

Invited Review

Mechanisms underlying eosinophil trafficking and their relevance *in vivo*

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Summary. After their formation in the bone marrow, eosinophils circulate with a short half-life and are distributed throughout the body, especially in mucosal and sub-mucosal regions. Although a small amount of these cells are normally seen in healthy tissue, blood and tissue eosinophilia is a hallmark of helminthic and allergic diseases. The role of eosinophils in the normal physiology of mucosal tissues is not understood, but there is good evidence to demonstrate that these cells protect the host at least against some intestinal helminths, specially those with a lung cycle. In addition, there are now many data that support a role for eosinophils in the pathophysiology of allergic diseases, such as asthma. Because helminthic diseases have been largely controlled in developed countries, there has been much interest in the development of drugs which affect eosinophil migration and/or activation in the tissue and which may, thus, be useful in the treatment of allergic conditions. The understanding of the mechanisms controlling eosinophil trafficking and/or activation are essential in the development of anti-eosinophil-based therapeutic strategies. The present paper reviews aspects of eosinophil biology with emphasis on the role of eosinophils in parasitic infections and allergy, the basic mechanisms underlying the trafficking of eosinophils into tissue and how these can be modulated pharmacologically.

Key words: Eosinophil, Allergy, Helminths, Trafficking, Interleukin-5, Cell adhesion molecules

Eosinophil morphology and granules

Eosinophils are a type of granulocyte derived from the bone marrow and distinguished by their morphological features, constituents, products, and

association with specific diseases. These cells are present in blood and characteristically in tissues with an epithelial interface with the environment, such as the respiratory, gastrointestinal, and lower genitourinary tracts (Weller, 1991). In blood, eosinophils account for only 1 to 3 percent of peripheral leukocytes in healthy subjects, and the upper limit of the normal range is 350 cells/ml of blood. The normal numbers of eosinophils in tissues is not known, but there are normally many more cells in tissues than in blood (Weller, 1991). In addition, as we will discuss below, eosinophil numbers in blood and tissue increase several times in the presence of certain conditions, such as asthma and helminth infection.

Typically, human eosinophils measure 8 µm in diameter, have a bilobed nucleus and contain characteristic granules that are known to have an intense avidity for eosin dye (Hirsch and Hirsch, 1980). Three types of granules are present in their cytoplasm: (i) primary granules, which are round, uniformly electron dense, and characteristically present in eosinophilic promyelocytes; (ii) specific or secondary granules, which are composed of an electron-dense core and an electron-lucent matrix; and (iii) small granules (Dvorak et al., 1988, 1991). Other cytoplasmic structures of the eosinophil are lipid bodies, which are non-membrane-bound, lipid-rich inclusions found in many types of cells (Dvorak et al., 1983). The latter structure appears to be particularly important for the production of lipid mediators by activated eosinophils (Bozza et al., 1997).

Eosinophils can express receptors for several molecules, including immunoglobulins (IgG, IgE, IgA), complement (C1q, C3b/C4b, iC3b, C5a), cytokines (IL-3, IL-5, GM-CSF), chemokines (eotaxin, eotaxin-2, RANTES, MCP-3, MCP-4), lipid mediators (PAF, LTB₄) and steroids (estrogens, glucocorticoids). It is via these receptors that eosinophils receive messages from and respond to their environment. Thus, the importance of each of these receptors will be dictated according to the environment in which the cells are situated. In addition to the receptors cited above, eosinophils express a range of cluster-determinant (CD) antigens on their surface. Although the function of several CD antigens is

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not known, their position on the plasma membrane suggests they may play an important role in cell/cell contact and communication. Among the CD antigens expressed in eosinophils, there are several cell adhesion molecules (CAMs) - CD11/CD18, VLA-4 ($\alpha 4\beta 1$), $\alpha 4\beta 7$ and CD62L (L-selectin).

Secretory products and cytokines

The ability of eosinophils to cause damage to cells or tissues appears to be related to their great ability to secrete a range of substances which include cationic proteins (see Table 1), cytokines, chemokines, lipid mediators and oxygen-derived radicals (Table 2). The action of each individual substance is beyond the scope of this review, but the wide array of substances produced and secreted gives a good indication of the ability of eosinophils to affect different cells or systems (see Giembycz and Lindsay, 1999 for review). Although the cationic proteins are usually thought to be specifically found in eosinophils, basophils have been shown to contain about one fourth as much MPB as eosinophils, and to contain detectable amounts of EDN, ECP and EPO, although at levels lesser than 7% of those in eosinophils (Abu-Ghazaleh et al., 1992). In addition, small amounts of EDN and ECP are also present in

neutrophils and could be synthesized by these cells (Abu-Ghazaleh et al., 1992).

Birth, life and death (IL-5)

Eosinophils are produced in bone marrow from pluripotential stem cells. The latter differentiate first into hybrid precursors with properties of basophils and eosinophils and then into a separate eosinophil lineage (Boyce et al., 1995). Three cytokines – interleukin-3 (IL-3), interleukin-5 (IL-5) and granulocyte-macrophage colony stimulating factor (GM-CSF) – have an essential role in the production of eosinophils. Interestingly, these cytokines are encoded by closely linked genes on chromosome 5q31 and bind to receptors that have a common beta chain and different alpha chains. Interleukin-5 (also known as eosinophil-differentiation factor) is the most specific colony factor for the eosinophil lineage and is responsible for selective differentiation of eosinophils (Sanderson, 1992). The critical role of IL-5 in the production of eosinophils is best demonstrated by genetic manipulation in mice. Overproduction of IL-5 in transgenic mice results in profound eosinophilia (Sanderson, 1992), and deletion of the IL-5 gene is accompanied by a marked reduction of eosinophils in the blood and tissue of naive animals or of sensitized animals after an allergen challenge (Foster et al., 1996; Kopf et al., 1996). In addition to a critical role for IL-5, much recent interest has been placed in the understanding of the role chemokine eotaxin and the eotaxin receptor (CCR3) play in the production and release of eosinophils from the bone marrow. It has been recently demonstrated that eotaxin alone or acting synergistically with IL-5 stimulates the release of eosinophils from bone marrow into the peripheral circulation (Collins et al., 1995). In addition, eotaxin was effective as GM-CSF at inducing eosinophil differentiation both *in vivo* and *in vitro* in mice (Peled et al., 1998). Thus, eotaxin induces not only the release of eosinophils from bone marrow, but also the generation of leukocytes needed for an inflammatory reaction. This is in line with findings that eotaxin knock-out mice had considerably lower eosinophil counts than control littermates in one study (Rothenberg et al., 1997). However, the latter findings have been recently disputed when the eotaxin gene was knocked out in mice from another genetic background (C57Bl6 x 129/Sv) (Yang et al., 1998).

In blood, eosinophils circulate with a half-life of approximately 18 hours (Spry, 1993). However, in

Table 1. Cationic proteins and other enzymes present in eosinophil granules

CATIONIC PROTEIN	MAIN ACTIVITY DESCRIBED
MBP (14 KD) major basic protein	- toxic to helminthic and protozoan parasites, tumor cells, bacteria, host cells - causes bronchospasm - induces histamine release - degranulates basophils - increases the expression of IL-8 mRNA
EPO (15-55 KD) Eosinophil peroxidase	- toxic to helminthic and protozoan parasites, bacteria, tumor cells and host cells. - induces histamine release - inactivates the peptide-leukotrienes - converts LTC ₄ to all-trans isomers of LTB ₄
ECP (18-21 KD) Eosinophil cationic protein	- toxic to helminthic and protozoan parasites, bacteria and host cells. - promotes degranulation of mast cells - neurotoxin
EDN (18 KD) Eosinophil-derived neurotoxin	- neurotoxin

Table 2. Eosinophil-derived substances

CLASS OF PRODUCTS	CYTOKINES	CHEMOKINES	LIPID MEDIATORS	NEUROPEPTIDES	OXYGEN-DERIVED RADICALS
Products known to be secreted	GM-CSF, IL-3, IL-5, IL-1 α , IL-2, IL-4, IL-6, IL-10, IL-16, TGF- β , TNF α , TGF- α	RANTES, eotaxin, MIP-1 α , IL-8, IP-10, MIG	PGE ₂ , PGD ₂ , PGF _{2α} , Thromboxane, LTC ₄ , PAF	SP, VIP, CGRP	O ₂ ⁻ , H ₂ O ₂ , OH ⁻ , NO, singlet oxygen

tissues eosinophils may survive for weeks (Spry, 1988), depending on cell matrix interactions and the cytokines present in the microenvironment (Rothenberg et al., 1987). For example, the VLA-4-dependent interaction of eosinophils with fibronectin induces the autocrine secretion of GMC-SF by eosinophils leading to their increased survival *in vitro* (Anwar et al., 1993).

