterone-induced inhibition of proliferative endometria and differentiation to secretory endometria (Balen, 2001; Hardiman et al., 2003). Some researchers found that a change in expression of ER-α66 (the classic estrogen receptor (α) was observed in endometria from PCOS patients, and that might lead to alteration of transcription
of genes involved in the cell cycle and induce development of endometrial hyperplasia in PCOS patients (Maliqueo et al., 2003; Quezada et al., 2006). However, the exact role of ER-α66 in endometria remains unclear as different investigations reported discrepant changes of expression of ER-α66.

Recently, ER-α36, a novel variant of estrogen receptor α, was identified and cloned with a molecular weight of 36 kDa. It was transcribed from previously unidentified promoter located in the first intron of the original classic estrogen receptor α 66 (ER-α66) gene (Wang et al., 2005). ER-α36 differs from ER-α66 by lacking both transcriptional activation domains (AF-1 and AF-2), but it retains DNA-binding domain and partial ligand-binding domains. It possesses a unique 27 amino acid domain that replaces the last 138 amino acids encoded by exons 7 and 8 of the ER-α66 gene. However, there is no clinical research on ER-α36 expression and its relationship with ER-α66 in endometrial hyperplasia in PCOS patients.

Therefore, the present research explored the role of ER-α36 and ER-α66 in the endometria of PCOS through examining the protein and mRNA expression of ER-α36 and ER-α66 in proliferative and hyperplastic endometria of PCOS women. These results will be useful for a better understanding of the high prevalence of endometrial hyperplasia in PCOS patients.

Materials and methods

Collection of samples

Women who received infertility treatment from Nov 2009 to Oct 2010 in Reproductive Medical Centre of Peking University Third Hospital were recruited. None of the women had received hormonal therapy or other medications for 3 months prior to recruitment into the study. PCOS was diagnosed on the basis of Rotterdam Consensus: oligo- or anovulation, clinical or biochemical evidence of hyperandrogenism and the presence of polycystic ovaries (Carmina, 2004). Women with hysteromyoma, endometriosis, endometrial polyps, endometritis and appendagitis were excluded. The control group was defined as non-PCOS women with tubal or male factor infertility.

The endometrial specimens were obtained with a pipelle suction curette from the corpus of the uterus. All the endometria were obtained in the follicular phase of the menstrual cycle and verified as proliferative phase by histopathological criteria of Noyes (Nejatbakhsh et al., 2012). The hyperplasia endometria of PCOS patients were diagnosed using the 2003 WHO classification (Steinbakk et al., 2011).

Endometria were classified as follows: CE group (n=30), proliferative endometria without hyperplasia from non-PCOS women with tubal or male factor infertility; PCOSE group (n=30), proliferative endometria without hyperplasia from PCOS patients; and HPCOSE group (n=15), hyperplasia endometria from PCOS patients. The diagnostic of endometrial hyperplasia was confirmed by histological studies by an experienced histopathologist.

Blood samples were collected on the 2nd to 5th day of menstrual cycle or when there was no follicle larger than 10 mm detected by transvaginal ultrasound. Total testosterone (TT), androstenedione (A) and estradiol were tested according to the Immulite 1000 assay based on chemiluminescence (DPC, USA).

The study was approved by the Ethics Committee of Peking University Third Hospital and all patients signed written informed consents.

Materials and reagents

ER-α36 specific antibody against the 20 unique amino acids at the C-terminal of ER-α36 was described before (Wang et al., 2005). The primers for specific amplification of cDNA of human ER-α36 (BX640939, 1145–1434 bp) were: forward, 5'-CAAGTGTTCTCCTGTTGCTAAGC-3' and reverse, 5'-TGTTGAGTGTTGGTTGCGAGG-3' (Invitrogen, USA); The primers for amplification of cDNA of human ER-α66 were: forward, 5 '-AAGTTCAGGCAACATTGGATG-3' and reverse, 5'-CCCTGGTACAGCTGTGTTAC-3' (Invitrogen, USA); Human glyceraldehyde -3-phosphate dehydrogenase (GAPDH) cDNA was amplified by the forward primer 5'-ACGGATTTGGTCGTATTGGG-3' and the reverse primer 5'-TGATTTGGAGGATCTCGC-3'.