Like other cells, eosinophils can be triggered to undergo programmed cell death or apoptosis, a process accompanied by condensation of the cytoplasm, segmentation of the nucleus and extensive degradation of chromosomal DNA, via specific surface death receptors (Simon and Alam, 1999). One of these death receptors expressed by eosinophils is CD95 (Fas/APO-1) (Matsumoto et al., 1995; Tsuyuki et al., 1995; Druilhe et al., 1996; Hebestreit et al., 1996). The ligand of CD95 (CD95L, FasL, APO-1L) is highly expressed by activated T cells (Green and Ware, 1997). The activation of CD95 leads to stimulation of a protease cascade, which, when started, is irreversible (Nagata, 1997). These proteases belong to the IL-1-converting enzyme (ICE) family of cysteine proteases, now called caspases (Alnemri et al., 1996), and appear to be directly responsible for the induction of apoptosis. There is evidence that at least two members of this family, caspase 3 and caspase 8, are involved in the regulation of eosinophil apoptosis (Simon and Alam, 1999). Inhibition of eosinophil apoptosis can be achieved by at least two mechanisms: (1) increased expression of eosinophil survival factors, and (2) disruption of death signals (Simon and Alam, 1999). *In vitro*, eosinophil survival can be maintained by the eosinophil haematopoietins, IL-3, IL-5 and GM-CSF (Her et al., 1991; Yamaguchi et al., 1991), that stimulate an anti-apoptotic signalling pathway via the common β chain of their receptors (Simon and Alam, 1999). Stimulation of eosinophils with IL-5 or other hematopoietins results in tyrosine phosphorylation of kinases Lyn (Pazdrak et al., 1995a; Yousefi et al., 1996), Jak1 (Ogata et al., 1998) and Jak2 (Pazdrak et al., 1995b; van der Bruggen et al., 1995; Simon et al., 1997), that activate the Stat family of nuclear factors. Indeed, IL-5 activates Stat 1 (Pazdrak et al., 1995b; van der Bruggen et al., 1995), Stat 3 (Caldenhoven et al., 1995) and Stat5 (Mui et al., 1995) nuclear factors. This pathway can be blocked by TGF- β , for example, that inhibits the tyrosine phosphorylation of Jak2 and Lyn tyrosine kinases (Pazdrak et al., 1995a). Besides the cytokines, nitric oxide can also disrupt the apoptotic signaling pathways initiated via CD95 (Hebestreit et al., 1998).

The role of eosinophil apoptosis in the control of eosinophil accumulation in tissue is not entirely understood. Theoretically, induction of eosinophil apoptosis in tissue would resolve tissue eosinophilia without activation of local inflammatory responses. However, to date this hypothesis has only been shown convincingly in the study by Tsuyuki and colleagues (1995) who showed that local treatment of mice with anti-CD95 resolved the lung eosinophilia following

challenge of sensitised mice with ovalbumin.

Eosinophil trafficking - Cell adhesion molecules

In response to an appropriate stimulus, circulating leukocytes interact with endothelial cells prior to leaving blood vessels and entering the tissue. The current model for the accumulation of leukocytes into tissues of the systemic circulation predicts that there are at least three stages of leukocyte/endothelial cell interaction (Carlos and Harlan, 1994; Springer, 1994; Teixeira et al., 1995). Initially, circulating leukocytes are captured and roll on the endothelial cells of post-capillary venules. The rolling leukocyte may then be activated by chemoattractants (e.g. eotaxin, interleukin-8, LTB4) and this leads to upregulation and increased avidity of integrins present on the surface of the leukocyte (e.g. CD11/CD18 and VLA-4). Integrins mediate the firm adhesion of activated leukocytes to endothelial cells by binding to ligands including ICAM-1 and VCAM-1. The leukocytes are then able to migrate to the interstitium, a process that also involves adhesion molecules, including the integrins and PECAM-1 (CD31), present at intercellular junctions. Here, the intention is not to review the basic cell adhesion pathways, but to highlight studies evaluating the effects of cell adhesion-based strategies on eosinophil recruitment *in vivo*.

There are several differences in the adhesion pathways utilised by eosinophils which are specific for these cells in relation to other leukocytes, specially neutrophils (Teixeira et al., 1995). These differences could help in the development of eosinophil-specific therapies without unwarranted side-effects on the host defence properties of other cell types (Teixeira et al., 1995). Of the known cell adhesion molecules much interest has been placed in understanding the role of selectins and the integrins CD18 and VLA-4 in mediating eosinophil recruitment *in vivo* (Lobb and Hemler, 1994; Teixeira et al., 1995) (see Table 3).

The first step in the adhesion cascade is the loose interaction between the circulating eosinophil and endothelial cells. This process is named rolling and is mediated by the selectin family of adhesion molecules present on eosinophils (L-selectin) or endothelial cells (P- and E-selectin) and their carbohydrate-expressing ligands (eg. PSGL-1) (Carlos and Harlan, 1994; Varki, 1994). The exact interplay between these molecules is not fully understood, although recent observations suggest that neutrophil rolling can be mediated by P-selectin and L-selectin sequentially (reviewed by Ley and Tedder, 1995; Kansas, 1996). Moreover, it is clear from studies assessing neutrophil migration *in vivo* that the function of selectins is partially redundant (Bosse and Vestweber, 1994; Labow et al., 1994; Henriques et al., 1996) and that the integrins VLA-4 and $\alpha 4\beta 7$ may also play a role in mediating the rolling of alpha4 integrin-positive cells *in vitro* and *in vivo* (Sriramarao et al., 1994; Alon et al., 1995; Kanwar et al., 1997). A role for selectins in mediating eosinophil recruitment *in vivo*

Table 3. Studies evaluating the effects of anti-cell adhesion molecules (CAMs) on eosinophil migration *in vivo*.

CAM TARGETED	STRATEGY	SPECIES	EFFECTS OBSERVED	REFERENCE
<i>Adhesion molecules present on endothelial cells:</i>				
P-selectin/ICAM-1/VCAM-1 Antibodies (Abs)	knock-outs (KO),	mice	Inhibition of rolling in P-selectin KO and inhibition of recruitment to the peritoneal cavity in P-selectin and ICAM-1 KO in the presence of anti-VCAM-1 antibodies.	Broide et al., 1998a
P-selectin/ICAM-1	KO	mice	Inhibition of eosinophil recruitment in the lung of ICAM-1 and P-selectin KO. Loss of inhibition in P-selectin KO with time.	Broide et al., 1998b
P-/E-selectin/CD11b/CD18	Abs	mice	Inhibition of eotaxin-induced eosinophil recruitment.	Das et al., 1997
P-selectin	KO	mice	Inhibition of eosinophil recruitment in the lung and inhibition of AHR	De Sanctis et al., 1997
P-/E-/L-selectins	Abs	mice	Inhibition of LPS-induced pleural eosinophilia; P- and E-selectins are functionally redundant.	Henriques et al., 1996
P/E-selectins/VLA-4	Abs	mice	Inhibition of eosinophil recruitment in skin sites; reliance on selectins depends on whether a direct-acting chemoattractant or an allergic reaction are studied.	Teixeira and Hellewell, 1998
ICAM-1	Ab	monkey	Blockade of tissue eosinophilia and AHR.	Wegner et al., 1990
ICAM-1	Ab	monkey	In animals with chronic AHR and airway eosinophilia, anti-ICAM-1 does not prevent disease but prevents recurrence after treatment with steroids.	Gundel et al., 1992
ICAM-1	KO	mice	Inhibition of airway hyperresponsiveness (AHR), lung eosinophilia.	Wolyniec et al., 1998
ICAM-1	Ab	rat	Inhibition of antigen-induced airway eosinophilia.	Chin et al., 1998
ICAM-1/ CD11/CD18	Ab	rat	Inhibition of allergen-induced nasal eosinophilia, antibodies given during immunisation.	Asakura et al., 1996
ICAM-1	Ab	rat	Inhibition of antigen-induced airway eosinophilia.	Richards et al., 1996
ICAM-1	Abs	rat	Inhibition of AHR but no effect on airway eosinophilia after antigen.	Sun et al., 1994
ICAM-2	KO	mice	Delayed eosinophil migration but prolonged accumulation, heightened AHR.	Gerwin et al., 1999
ICAM-1/VCAM-1	KO	mice	Both inhibit eotaxin-induced transmigration, but VCAM-1 appears more relevant later.	Jia et al., 1999
VCAM-1/ICAM-1	KO	mice	Inhibition of pulmonary eosinophilia.	Gonzalo et al., 1996
VCAM-1	Ab	mice	Inhibition of cutaneous eosinophilia in contact hypersensitivity.	Satoh et al., 1997
VCAM-1/VLA-4/ICAM-1	Ab	rat	Inhibition of IL-4-induced cutaneous eosinophilia, no effect of anti-ICAM-1.	Sanz et al., 1998
VCAM-1/VLA-4	Ab	rat	Inhibition of TNF α -induced cutaneous eosinophilia.	Sanz et al., 1997
<i>Adhesion molecules present on leukocytes:</i>				
L-selectin/VLA-4	Abs	guinea pig	Inhibition of eosinophil recruitment, AHR and M2 receptor dysfunction with anti-VLA-4, but not anti-L-selectin.	Fryer et al., 1997
L/P-Selectins	Fucoidin	guinea pig	The selectin-binding polysaccharide fucoidin blocks eosinophil migration by blocking both P- and E-selectin function.	Teixeira and Hellewell, 1997
CD11/CD18	Ab	guinea pig	Inhibition of eosinophil migration in skin.	Teixeira et al., 1994b
CD11/CD18	Abs	guinea pig	Inhibition of eosinophil numbers in BAL, but not tissue, inhibit AHR (one Ab).	Milne and Piper, 1994
CD11/CD18	Ab	guinea pig	Blockade of tissue eosinophilia and AHR.	Noonan et al., 1991
CD11/CD18	Abs	guinea pig	Blocks eosinophil migration in naive and primed skin sites.	Macari et al., 1996
CD11/CD18/VLA-4	Abs	guinea pig	Anti-CD18 blocks eosinophil migration in naive and primed sites. Anti-VLA-4 blocks responses induced by TNF α only.	Macari et al., 1998
CD11/CD18/VLA-4	Abs	guinea pig	Either treatment alone inhibits BAL, but not tissue, following sephadex. Combined treatment affects tissue eosinophilia.	Das et al., 1995
CD11/CD18/VLA-4	Abs	rat	Both antibodies block AHR. Airway eosinophilia is blocked by anti-CD11b, but not anti-VLA-4.	Laberge et al., 1995
CD11/CD18/VLA-4	Abs	rat	Inhibition of 5-oxo-ETE-induced lung eosinophilia by anti-CD11a and anti-VLA-4, not anti-CD11b.	Stamatiou et al., 1998
CD11/CD18/VLA-4	Abs	rat	Partial inhibition of BAL and parenchymal lung eosinophilia by either anti-CD18 or anti-VLA-4. Combined treatment abrogates eosinophilia	Schneider et al., 1999
CD11/CD18/VLA-4	Abs	mice	Inhibition of eotaxin-induced lung eosinophilia and and BHR in IL-5 transgenic mice by anti-VLA-4 but not anti-CD11b; synergism when both used together.	Hisada et al., 1999
VLA-4	Ab	guinea pig	Inhibition of lung and BAL eosinophilia, no effect on AHR.	Milne and Piper, 1995
VLA-4	Ab	guinea pig	Inhibition of airway eosinophilia and AHR after antigen challenge.	Sagara et al., 1997
VLA-4	Ab	rat	Inhibition of airway eosinophilia and AHR after antigen challenge.	Richards et al., 1996
VLA-4	Ab	guinea pig	Inhibition of airway eosinophilia, free EPO and AHR after antigen challenge.	Kranevel et al., 1997
VLA-4	Ab	guinea pig	Inhibition of nasal eosinophilia and eosinophil activation.	Terada et al., 1996
VLA-4	Ab	guinea pig	Inhibition of airway eosinophilia and AHR after antigen challenge.	Pretolani de al., 1994
VLA-4	Abs	sheep	Blockade of AHR, late phase reaction with little effect on eosinophil influx.	Abraham et al., 1994
VLA-4	Ab	guinea pig	Blockade of eosinophil recruitment in skin.	Weg et al., 1993
VLA-4/VCAM-1/ICAM-1 CD11/CD18	Abs	mice	Anti-VLA or anti-VCAM-1, but not anti-CD11b or anti-ICAM-1, block eosinophil recruitment in the trachea of antigen-challenged mice.	Nakajima et al., 1994

was initially demonstrated in a murine model of lipopolysaccharide (LPS)-induced pleural eosinophilia (Henriques et al., 1996). In this model, blocking antibodies against L-selectin or a combination of anti-P- and anti-E-selectin monoclonal antibodies (mAbs) virtually abolished LPS-induced eosinophil recruitment. In addition, several studies with knock-out mice have confirmed the importance of the selectins, specially the endothelial selectins (P and E) in mediating eosinophil rolling and recruitment *in vivo* (see Table 3). In these experiments however, it has been difficult to evaluate whether the antibodies or the knocked-out gene inhibited eosinophil recruitment directly or indirectly by modulating the recruitment and/or activation of lymphocytes and macrophages. More recently, we have shown that the combined treatment with anti-selectin antibodies abolished eosinophil recruitment induced by direct-acting chemoattractants or in allergic reactions in murine skin (Teixeira and Hellewell, 1998). The latter studies clearly demonstrate that selectin-based therapies do have a direct effect on eosinophil trafficking *in vivo*. Moreover, it was clear from these studies that eosinophil recruitment in response to direct acting mediators was mostly P-selectin-dependent, whereas that in a delayed-onset allergic reaction it was dependent on both P- and E-selectin (Teixeira and Hellewell, 1998). These findings highlight the redundant function of P- and E-selectin at mediating both neutrophil and eosinophil recruitment in models of chronic inflammation. Moreover, they suggest that blockade of both endothelial selectins is necessary if effective inhibition of leukocyte recruitment at sites of chronic inflammation is to be achieved pharmacologically. Interestingly, in a delayed-type hypersensitivity (DTH) reaction, eosinophil recruitment was independent of P-, E- or L-selectin (Teixeira and Hellewell, 1998). The inability of anti-selectin antibodies to block eosinophil recruitment in the DTH reaction suggests there are additional selectin-independent adhesion pathways which may mediate eosinophil rolling in these late phase skin reactions. Two possibilities include a role for VLA-4 in mediating eosinophil rolling in conditions of chronic inflammation mimicked by the DTH reaction or the recently described cell adhesion molecule which plays an important role in mediating bovine lymphocyte rolling *in vitro* (Jutila et al., 1997). The ability of anti-VLA-4 antibodies to block eosinophil recruitment in sites of DTH reactions support the former hypothesis, but definite proof that anti-VLA-4 blocks rolling and not firm adhesion (see below) in the model is still lacking (Teixeira and Hellewell, 1998). Nevertheless it is worth noting that studies have demonstrated that anti-VLA-4 blocked the rolling of eosinophils *in vivo* (eg. Sriramarao et al., 1994).

Studies with human eosinophils highlight differences in the ability of neutrophils and eosinophils to use differentially P- and E-selectin for rolling *in vitro*. This is in line with the idea that the selectin ligand(s) on eosinophils is different from that of neutrophils (Symon et al., 1996). If such a concept is true, it would then be

possible to devise eosinophil-selective drugs which would block their migration, but not those of neutrophils, *in vivo*. The inhibition of eosinophil migration by selectin-active drugs (eg. fucoidin) has been demonstrated *in vivo* (Teixeira and Hellewell, 1997), but no studies have proved the concept that a differential action on eosinophils versus neutrophils is possible *in vivo*.

The interaction selectin/selectin ligand approximates eosinophils to endothelial cells and allows these cells to roll on the endothelial surface. The rolling leukocyte is activated by molecules acting on 7-transmembrane receptors and this activation leads to an increase in the function of integrins on the eosinophil surface. Proof that activation of such receptors is relevant for eosinophil migration *in vivo* has been recently provided using pertussis toxin which binds to and inactivates certain G proteins coupled to 7-transmembrane receptors (Teixeira et al., 1997a). Several studies have evaluated the ability of anti-integrin antibodies on the recruitment of eosinophils *in vivo* (see Table 3). Although anti-CD18- or anti-ICAM-1-based strategies are very effective at suppressing eosinophil recruitment to a range of different tissues, these strategies also interfere with the ability of neutrophils to migrate into sites of tissue inflammation (Springer, 1994; Henriques et al., 1996). Thus, specificity would be lost and most interest is now centred on the role of VLA-4 or its ligands (especially VCAM-1) in mediating eosinophil recruitment *in vivo*.

VLA-4 integrin is constitutively expressed on the surface of eosinophils, but not neutrophils (with the exception of rat neutrophils), and appears to play an important role in the binding of eosinophils to cytokine-activated endothelial cells *in vitro* (Teixeira et al., 1995).

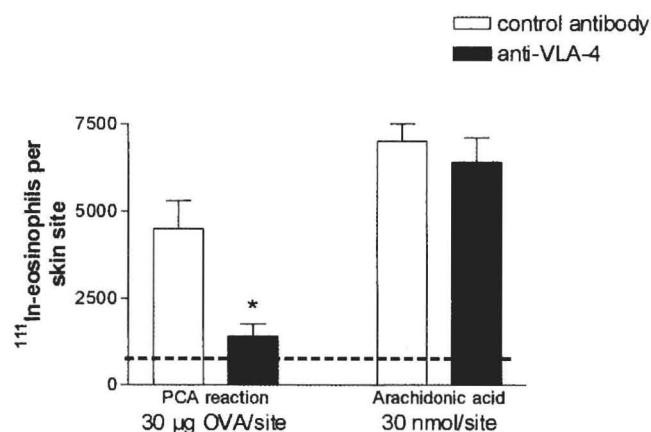


Fig. 1. Effect of an anti-VLA-4 monoclonal antibody (2B4) on the migration of eosinophils induced by arachidonic acid or in a passive cutaneous anaphylactic (PCA) reaction in guinea pig skin. Animals were injected i.v. with the monoclonal antibody (2B4, 3mg/kg) and antigen (ovalbumin, OVA) or arachidonic acid injected intradermally. ^{111}In -eosinophils (3×10^6 /animal) were then administered i.v. and their accumulation in the skin sites assessed over a 2-hour period. Results are the mean \pm s.e. mean of 5 animals. *: when $P < 0.05$ when compared to animals injected with control antibody (MOPC21).

Table 4. Examples of chemoattractant molecules known to induce eosinophil migration *in vivo*.

Lipid mediators: PAF, LTB ₄ , 5-HETE
Complement fragments: C5a, C3a
Chemokines: eotaxin, RANTES, MCP-3, eotaxin-2, MCP-4, MIP-1a, IL-8
Cytokines ¹ : IL-2, IL-5, TNF α , IL-16
Components of microorganisms: FMLP, lipopolysaccharide ¹
Others ² : histamine, ECF-a, PGD ₂

¹: usually an indirect effect via the release of chemokines or lipid mediators *in vivo* and chemokinetic *in vitro*; ²: small activity *in vitro* and very weak activity *in vivo*.

Blockade of VLA-4 is followed by suppression of eosinophil recruitment to sites of inflammation in most, but not all, studies (Table 3). Common to the studies which observed an inhibition was the prolonged time-course required for eosinophil recruitment, usually occurring after or concomitantly with other cell types (e.g. lymphocytes, monocytes, mast cells) which are themselves inhibited by anti-VLA-4 treatment. Using two different anti- α 4 mAbs we evaluated the role of α 4-dependent pathways on the rapid accumulation of ¹¹¹In-eosinophils into sites of allergic and non-allergic inflammation. We demonstrated that ¹¹¹In-eosinophil accumulation induced in a passive allergic reaction, but not after injection of arachidonic acid, was effectively inhibited by pretreatment with anti- α 4 antibodies (Fig. 1). The choice of arachidonic acid to compare to the allergic reactions was based on our previous findings that both reactions were dependent on the local release of leukotrienes (Teixeira and Hellewell, 1994). In addition, pretreatment of eosinophils with anti- α 4 mAbs prior to their injection *in vivo* was not followed by inhibition of their recruitment. These results suggest that the eosinophil may not be the only target of the inhibitory effects of systemically administered anti- α 4 mAbs, specially when rapid accumulation of these cells is studied. Other cell targets include an effect of anti- α 4 antibodies on the recruitment of monocytes, lymphocytes and, possibly, basophils, in addition to an effect on the function of mast cells (Carlos and Harlan, 1994; Springer, 1994).