Immunofluorescence and confocal microscopy

The cellular localization of ER-α36 was determined by indirect immunofluorescence, according to a previous method (Lin et al., 2009). Frozen sections of endometrial biopsies of 5 μm were fixed for 10 minutes in acetone stored overnight at 4°C and then rinsed with PBS for 10 min. After being permeabilized with 0.5% Triton X-100 for 60 min at room temperature, tissues were blocked in 10% BSA-supplemented PBS for 1.5 hours and incubated overnight at 4°C with anti-ER-α36-specific antibody or normal rabbit IgG. After three washes in PBS, the tissues were labeled with FITC-conjugated secondary antibody for 2 hours protected from light. After three washes in PBS, the DNA dye PI was used for nuclear staining. Microscopic analysis was performed using a Confocal Laser-Scanning Microscope (Zeiss LSM 510 META, Germany).

Immunohistochemistry

Tissue slides were deparaffinized with xylene and rehydrated through a graded alcohol series. Antigen retrieval was carried out by immersing the slides in natrium citrate buffer and boiling in a waterbath at 100°C for 30 minutes. The endogenous peroxidase activity was blocked by incubation in a 3% hydrogen peroxidise solution for 10 minutes. Endometrial sections were incubated overnight at 4°C with anti-ER-α36 specific antibody at 1:300 dilution in 2.5% BSA-PBS. After incubation, the sections were washed in PBS, incubated with biotinylated goat anti-rabbit IgG 1:100, washed again in PBS then labeled with streptavidin-horse radish peroxidase 1:100 for 30 min. The endogenous peroxidase activity was blocked by incubation in 3% hydrogen peroxidise solution for 10 minutes. Nonspecific staining was blocked using normal goat serum 1:100 for 30 min. Staining was developed with 3,3′-diaminobenzidine tetrahydrochloride (DAB) solution. The slides were counterstained with hematoxylin.
peroxide/methanol buffer for 10 minutes. The slides were rinsed in PBS and incubated with normal goat serum to block nonspecific staining. The slides were then incubated with ER-α36 specific antibody or normal rabbit IgG overnight at 4°C in a humidified chamber. The sections were incubated with the second antibody for 30 minutes. Diaminobenzidine was used as a chromogen, and sections were counterstained with hematoxylin. Before nuclear counterstaining, pictures were taken by Leica Q 550cw Optical microscope with a Leica DC 300F camera (Leica Company, Germany) and the staining intensity of glandular cells of endometria was evaluated by using Leica Qwin Standard V2.8 image software for the average optical density. As a negative control, duplicate sections were immunostained without exposure to primary antibodies, which was replaced by PBS.

**RNA extractions and cDNA synthesis**

Total RNA was extracted from tissues frozen in liquid nitrogen using TRIzol reagent according to the protocol provided by the manufacturer (Invitrogen, Carlsbad, CA, USA). Total RNA was reverse-transcribed into single-strand complementary DNA (cDNA) using moloney-murine leukemia virus (M-MLV) reverse transcriptase (TianGen, Beijing, China). Briefly, the RNA was denatured by heating for 5 min at 70°C followed by rapid cooling on ice, and then used for reverse transcription (5 μg of total RNA, 20 U of RNase inhibitor, 2.5 mmol/L of each dNTP, 10 μmol/L reverse primer and 200 U of M-MLV reverse transcriptase in a total volume of 20 μl). For reverse transcription, tubes were incubated at 42°C for 50 min.