A few studies have evaluated the ability of anti-CD18 and anti- α 4 to synergise at inhibiting allergen-induced eosinophil recruitment *in vivo* (see Table 3). In the mouse trachea, an anti-CD18 mAb had no effect on antigen-induced eosinophil infiltration when used alone, but significantly enhanced the inhibitory effects of an anti- α 4 mAb (Nakajima et al., 1994). In contrast, in a rat model of lung inflammation, an anti- α 4 mAb had no effect on eosinophil influx and did not increase the ability of an anti-CD18 mAb to inhibit bronchoalveolar lavage or tissue eosinophilia (Laberge et al., 1995). In our guinea pig skin model, eosinophil recruitment in response to non-allergic inflammatory stimuli was CD18-dependent and there was no further inhibition when an anti- α 4 mAb was used (Teixeira and Hellewell,

unpublished). Allergen-induced inflammation was inhibited by anti-CD18 and anti- α 4 mAbs and there was no synergy when both mAbs were used. This last piece of evidence suggests that these two adhesion pathways, CD18-dependent and α 4-dependent, are positioned in series and do not represent redundancy of the system. One possibility, at least in the guinea pig cutaneous model of inflammation, is that the α 4-dependent pathway is important for the adequate release of mediators from mast cells and the CD18 pathway is responsible for the adhesion of eosinophils to endothelial cells.

There are no reports on the ability of anti-integrin-based strategies to modulate eosinophil recruitment in human allergic diseases. One interesting observation is the presence of eosinophils in tissues of patients with leukocyte adhesion deficiency I (LAD I, lack CD18-dependent adhesion) (Springer, 1994). The latter observation suggests that alternative pathways, presumably VLA-4/VCAM-1, mediate the migration of these cells. Studies in human diseases are needed to understand the efficacy and specificity (side-effects) of the blockade of anti-VLA-4/VCAM-1 versus anti-CD18/ICAM-1 as modulators of eosinophil recruitment.

Eosinophil trafficking - chemoattractant molecules

The activation of eosinophils by chemoattractant molecules which act on 7-transmembrane receptors is a critical step in the adhesion cascade (Teixeira et al., 1997a). The activation of such receptors induces an elevation of intracellular calcium in eosinophils which precedes their migration *in vivo* (Teixeira et al., 1997a). Thus, it appears that molecules acting on serpentine receptors are capable of inducing eosinophil migration *in vivo* by directly activating the eosinophil. These molecules include lipid mediators (e.g. PAF and LTB₄), complement fragments (e.g. C5a) and chemokines (e.g. eotaxin) (see Table 4). In addition, several mediators which act on receptors distinct from the serpentine receptors, including the cytokines IL-5, TNF α and IL-2, have been shown to induce the chemokinesis of eosinophils *in vitro* and their accumulation in tissues (Weller, 1991; Milne et al., 1995; Gienbycz and Lindsay, 1999). For example, TNF α is chemokinetic for eosinophils *in vitro* and its *in vivo* administration induces the accumulation of eosinophils in the skin of guinea pigs (Macari et al., 1998 and references therein). However, TNF α appears to act indirectly via the production of a yet unidentified chemoattractant molecule (Macari et al., 1998). Similarly, the effects of IL-2 and IL-5 on the accumulation of eosinophils *in vivo* is not demonstrated in all studies (e.g. Milne et al., 1995) and appears to be indirect, possibly dependent on the release of chemokines such as eotaxin.

Recently, there has been much interest in a group of chemoattractants, the chemokines, which appear to play a major role in mediating eosinophil recruitment *in vivo*. Chemokines are a group of chemoattractant proteins

with molecular weight usually ranging from 8 to 12 kDa and amino acid sequence identity of 20 and 90% (Power and Wells, 1996). These proteins possess 4 conserved cysteine residues as part of their structure and, depending on the presence or absence of one amino acid between the first two cysteines adjacent to the N terminus, are classified in C-C or C-X-C chemokines. There are two other additions to the family – a C chemokine (lymphotactin) which possesses only two cysteine residues and a C-X₃-C chemokine (fractalkine) which possesses three intervening amino acids (see Baggiolini, 1998 for review). Chemokines are produced in both acute and chronic inflammatory reactions, but also appear to control leukocyte trafficking under basal conditions (Matthews et al., 1998). The great interest in chemokines stems from their ability to selectively activate and recruit particular leukocyte subsets. This selectivity is based on the differential expression of chemokine receptors among different leukocyte subsets (Murphy, 1996; Baggiolini, 1998). For example, human eosinophils express high levels of the chemokine

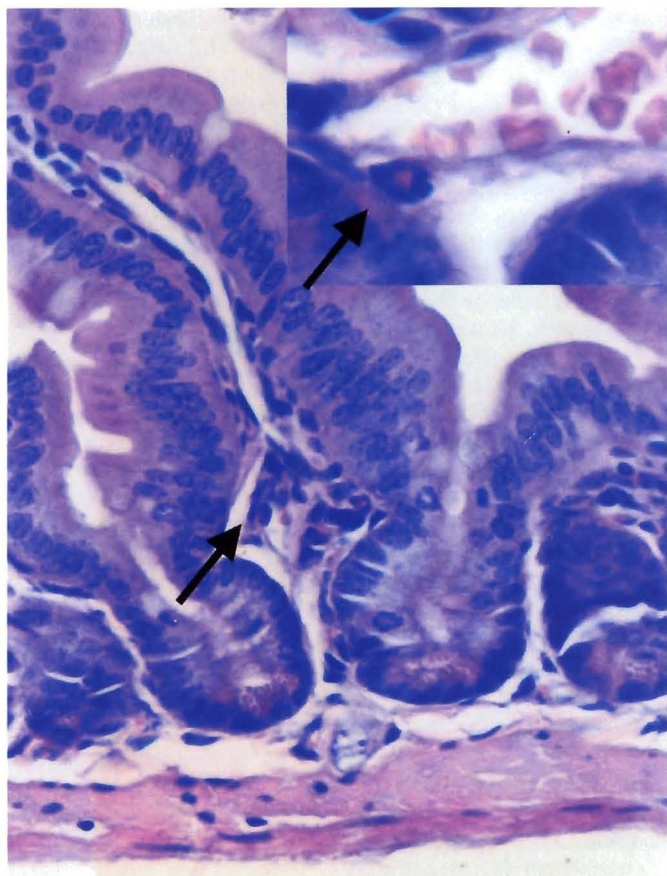


Fig. 2. Histological aspect of the intestinal mucosa of a germ-free mouse. (x 400) Note the marked decrease in the number of cells infiltrating the submucosa of these animals. A few eosinophils are also seen (arrow). In the insert, an eosinophil is seen leaving a blood vessel (x 1000)

receptor CCR3 (40,000 to 400,000 receptors/eosinophil) and low levels of the CCR1 receptor (1 to 5% of the CCR3 levels) (Daugherty et al., 1996; Ponath et al., 1996). Thus, chemokines which activate the CCR3 receptor (e.g. eotaxin, RANTES, MCP-3) are capable of activating several eosinophil functions *in vitro*. The importance of the CCR1 receptor for the activation of human eosinophils is a matter of controversy, but, at least in some allergic individuals, chemokines which activate this receptor (e.g. MIP-1 α) do activate the function of eosinophils.

In a murine model, we have recently shown that the intradermal injection of the C-C chemokines eotaxin and MIP-1 α , but not RANTES or MCP-5, induced the recruitment of eosinophils to skin sites (Teixeira et al., 1997b; Teixeira, 1998). Interestingly, an anti-eotaxin antibody, but not an anti-MIP-1 α antibody, inhibited the recruitment of eosinophils to sites of active anaphylactic reactions in mouse skin (Teixeira et al., 1997b). Together these data demonstrate the effectiveness of eotaxin on inducing eosinophil migration *in vivo* and the importance of endogenous eotaxin for the migration of eosinophils to sites of allergic reactions. This is in agreement with recent findings of defective eosinophil migration to sites of pulmonary anaphylaxis in eotaxin knock-out mice (Rothenberg et al., 1997). However, even in the knock-out animals, there was marked eosinophil migration in the later stages following antigen challenge and, in one study, no inhibition of eosinophil migration was observed (Rothenberg et al., 1997; Yang et al., 1998). These studies highlight an important aspect not to be missed when evaluating the effects of chemokines – there is much redundancy in the system and inhibition of one chemokine may be overcome by some other chemokine with a similar functional profile. In the case of eosinophils, blockade of eotaxin may be insufficient for inhibiting eosinophil recruitment since other chemokines, such as RANTES, MCP-3 and eotaxin-2, may replace eotaxin functionally. As eosinophils have one dominant chemokine receptor, one possibility to overcome such a problem would be the development of CCR3 receptor antagonists. At least one study has shown the feasibility of this approach. Pre-treatment of animals with the CCR3 receptor antagonist MetRANTES effectively inhibited the recruitment of eosinophils in anaphylactic reactions in mouse skin (Teixeira et al., 1997b).

What is the role of the eosinophil?

The functional importance of eosinophils in immunology is usually considered under pathophysiological conditions. In other words, studies which evaluate the role of eosinophils are usually carried out in disease states, such as asthma or parasitic infections. This is most likely a problem of immunology itself which tends to evaluate the role of cells, immunoglobulins or reactions in the light of defence against invading organisms, but not in the light of a normally

functioning organism. Thus, there remains an important question to be evaluated: what role do eosinophils play under normal conditions, i.e. what is the physiological role of eosinophils? As mentioned above, eosinophils are typically seen in mucosal tissues and are even found in the intestinal mucosa of germ-free animals (Fig. 2). It would be logical then to associate the physiological function of these cells with the normal function of mucosal tissues, which itself is difficult to define. As mucosal tissues are an important interface between the organism and its environment and, if we believe these are essential places for the entrance of antigens, eosinophils are at an important interface to function as antigen-presenting cells (reviewed by Weller and Lim, 1997). In this way, eosinophils would deal not only with antigens from invading microorganisms, but also, and more importantly, with the great majority of antigens we meet everyday - those which are ingested or inhaled. The understanding of this physiological role of eosinophils is far from clear but it may shed light on the understanding of the role these cells play in human disease. Below, we will consider the role eosinophils play in helminthic infections and allergic diseases.