**Real-Time Quantitative PCR**

SYBR Green Real-Time Quantitative PCR was performed using iQ5 Real-Time PCR Detection System (Bio-Rad, USA). The Real-Time Quantitative PCR reaction was carried out in triplicate for each sample. Briefly, 20 μl of reaction mixture containing 2 μl of cDNA template was amplified as follows: ER-α36 denaturation at 95°C for 15 min and 40 cycles at 95°C for 15 s, 68°C for 60 s; ER-α66, denaturation at 95°C for 15 min and 40 cycles at 95°C for 15 s, 63°C for 60 s. Quantitative analysis was performed using the comparative threshold cycle (CT) method (Livak and Schmittgen, 2001).

**Statistical analysis**

All data was analyzed using SPSS 18.0 software package (SPSS Inc., Chicago, IL, USA). The difference in expression of ER-α36 and ER-α66 among three groups was compared using the one-way ANOVA test or nonparametric Kruskal-Wallis H test. Correlation analysis was done using Pearson or Spearman correlation coefficient. p<0.05 was considered statistically significant.

**Results**

**Location of ER-α36 and ER-α66 in glandular epithelial cells of endometrium**

ER-α36 is a novel variant of ER-α66 generated by alternative promoter usage and alternative splicing. To examine the cellular localization of ER-α36, immunofluorescence assay was performed using ER-α36 specific antibody against the unique 20 amino acids at the C-terminal of ER-α36. As shown in Fig. 1, ER-α36 protein (green) was localized on the cell surface, cytoplasm and nucleus of glandular epithelial cells of endometria, mainly on the cell surface. Moreover, the results of immunohistochemistry also
affirmed the localization of ER-α36. The expression of ER-α36 is mainly localized on the cell surface in the epithelial cells of CE, PCOSE and HPCOSE (Fig. 2A-C).

Immunohistochemistry showed that ER-α66 was localized on the nucleus of glandular epithelial cells of CE, PCOSE and HPCOSE (Fig. 3A-C).

Expression change of ER-α36 and ER-α66 in endometria

The staining of ER-α36 gradually decreased from CE, PCOSE to HPCOSE group according to immunohistochemistry results. Furthermore, the staining intensity of ER-α36 of CE, PCOSE and HPCOSE groups was analyzed using mean optical density (OD) value. The mean OD value of ER-α36 of the three groups was 0.342±0.045, 0.311±0.027 and 0.282±0.022 (Fig. 2D) respectively, and had significant differences among groups (p<0.001). The mRNA expression of ER-α36 of the three groups is shown in Fig. 2E. The relative mRNA level of ER-α36 of CE, PCOSE and HPCOSE group was 43.612 (18.008-86.149), 15.324 (3.637-50.992) and 2.533 (0.758-10.289) respectively, and had significant differences among groups (p<0.001).

Analysis of staining intensity of ER-α66 in CE, PCOSE and HPCOSE group by immunohistochemistry showed that the mean OD value of ER-α66 was 0.398±0.058, 0.361±0.030 and 0.413±0.030 (Fig. 3D) respectively. The staining intensity between PCOSE group and HPCOSE group had a significant difference (p<0.001). The mRNA expression of ER-α66 of three groups is shown in Fig. 3E. The relative mRNA level of ER-α66 in CE, PCOSE and HPCOSE group was 8.021 (2.402-19.627), 6.571 (4.074-14.448) and 9.883 (5.756-25.107), and had no significant difference among groups (p=0.272).

Fig. 2. The expression change of ER-α36. The localization of ER-α36 was examined in proliferative endometria of control group (CE, A), proliferative endometria of PCOS group (PCOSE, B) and hyperplasia endometria of PCOS group (HPCOSE, C). D. The mean optical density (OD) value of ER-α36 in CE, PCOSE and HPCOSE groups. The mean OD value of the three groups was 0.342±0.045, 0.311±0.027 and 0.282±0.022 respectively. *: difference between groups were significant (p<0.001). E. The expression level of ER-α36 mRNA in CE, PCOSE and HPCOSE groups. The relative mRNA level of the three groups was 43.612 (18.008-86.149), 15.324 (3.637-50.992) and 2.533 (0.758-10.289) respectively, shown as median (25% quartile-75% quartile). #: difference between groups was significant (p<0.001).
Relative expression of ER-α36 to ER-α66 in endometria

We further evaluated the expression ratio of ER-α36 to ER-α66 in endometria. As shown in Fig. 4A, the mean OD ratio of ER-α36 to ER-α66 was 0.926 (0.854-0.975) in CE, 0.879 (0.812-0.965) in PCOSE and 0.709 (0.696-0.738) in HPCOSE respectively, and there was a significant difference between CE and HPCOSE, PCOSE and HPCOSE (p<0.001). The mRNA expression ratio of ER-α36 to ER-α66 was 5.764 (1.761-13.866) in CE, 1.733 (0.726-5.902) in PCOSE and 0.591 (0.067-0.771) in HPCOSE respectively, and there was a significant difference among groups (p<0.05) (Fig. 4B).

Correlation between expression of ERs and serum hormone level

We further analyzed the effect of serum hormone level on expression of ERs among three groups, including luteinizing hormone (LH), follicle stimulating hormone (FSH), LH/FSH, estradiol, testosterone and androstenedione.

We found no significant correlation between serum hormone level and expression of ER-α66, neither at

Table 1. Correlation between expression level of ER-α36 and serum hormone level.

<table>
<thead>
<tr>
<th></th>
<th>LH</th>
<th>LH/FSH</th>
<th>Estradiol</th>
<th>Testosterone</th>
<th>Androstenedione</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mean OD value of ER-α36</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>r</td>
<td>-0.48</td>
<td>-0.57</td>
<td>-0.334</td>
<td>-0.52</td>
<td>-0.469</td>
</tr>
<tr>
<td>p</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
<td>0.02</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Relative mRNA expression of ER-α36</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>r</td>
<td>-0.347</td>
<td>-0.803</td>
<td>-0.403</td>
<td>-0.233</td>
<td>-0.254</td>
</tr>
<tr>
<td>p</td>
<td>0.01</td>
<td>&lt;0.001</td>
<td>&lt;0.001</td>
<td>0.042</td>
<td>0.028</td>
</tr>
</tbody>
</table>

Fig. 3. The expression change of ER-α66. The localization of ER-α66 was examined in proliferative endometria of control group (CE, A), proliferative endometria of PCOS group (PCOSE, B) and hyperplasia endometria of PCOS group (HPCOSE, C). D. The mean optical density (OD) value of ER-α66 in CE, PCOSE and HPCOSE groups. The mean OD value of the three groups was 0.398±0.058, 0.361±0.030 and 0.413±0.030. *: p<0.001 compared to PCOSE vs. HPCOSE group. E. The expression level of ER-α66 mRNA in CE, PCOSE and HPCOSE groups. The relative mRNA level of the three groups was 8.021 (2.402-19.627), 6.571 (4.074-14.448) and 9.883 (5.756-25.107) respectively, shown as median (25% quartile-75% quartile), and had no significant difference among groups (p=0.272).
protein level nor at mRNA level. However, the mean OD value of ER-α36 of the three groups was negatively correlated with serum level of LH (r=-0.48, p<0.001), LH/FSH (r=-0.57, p<0.001), estradiol (r=-0.334, p=0.02), testosterone (r=-0.520, p<0.001) androstenedione (r=-0.469, p<0.001). The relative mRNA level of ER-α36 of the three groups was also negatively correlated with serum level of LH (r=-0.347, p=0.01), LH/FSH (r=-0.603, p<0.001), estradiol (r=-0.403, p<0.001), testosterone (r=-0.233, p=0.042) and androstenedione (r=-0.254, p=0.028) (table 1).