Eosinophils and helminth infections

Helminth infections are highly prevalent in human populations, particularly in tropical and subtropical countries. Twenty-six species of helminth parasites have been reported to infect humans. The four most prevalent species of nematodes - *Ascaris lumbricoides*, *Necator americanus*, *Ancylostoma duodenale* and *Trichuris trichiura* - infect almost three billion people worldwide (Warren et al., 1990; Bundy, 1994).

The three hallmarks of gastrointestinal (GI) infection are eosinophilia, intestinal mastocytosis and IgE production (Love et al., 1976; Ruitenberget al., 1979; Jarrett and Miller, 1982). These three elements are also associated with the immune response during allergic disease, like asthma (see below). The IgE production, eosinophilia and mastocytosis are regulated by cytokines produced predominantly by a T helper cell subset in human and mice designated Th-2 (Mossmann and Coffman, 1989; Romagnani, 1994). The Th-2 subset produces predominantly IL-4, IL-5, IL-9, IL-10 and IL-13; these cytokines regulate the humoral response, promoting B cell proliferation and immunoglobulin switching to non-complement-fixing IgG isotypes, IgG1 in mice or IgG4 in human, and to IgE-producing plasma cells (reviewed by Abbas et al., 1996; Mossmann and Sad, 1996). Moreover, Th-2 cytokines, especially IL-5, induce eosinophil proliferation, differentiation and survival (see above). Although the CD4+ T cell separation into Th-1 and Th-2 has been very well demonstrated *in vitro* and associated, *in vivo*, with the outcome of some infections, such as *Leishmania major* infection in mice, it appears that such division is an oversimplification of the immune response. For example, the measurement of cytokines in the thoracic

duct lymph of *Trichinella spiralis*-infected rats demonstrated that, even though the intestinal immune response has a predominant Th-2 profile with high levels of IL-5 and IL-4, IFN- γ was also present (Ramaswamy et al., 1996).

Recent evidence favours the idea that a protective role exists for Th2-like responses in helminthic infection (Urban et al., 1992). There are at least two independent studies of murine infection with *Trichuris muris* (Else et al., 1992) and *Heligmosomoides polygyrus* (Urban et al., 1991) that have directly demonstrated a protective role of the Th2 response. In *T. muris* infection, the mouse strains that predominantly produce Th-1 responses develop a chronic infection and are defined as susceptible strains for this parasite. The mouse strains capable of mounting a Th2 response against *T. muris* infection are able to eliminate worms before they mature into egg-producing adult worms, and are designated resistant strains. Susceptible mouse strains are able to cure the chronic infection produced by *T. muris* if they are treated with IL-4 (Else et al., 1994).

The mechanism by which IL-4 acts to control a helminth infection is still unknown. IL-4 is essential for the induction of IgE responses (Finkelman et al., 1988) and, together with the IL-4-induced Th2 response, is also necessary for the intestinal mastocytosis, at least in mice (Madden et al., 1991). Also, IL-4 production is normally associated with increase in IL-5 and, consequently, eosinophilia. Therefore, any or all of these three hallmarks of helminthic infection - eosinophilia, IgE and mastocytosis - might participate in the protection. We will review and discuss the association of IgE, mast cell and eosinophils with helminthic infections and the possible role of these immune mechanisms in protective immunity. However, it is important to underline that the importance of the IL-4-dependent response observed during helminthic infection is not fully understood. Urban et al. (1995) have recently suggested that IL-4 may have a direct effect on the parasite and/or the intestinal mucosa that might contribute to worm elimination. *In vitro*, prolonged stimulation with IL-4 stimulation produced a dose-dependent proliferation of a rat intestinal epithelial cell line - IEC-6 (McGee and Vitkus, 1996). Colgan et al. (1994) also showed that IL-4 stimulation of the T84 cell line (a human gut epithelium cell line) resulted in a modulation of the barrier function and ion transport displayed by the monolayer. In the same system, IL-4 regulated the expression of accessory molecules that mediate neutrophil adhesion on the monolayer. Finally, Ramaswamy et al. (1994) reported an IgE transport mechanism in the intestine of *T. spiralis*-infected rats, which is also IL-4 dependent. Thus, IL-4 may have a host protective role against intestinal helminths by a mechanism not entirely dependent on its ability to modulate IgE or mast cell activities.

The participation of IgE in protective immunity against helminthic infections is still unclear. However, several studies have reported a correlation between

specific IgE levels and protection against infection with different species of helminths - *Taenia taeniformis* (Musoke et al., 1978), *Trichinella spiralis* (Dessein et al., 1981; Negrão-Correa et al., 1999), *Brugia malayi* (Kurniawan et al., 1993), *Strongyloides ratti* (Korenaga et al., 1986), *Necator americanus* (Pritchard et al., 1995) and *Schistosoma* spp (Dunne et al., 1992; Hagan, 1993). A more direct evidence for IgE involvement in protective immunity has been achieved by transfer of parasite-specific IgE in two infection models in rats. In *S. mansoni* infection, a purified IgE-monoclonal antibody specific for adult worm metabolic antigen passively transferred a significant level of protection to adoptive recipients (Verwaerde et al., 1987). In *T. spiralis*-infected mice the transfer of immune IgE to intestinal primed rats will lead to rapid expulsion of the parasite (Ahmad et al., 1991). Moreover, IgE immunoprecipitated from 14 day-*T. spiralis*-infected rats was able to abolish the ability of adult worm to invade IEC-6 (rat epithelial cell line) monolayer (Negrão-Corrêa, 1997).

Due to the low concentration of IgE in serum, its function is normally associated with the receptor(s) to which IgE binds on cells. Therefore, two mechanisms are postulated to explain the protective role of IgE against helminthic infection: immediate hypersensitivity and antibody-dependent cellular cytotoxicity (ADCC) reaction. The high affinity receptor for IgE, FcεRI, is expressed in high density by basophils and mast cells. Immediate hypersensitivity, the principal known function of IgE *in vivo*, requires binding of IgE to FcεRI receptors on mast cells or basophils. The cross-linking of IgE on these cells by antigen triggers the release of preformed mediators, such as histamine, heparin, proteases and cytokines and the synthesis and secretion of newly formed lipid-derived mediators and cytokines (Schwartz, 1994). The release and action of these mediators induce the characteristic symptoms observed in an allergic reaction and contribute to the chronic inflammation that can appear in the surrounding tissue. The FcεRI receptor has also been detected on dendritic cells (Grabbe et al., 1993) and on human eosinophils purified from hypereosinophilic patients (Gounni et al., 1994). The function of IgE on either of these cell types is still unknown. In human eosinophils, IgE binding to FcεRI can lead to an ADCC reaction which has been associated with killing of schistosomula of *Schistosoma mansoni* *in vitro* (Gounni et al., 1994). However, murine eosinophils do not express the high (or low) affinity IgE receptor (de Andres et al., 1997).

IgE also has a low affinity receptor, FcεRII (CD23), present on a variety of cell types, including platelets, lymphocytes, enterocytes and eosinophils (Capron and Joseph, 1991). IgE also binds to epsilon-binding proteins (εBP) detected on the surface of enterocytes (Brassart et al., 1992), eosinophils and neutrophils (Truong et al., 1993a,b), mast cells and macrophages (Frigeri and Liu, 1992). Although binding of IgE to these cells and triggering with specific antigen may mediate ADCC

killing of parasite larvae or other cellular functions *in vitro*, IgE-dependent cell functions *in vivo* for most of these cell types have not been unequivocally established.

The participation of mast cells in the protective mechanisms against GI infection was initially suggested based on the temporal coincidence of intestinal mastocytosis and worm expulsion. The theoretical mechanism, known as "Allergic Inflammation" (Larsh and Race, 1975), suggested that the local inflammatory reaction stimulated by mast cell degranulation led to worm elimination by non-specific mechanisms. The release of mast cell mediators in the intestinal mucosa may induce changes in mucosal permeability (Murray, 1972), plasma leakage and further infiltration of leukocytes (Askenase, 1979). Large amounts of histamine could also increase the production and release of mucus from goblet cells (Lake et al., 1980) and increase intestinal motility (Goldhill et al., 1997). All these alterations were thought to make the intestinal mucosa an unsuitable environment for the parasite. The idea was reinforced with the demonstration that *T. spiralis* worms transplanted into a normal rat intestine were able to survive for many days while the worms transplanted into inflamed intestine were eliminated rapidly (Wakelin and Wilson, 1979). Indication of a protective role for mast cell against intestinal helminths has also been demonstrated in mast cell-deficient mice (W/W^v mice). W/W^v mice showed a more severe *T. spiralis* (Alizadeh and Murrell, 1984), *S. ratti* (Nawa et al., 1985) and *Hymenolepis nana* (Watanabe et al., 1994) infection, with a slower worm expulsion than control mice. The direct connection between the response in W/W^v mice with mast cell function exclusively should be considered more carefully. The mutation observed in W/W^v mice affects a tyrosine kinase (c-kit) receptor for stem cell factor (SCF) that is also involved in the growth and differentiation of erythroid and granulocyte precursors, as well as lymphoid stem cells (Witte, 1990).