Discussion

Recently, ER-α36, a novel variant of estrogen receptor α was identified and cloned. Increasing research focused on its expression and functions. Our present study first explored the role of ER-α36 and its relative expression level to classic estrogen receptor, ER-α66 in endometrial hyperplasia in PCOS patients. The present study found that ER-α36 was mainly localized on the cell surface of glandular epithelial cells of endometria in both control and PCOS groups, and ER-α66 was localized on the nucleus. The protein and mRNA level of ER-α36 was gradually decreased from CE, PCOSE to HPCOSE group. However, there was no obvious difference in expression of ER-α66 among the three groups. The expression ratio of ER-α36 to ER-α66 presented a decreasing tendency from CE, PCOSE to HPCOSE group. Furthermore, the expression of ER-α36 was negatively correlated with serum concentration of LH, LH/FSH, estradiol and testosterone.

The immunofluorescence and immunohistochemistry results have shown that ER-α36 was localized on the cell surface, cytoplasm and nuclei of glandular epithelial cells of endometria, mainly on the cell surface, which is consistent with recent findings (Lin et al., 2010). ER-α66, the classic estrogen receptor, belongs to the nuclear receptor family of transcription factors and its transcriptional activation is mediated by two activation function domains, AF-1 and AF-2. However, ER-α36, the new variant of estrogen receptor, is generated from a promoter located in the first intron of the ER-α66 gene. ER-α36 differs from ER-α66 by lacking both AF-1 and AF-2, but it retains the DNA-binding domain, partial dimerization and ligand-binding domains. Several clinical investigations suggested that ER-α36 played negative role in cancers, such as colorectal cancer and breast cancer (Jiang et al., 2008; Zheng et al., 2010). Present immunohistochemistry and Real-Time PCR analysis demonstrated that the expression of ER-α36 and its relative expression level to ER-α66 was lower in HPCOSE group than in CE and PCOSE groups. Wang found that ER-α36 inhibits the transcription activities mediated by AF-1 and AF-2 domains of ER-α66 (Wang et al., 2006). It is possible that lower expression of ER-α36 induced ER-α66 transcription activity and then led to endometrial hyperplasia. However, the exact role of ER-α36 needs further research.

Our research also found that the expression level of ER-α36 was negatively correlated with serum level of LH, LH/FSH, estradiol, testosterone and androstenedione. Previous studies suggested that serum level of LH, LH/FSH, testosterone and androstenedione was elevated in PCOS patients (Skampardonis et al., 2011; Ma et al., 2011). High tissue concentration of androstenediol is generated in the PCOS endometrium,
which possesses estrogenic activity (Wood and Strauss, 2002; Jamnongjit and Hammes, 2006; Plaza et al., 2010). It is possible that the elevated molecules with estrogenic activity in endometrium influence the expression of ER-α36 and further weaken its dominant-negative effect in the ER-α66, which induces endometrial hyperplasia in PCOS patients. However, the mechanism should be explored in the future.

In summary, we report here that ER-α36 plays a negative role in endometrial hyperplasia of PCOS patients. Our present results give important information to further understand the mechanism of higher prevalence of endometrial hyperplasia in PCOS patients. The role of ER-α36 in endometria of PCOS patients will provide useful information for designing new drugs for the treatment of endometrial problems of PCOS patients.

Acknowledgements: We would thank Dr. Zhao-Yi Wang for kindly providing ER-α36 specific antibody and reviewing the manuscript. This study was supported by National Natural Science Funds for Distinguished Young Scholars (30825038), National Scientific and Technical Supporting Programs funded by the Ministry of Science and Technology of China (2007BAI04B04), National Natural Science Funds for China for Young Scholars (81200437).

References


Plaza F., Gabler F., Romero C., Vantman D., Valladares L. and Vega M. (2010). The conversion of dehydroepiandrosterone into androst-5-ene-3beta,17beta-diol (androstenediol) is increased in endometria from untreated women with polycystic ovarian syndrome. Steroids 75, 810-817.


Accepted May 15, 2013

ER-α36 is associated with endometrial hyperplasia