Mast cell degranulation cannot entirely explain the participation of specific IgE in protective mechanisms operating against all helminthic infections, since the course of *N. brasiliensis* infection in W/W^v mice was unchanged (Uber et al., 1980). Moreover, in *T. spiralis* infection, the transfer of immune IgE plus immune CD4+ OX22- T cells resulted in rapid expulsion of the infective larvae from the rat intestine (Ahmad et al 1991). However the protective T cell population (CD4+ OX22- T cells) induced an intestinal eosinocytosis, but not an intestinal mastocytosis (Wang et al., 1990). Furthermore, during worm elimination, our preliminary work using an *in vitro* system indicated that intestinal-specific IgE might participate directly in protective mechanism independently of mast cells (Negrão-Corrêa, 1997). Thus, although mast cells may play a role in protection against some helminthic infections and mast cell-derived products affect parasite survival in some systems, direct evidence for a role for mast cells has not been convincingly obtained for all the helminthic models *in vivo*.

Table 5. The effects of IL-5 on helminthic infections: IL-5 neutralization by treatment with anti-IL-5 or anti IL-5 receptor (IL-5R) antibodies.

PARASITE	EFFECT	REFERENCE
<i>Schistosoma mansoni</i>	After vaccination with irradiated cercariae, anti IL-5-treated and untreated mice eliminated similarly the challenge infection.	Sher et al. 1990
<i>Schistosoma japonicum</i>	Anti IL-5-treated and untreated mice showed similar egg elimination, although the granuloma volume was reduced in anti IL-5-treated mice.	Cheever et al., 1991
<i>Trichinella spiralis</i>	Anti IL-5-treated and untreated mice had similar muscle larvae burden after the primary and the secondary infection.	Herdon and Kayes, 1992
<i>Strongyloides venezuelensis</i>	During the primary infection, the anti IL-5 plus anti IL-5R treatment did not alter the egg production or adult worm elimination time. However, the number of adult worms in intestine was significantly higher in challenged infected IL-5-depleted mice.	Korenaga et al., 1991
<i>Strongyloides venezuelensis</i>	During primary infection, anti IL-5-treated mice had significantly higher number of intestinal adult worms than untreated mice	Korenaga et al., 1994
<i>Angiostrongylus cantonensis</i>	Anti IL-5-treated mice showed more intracranial worms than untreated mice.	Sasaki et al., 1993
<i>Onchocerca volvulus</i>	Vaccination with irradiated L3 larvae was capable of killing larvae implanted in untreated mice but not anti IL-5- or anti IL-4-treated mice	Lange et al., 1994

In the same way, the role of eosinophils in protective mechanisms against helminth have been very contradictory. As mentioned above, in most helminth-infected hosts, like *S. mansoni*-infected mice (Sher et al., 1990), *Protostrongylus rufescens* and *Haemonchus contortus*-infected lambs (Mansfield and Gamble, 1995), *Paragonimus westermani* and lymphatic filariasis in humans (Kan et al., 1995; Magnussen et al., 1995) *Trichinella spiralis*- and *Nippostrongylus brasiliensis*-infected rats (Watanabe et al., 1988) and in *Strongyloides ratti*-infected rats (Moqbel, 1980), the number of eosinophils in peripheral blood and tissues involved in parasite migration increases dramatically. The association of eosinophil and protective mechanisms against helminthic infections was proposed after the demonstration of the effectiveness of the depletion of eosinophils by anti-eosinophil antiserum in modulating infection *in vivo*. Thus, in *Trichostrongylus colubriformis*-infected guinea pigs (Gleich et al., 1979), anti-eosinophil antibody treatment was able to increase the host susceptibility to the parasite, and, in *T. spiralis*-infected mice treated with anti-eosinophil antibodies (Grove et al., 1977), the number of muscle larvae recovered was twice as much as in the control infected mice. At the same time as these *in vivo* studies were published, some authors also demonstrated that immune serum plus rat or human eosinophils were capable of mediating killing of *Schistosoma mansoni* schistosomula and *Trichinella spiralis* larvae (Capron et al., 1981; Butterworth, 1984) or *S. hematobium* schistosomula (Hagan et al., 1985) *in vitro*. The cytotoxic effect of eosinophils on antibody-coated helminth larvae has also been demonstrated *in vivo* on *T. spiralis* newborn larvae of infected rats (Wang and Bell, 1992) and on *Onchocerca volvulus* microfilaria *in vitro* (Folkard et al., 1996). Eosinophils were the main effector cells involved in the killing of *S. stercoralis* L3 larvae implanted, inside a chamber, into the peritoneal cavity of immune mice (Abraham et al., 1995). The

mechanisms underlying the ability of eosinophils to kill parasite larvae is not entirely understood, but is most likely related to the ability of eosinophils to secrete various cytotoxic proteins and mediators (see Tables 1 and 2), specially the toxic activity of cationic proteins and oxygen-derived radicals.

The regulation of eosinophil production and differentiation in the bone marrow, its migration to the tissue and survival in tissue are effectively and selectively influenced by IL-5 (see above). In this sense, strategies which block the production and/or function of IL-5 are specially important for the understanding of eosinophil function *in vivo*. Pioneering studies by Coffman et al. (1989) showed that the injection of monoclonal antibody to IL-5 suppressed the blood eosinophilia and tissue eosinophil infiltration in mice infected with *Nippostrongylus brasiliensis*. Therefore, depletion of IL-5 by specific antibody treatment or genetic manipulation of the IL-5 gene-disruption of IL-5 in IL-5-deficient mice (Kopf et al., 1996) and increased IL-5 expression in IL-5 transgenic mice (Dent et al., 1990) have been used as models to study the role of eosinophils in the protective mechanisms against helminthic infections. In Tables 5 to 7, we summarize the results obtained with different helminthic infections in these IL-5 manipulated hosts. The most obvious conclusion when analysing data in these tables is that, although all the helminthic infection stimulated an IL-5-dependent eosinophilia, IL-5 manipulation did not produce a similar effect on infection outcome of all the species evaluated. Evidence for a protective role of IL-5 has been observed mainly on Nematodes with a pulmonary phase in the life cycle (Tables 5 to 7). As discussed above, diversity in the protective mechanism against different species of helminths has also been observed when evaluating the role of mast cells, goblet cells and IgE. Thus, IL-3 treatment of nude mice restored mucosal mastocytosis and the capacity for elimination of *S. ratti* infections; however, this treatment

Table 6. The effects of IL-5 on helminthic infections: IL-5 transgenic mice.

PARASITE	EFFECT	REFERENCE
<i>Mesocestoides corti</i>	The infection was fatal in IL-5 transgenic and non-transgenic mice.	Strath et al., 1992
<i>Schistosoma mansoni</i>	After vaccination with irradiated cercariae, IL-5 transgenic mice and non-transgenic littermates showed similar resistance to challenge infection.	Freeman Jr et al., 1995
<i>Schistosoma mansoni</i>	After vaccination with irradiated cercariae, IL-5 transgenic mice carried more adult worms of challenge infection than the littermate control.	Dent et al., 1997
<i>Trichinella spiralis</i>	Infection in IL-5 transgenic mice or in the littermate controls showed no difference in the muscle larvae and intestinal adult worm recovery, or in the female fecundity.	Hokibara et al., 1997
<i>Trichinella spiralis</i>	Infection in IL-5 transgenic (low transgenic Tg1) mice resulted in more muscle larvae than in infected non-transgenic control.	Dent et al., 1997
<i>Toxocara canis</i>	After ES antigen immunization, the challenge infection in IL-5 transgenic mouse resulted in similar number of larvae than in non-transgenic controls	Sugane et al., 1996
<i>Nippostrongylus brasiliensis</i> or <i>Toxocara canis</i>	The number of tissue worms was similar in <i>T. canis</i> infected IL-5 transgenic and non-transgenic littermates. IL-5 transgenic mice had less intestinal <i>N. brasiliensis</i> and the females were less fecund than the controls.	Dent et al., 1999
<i>Nippostrongylus brasiliensis</i>	During primary infection, IL-5 transgenic mice showed lower number of lung and intestine worms than the littermate control. Moreover eosinophil transference from the transgenic to naive mice resulted in lower larvae burden in lungs.	Shin et al., 1997
<i>Angiostrongylus cantonensis</i>	IL-5 transgenic mice showed lower numbers of intracranial worms than non-transgenic littermates. The female worms recovered from IL-5 transgenic mice were smaller than the controls.	Sugaya et al., 1997

was not effective against *N. brasiliensis* infection (Abe et al., 1992; Horii et al., 1993; Ishikawa et al., 1994). Goblet cell hyperplasia, and more precisely, alteration of the terminal sugars of goblet cell mucins during *N. brasiliensis* infection is thought to be a key event leading to the elimination of adult worms (Ishikawa et al., 1993). Recently, we described (Negrão-Corrêa et al., 1996) a powerful intestinal IgE response that was initiated during a *T. spiralis* infection of rats. However, the intestinal IgE response of rats was not a universal consequence of a nematode GI infection. In rats infected with *T. spiralis* and *H. polygyrus*, IgE could be measured in the lumen of the small intestine; however, IgE was not detected in the intestinal lumen of *N. brasiliensis*-infected rats (Negrão-Corrêa et al., 1999). All these results reinforce the idea that, even though the immune response to different nematodes seems similar, the mechanism responsible for worm elimination may be selective and specific for each (or some) nematode species.

Interestingly, although the IL-5 manipulation work has been performed in mice (Tables 5 to 7), most of the *in vitro* protection and killing assays routinely use human or rat eosinophils. In this respect, it is worth remembering that De Andres et al. (1997) were not able to demonstrate expression of IgE receptors on mice eosinophils. The lack of IgE receptors in mice eosinophils should affect the ADCC killing of the parasite, suggesting that mice may not be a good model to evaluate the importance of eosinophils in helminthic infections. Another important consideration is that IL-5-deficient mice have proportionally less B1 cells; a B cell population that expresses CD5 antigen and localizes in peritoneal cavity and intestinal mucosa (Murakami and

Honjo, 1995; Kopf et al., 1996). A possible role of these cells in protection against helminthic parasites has not been evaluated.

Overall, these infection models do suggest that eosinophils might not be essential for the protective immune response against many helminths in mice. However, little data are available in other species and, specially, in humans. These are essential experiments to be carried out in order to evaluate the impact of eosinophil-based treatment on helminthic infectivity.

Eosinophils and allergic diseases

There is much circumstantial evidence to suggest a role for Th2 lymphocytes and eosinophils in the pathophysiology of allergic diseases, such as rhinitis (Klementsson, 1992), dermatitis (Bruijnzeel-Koomen et al., 1992; Grewe et al., 1998) and conjunctivitis (Foster et al., 1991). In these conditions, eosinophil numbers or eosinophil-derived mediators are elevated in tissue and appear to correlate positively with symptoms and/or loss of organ function. However, the case for a role of eosinophils in the pathophysiology of asthma is much stronger and a considerable amount of work has been published on the subject. Asthma is a common disease worldwide and its prevalence appears to be increasing, specially in developed and urban populations (Lebowitz and Spinaci, 1993). Furthermore, asthma sufferers possess a significant degree of disability, and loss of working-days can lead to important economic losses.

Initial evidence of a possible role for eosinophils in the pathophysiology of asthma derived from studies in peripheral blood demonstrating that eosinophils were primed, i.e. were hypodense or secreted more lipid

Table 7. The effects of IL-5 on helminthic infections: IL-5 or IL-5 receptor alpha chain (IL-5R) deficient mice.

PARASITE	EFFECT	REFERENCE
<i>Hymenolepis diminuta</i>	Infection in IL5 (-/-) mice did not prolong the worm survival or improve the parasite growth.	Ovington and Behn, 1997
<i>Mesocestoides corti</i>	IL5 (-/-)-deficient mice did not alter the infection outcome.	Kopf et al., 1996
<i>Brugia malayi</i>	After immunization with killed microfilariae, the challenge infection in IL5 (-/-) did not produce lung pathology or airway hyperresponsiveness (AHR) to cholinergic agonists as non-deficient mice did. The infection level was not measured by the authors.	Hall et al., 1998
<i>Toxocara canis</i>	Lung pathology due the infection was less intense in IL5 (-/-) mouse, however the larvae number and the location did not alter in IL5 (-/-) compared to the non-deficient control.	Takamoto et al., 1997
<i>Trichinella spiralis</i>	The IL-5 (-/-) mice showed higher worm burden after the challenge infection than the non-deficient littermates.	Revised by Matthaei et al., 1997
<i>Strongyloides ratti</i>	During the primary infection, IL5 (-/-) mice had more intestinal worms and eliminated more eggs/larvae than the non-deficient mice, however the adult elimination timing was similar, and both animals were resistant to secondary infection.	Ovington et al., 1998
<i>Angiostrongylus cantonensis</i>	IL5 R (-/-) mice showed an enhanced survival of worms than the controls.	Yoshida et al., 1996
<i>Angiostrongylus cantonensis</i>	IL5 R (-/-) mice yielded a higher number of worms at 20 dpi than the non-deficient littermates.	Sugaya et al., 1997
<i>Heligmosomoides polygyrus</i>	IL 5 (-/-) mice showed higher parasite burden than the controls.	Revised by Matthaei et al., 1997

mediators or enzymes when activated (Smith, 1992). More recently, studies have used invasive techniques to evaluate the lung (e.g. bronchoscopy) and these techniques have been instrumental in the understanding of the pathological and immunological alterations present in the asthmatic lung (Djukanovic et al., 1990). Thus, bronchoscopic studies assessing bronchoalveolar lavage and biopsies of the lungs of asthmatic individuals under basal conditions and following antigen challenge have demonstrated that: (i) airway inflammation is central to the pathophysiology of asthma and appears to play an important role in the development of airway hyperresponsiveness to non-specific stimuli causing bronchoconstriction (reviewed by Djukanovic et al., 1990; Bousquet et al., 1992); (ii) the infiltration of eosinophils in the mucosa, a characteristic feature of the inflammation in the asthmatic lung, is present even in patients with newly diagnosed asthma (e.g. Laitinen et al., 1993); (iii) the numbers and/or activation state of eosinophils or the levels of eosinophil-derived secretory products correlate positively with bronchial hyperresponsiveness (e.g. Wardlaw et al., 1988; Djukanovic et al., 1990); (iv) there is an increase in the levels of eosinophils following antigen challenge and this appears to correlate positively with the ensuing late phase response (e.g. Dupuis et al., 1992; O'Byrne et al., 1987); (v) the eosinophilic lung inflammation is maintained by cytokines predominantly of the TH2 phenotype (Robinson et al., 1992); (vi) these TH2 cytokines are secreted mostly by activated CD4+ T cells (Ying et al., 1993; Walker et al., 1994), but also by mast cells (Bradding et al., 1994); and (vii) treatment with steroids effectively inhibits lung inflammation and that this appears to be the mechanism underlying the effects of these drugs in asthma (Barnes, 1998a). Thus, the current

hypothesis is that asthma is a T lymphocyte-driven inflammatory disease of the airways where an increased number of activated eosinophils appear to act as major effector cells. Very importantly, although the clinical studies demonstrate the correlation between eosinophil numbers or activation state with asthma severity, no studies have proved that the correlation is indeed causal. The lack of proof derives from the lack of pharmacological strategies to inhibit specifically the function and/or activation state of eosinophils in human diseases.

Overall, animal studies demonstrate that there is a good correlation between the development of airway hyperresponsiveness (AHR) following antigen challenge and influx of activated eosinophils in the lung (Teixeira et al., 1995). In agreement with this finding, specific blockade of eosinophil migration with antibodies against IL-5 blocks the development of AHR in several species (Reviewed by Teixeira et al., 1995). For example, pretreatment of guinea pigs with an anti-IL-5 antibody (TRFK-5) abrogated both the recruitment of eosinophils and the development of AHR following antigen challenge (eg. Chand et al., 1992; Mauser et al., 1993). Similar results have been obtained in rats, mice and primates (Kung et al., 1995; Mauser et al., 1995; Egan et al., 1997; Yagi et al., 1997). Significantly, pulmonary eosinophilia, AHR and lung damage are abolished in sensitised and challenged IL-5-deficient mice (Foster et al., 1996). However, reconstitution of IL-5 with recombinant vaccinia viruses completely restored aeroallergen-induced eosinophilia and airway dysfunction (Foster et al., 1996). As seen in Table 3, inhibition of eosinophil migration with anti-CAMs is also commonly associated with inhibition of AHR.

One alternative approach to evaluate the role of

eosinophils in the development of AHR is to induce an accumulation of eosinophils in the lung with various strategies and then evaluate lung function. For example, we have recently shown that the intratracheal injection of the chemokine eotaxin into the lungs of IL-5-transgenic mice is associated with a significant increase in eosinophil numbers in the airways, free EPO levels in BAL fluid and AHR (Hisada et al., 1999). However, there are several studies which demonstrate that the simple presence of eosinophils in the lung is not sufficient to induce AHR, these cells must be activated (Pretolani et al., 1994). In this regard, Lefort et al. (1996) have recently shown that an antibody against MBP effectively blocked the AHR following antigen challenge in guinea pigs.

Despite a great deal of work demonstrating the importance of airway eosinophilia to the development of AHR, there are several studies which convincingly dissociate airway eosinophilia and AHR (reviewed by Smith, 1992; Morley, 1993). For example, the pretreatment of guinea pigs with anti-VLA-4 antibody reduced airway eosinophilia but had no effect on AHR in one study (Milne and Piper, 1995). Similarly, there was no correlation between the presence of airway eosinophilia and AHR in different strains of ovalbumin-sensitised and challenged mice (Wilder et al., 1999). In addition, it is also important to reiterate that animal models of allergic asthma only mirror part of the pathophysiology of asthma and not the disease as seen in man. For example, the changes in AHR seen in sensitised/challenged animals when compared to naive animals is orders of magnitude smaller than those observed in asthmatic versus normal individuals (Smith, 1992). So, what can we learn from animal models and how can we extrapolate it to the human diseases? Possibly, the most important message from studies in animal models is that eosinophils do appear to play an important role in the pathophysiology of AHR, but there are certainly other factors which contribute to the modification of pulmonary function. Moreover, it appears that specific inhibition of eosinophil recruitment and/or function is an important strategy to be pursued when developing new anti-asthma drugs (Teixeira et al., 1995). However, definite proof of a role of eosinophils in the pathophysiology of human asthma is still to be shown.

Pharmacological modulation of eosinophil trafficking

Thus, if eosinophils are leukocytes with an important effector function in allergic diseases, it is possible that drugs which modulate eosinophil recruitment and/or activation may be useful therapeutic agents for these diseases. In addition, novel therapeutic strategies may arise from the understanding of the mechanisms underlying eosinophil recruitment and/or activation. The central point in this working hypothesis is the central role of eosinophils in the pathophysiology of allergic conditions, such as asthma. As mentioned above, the

evidence towards such an unique role is very strong but definite proof is still lacking. Thus, it is not known whether specific inhibition of eosinophil recruitment and/or action will result in improvement of asthma. In addition, one central assumption in the use of anti-eosinophil-based strategies is that these will be used in patients (from developed countries) in whom the risk of helminthic infection is low.

Glucocorticosteroids are the "gold standard" for the treatment of eosinophilic conditions in man and against which novel drugs and therapies must be compared. Indeed, steroids are probably the most effective inhibitors of the recruitment of eosinophils in allergic models of inflammation in animals and possibly the only class of drugs which have been successfully used against eosinophilic diseases in man (Teixeira et al., 1995; Barnes, 1998a). At the molecular level, steroids bind to specific receptors in the cytoplasm and that leads to the translocation of the complex to the nucleus of the cell where it will inhibit or induce the synthesis of a great range of proteins (Barnes, 1998b). For example, steroids block the synthesis of the Th2 cytokines IL-4 and IL-5 and this inhibition may account for some of the inhibitory effects of steroids on eosinophil recruitment observed *in vivo*. Steroids may also induce the synthesis of a group of proteins named lipocortins which have been shown to modulate the recruitment of leukocytes (Das et al., 1997 and references therein). Nevertheless, in one study, blockade of lipocortin had no effect on the inhibitory actions of dexamethasone in mice (Teixeira et al., 1998). Finally, a direct effect of steroids on eosinophils appears to play a role in the modulation of the recruitment of these cells *in vivo* (Teixeira and Hellewel, 1996). Despite their efficacy, the use of high doses of steroids is accompanied by a series of important side effects, such as metabolic disturbances and decreased resistance to infections. Thus, a new "anti-eosinophil" drug should ideally be as effective as or more effective than the steroids but possess fewer side-effects. So far, no such drug exists but there are a few potential candidates. The possible use of anti-CAMs as therapeutic agents is discussed above. Although most experiments to date have used antibodies, there are several companies developing small molecules with anti-CAM activity which could be administered orally. The availability of these molecules for testing in different animal models and clinical trials is eagerly awaited. Nevertheless, inasmuch as strategies which block the function of CAMs do not appear to be eosinophil-specific, a range of side effects related to inhibition of other leukocytes (e.g. increased susceptibility to infection) may also occur in the presence of such drugs.

The development of drugs which block the action of chemoattractant mediators is probably the strategy that has deserved the greatest attention throughout the years (Teixeira et al., 1995). The availability of a defined receptor with a defined binding molecule facilitates the development of antagonists, specially with molecular biology techniques and robotics that allow large-scale

screening. There are good antagonists that block PAF and leukotriene B₄ receptors and there has been much recent interest in the development of chemokine receptor antagonists, specially antagonists for the CCR3 receptor. Although this strategy achieves a great degree of specificity and, presumably, fewer side-effects, there is much concern about the efficacy of using antagonists at specific receptors. In both acute and chronic inflammatory reactions, there is much mediator redundancy. In other words, there could be several eosinophil chemoattractants released at the site of inflammation and the inhibition of only one may not be sufficient for inhibition of eosinophil recruitment. Nevertheless, these questions are only theoretical and clinical trials must definitely be carried out to refute or confirm the effectiveness of any mediator receptor antagonist. To use an example from another system: although there are many factors which control blood pressure, treatment with angiotensin receptor antagonists is an effective means of treating hypertension. A few inflammatory mediator receptor antagonists have been tested in clinical trials of human asthma. Although preclinical studies, specially in guinea pigs, suggested an important role for PAF receptor antagonists in the treatment of asthma, most clinical studies have failed to demonstrate any significant effect of PAF receptor antagonists in clinical asthma (see for example Kuitert et al., 1995). In contrast, leukotriene receptor antagonists (e.g. zafirlukast and montelukast) do provide therapeutic benefit in clinical asthma when given alone or in addition to inhaled steroids (Busse et al., 1999; Kemp et al. 1999; Malmstrom et al., 1999). However, leukotriene receptor antagonists were no better than currently used long-acting β_2 -adrenoceptor agonists (salmeterol, Busse et al., 1999) or inhaled steroids (budesonide, Malmstrom et al., 1999) for asthma control.

One other alternative to controlling eosinophil influx is via the use of drugs, called functional antagonists, which block the recruitment of these cells whatever the stimuli or mediators involved. The glucocorticosteroids are an example of functional antagonists. More recently, much interest has been placed on drugs which elevate cyclic AMP as modulators of eosinophil recruitment *in vivo* (Teixeira et al., 1997c). It is hoped that these drugs will be as effective as steroids but with fewer side-effects or, at least, side-effects which would be easily reversed upon discontinuation of the drug. The levels of cyclic AMP in a given leukocyte are controlled by a balance between the rate of cyclic AMP production via G protein-coupled adenylate cyclase and the rate of cyclic AMP degradation by phosphodiesterases (PDEs) (Teixeira et al., 1997c). Although agents which elevate cyclic AMP by activating adenylate cyclase possess the ability to block eosinophil migration and/or activation acutely, it appears that most effects of these drugs are lost chronically (Giembycz, 1996). The reasons for such a loss of anti-inflammatory effects is not entirely understood but could be related to the ability of elevated levels of cyclic AMP to induce PDEs in different cell

types (Teixeira et al., 1997c; Giembycz and Lindsay, 1999).

The PDE family of enzymes is subdivided into 7 subfamilies based on their biochemical, pharmacological and genetic characteristics. One interesting aspect of the biology of PDEs is that a subfamily of enzymes, namely PDE4, appears to be the major modulator of cyclic AMP levels in cells which participate in the inflammatory process (reviewed in Teixeira et al., 1997c; Giembycz and Lindsay, 1999). As such, PDE4 inhibitors are effective inhibitors not only of the functional response of eosinophils but also of their ability to migrate into sites of inflammation *in vivo* (Teixeira et al., 1994a, 1997d; Giembycz and Lindsay, 1999). In addition to their anti-inflammatory activity, PDE4 inhibitors also possess some bronchodilatory activity which may have an additional usefulness in the treatment of asthma (Giembycz and Dent, 1992). Moreover, PDE4 inhibitors do not appear to lose their anti-inflammatory efficacy after continuous administration *in vivo*. Although no PDE4 inhibitor has been tested successfully in the clinic, clinical usefulness derived from the treatment of asthmatic patients with the non-specific PDE inhibitor theophylline suggests this to be an interesting and promising approach.

The central role of IL-5 in eosinophilopoiesis, maturation, priming and survival in tissue (see above) makes the inhibition of this cytokine one of the most powerful targets to inhibit eosinophil recruitment and activation *in vivo*. However, the development of drugs which bind to and block the IL-5 receptor has proven very difficult and no inhibitor has been reported to date. Nevertheless, there are several antibodies which bind to and inactivate IL-5 function (see above). Results from clinical trials evaluating the use of these antibodies in human disease are eagerly awaited and should strongly make a case for (or against) the efficacy and safety of eosinophil-based therapeutic strategies. Whether the use of antibodies for diseases is feasible in chronic conditions such as asthma is unlikely, but a few studies in animals demonstrate that anti-IL-5 antibodies are effective even 3 to 6 months after their administration (Egan et al., 1997).

Concluding remarks

Much is now known about the mechanisms underlying eosinophil recruitment *in vivo* and the ability of these cells to release several toxic substances in tissues. In addition, much data have been gathered to suggest an important role for eosinophils in the physiopathology of allergic diseases, such as asthma. In the next few years, we will be able to determine whether this suggestion is correct and whether eosinophil-based therapeutic strategies are indeed effective for the treatment of allergic diseases. However, if these strategies are effective and patients are treated in such a way, we will need to very carefully examine their effects on helminthic diseases, specially in developing countries where they are more prevalent. Meanwhile, much research is needed to understand the physiology of eosinophils in the absence of

disease and the role of these cells in helminthic infection models in species other than the mouse.

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References

- Abbas A.K., Murphy K.M. and Sher A. (1996). Functional diversity of helper T lymphocytes. *Nature* 383, 787-793.
- Abe T., Sugaya H., Yoshimura K. and Nawa Y. (1992). Induction of the expulsion of *Strongyloides ratti* and retention of *Nippostrongylus brasiliensis* in athymic nude mice by repetitive administration of recombinant IL-3. *Immunology* 76, 10-14.
- Abraham W.M., Sielczak M.W., Ahmed A., Cortes A., Lauredo I.T., Kim J., Pepinsky B., Benjamin C.D., Leone D.R. and Lobb R.R. (1994). Alpha 4-integrins mediate antigen-induced late bronchial responses and prolonged airway hyperresponsiveness in sheep. *J. Clin. Invest.* 93, 776-787.
- Abraham D., Rotman H.L., Haberstroh H.F., Yutanawiboonchai W., Brigandi R.A., Leon O., Nolan T.J. and Schad G.A. (1995). *Strongyloides stercoralis*: protective immunity to third-stage larvae in BALB/cByJ mice. *Exp. Parasitol.* 80, 297-307.
- Abu-Ghazaleh R.I., Dunnette S.L., Loegering D.A., Checkel J.L., Kita H., Thomas L.L. and Gleich G.J. (1992). Eosinophil granule proteins in peripheral blood granulocytes. *J. Leukoc. Biol.* 52, 611-618.
- Ahmad A., Wang C.H. and Bell R.G. (1991). A role for IgE in intestinal immunity: Expression of rapid expulsion of *Trichinella spiralis* in rats transfused with IgE and thoracic duct lymphocytes. *J. Immunol.* 146, 3563-3570.
- Alizadeh H. and Murrell K.D. (1984). The intestinal mast cell response to *Trichinella spiralis* infection in mast cell-deficient w/wv mice. *J. Parasitol.* 70, 767-773.
- Alnemri E.S., Livingston D.J., Nicholson D.W., Salvesen G., Thornberry N.A., Wong W.W. and Yuan J. (1996). Human ICE/CED-3 protease nomenclature. *Cell* 87, 171.
- Alon R., Kassner P.D., Carr M.W., Finger E.B., Hemler M.E. and Springer T.A. (1995). The integrin VLA-4 supports tethering and rolling in flow on VCAM-1. *J. Cell Biol.* 128, 1243-1253.
- Asakura K., Saito H. and Kataura A. (1996). *in vivo* effects of monoclonal antibody against ICAM-1 and LFA-1 on antigen-induced nasal symptoms and eosinophilia in sensitized rats. *Int. Arch. Allergy Immunol.* 111, 156-160.
- Anwar A.R., Moqbel R., Walsh G.M., Kay A.B. and Wardlaw A.J. (1993). Adhesion to fibronectin prolongs eosinophil survival. *J. Exp. Med.* 177, 839-843.
- Askenase P.W. (1979). Immunopathology of parasitic diseases: Involvement of basophils and mast cells. *Springer Semin. Immunopathol.* 2, 2.
- Baggiolini M. (1998). Chemokines and leukocyte traffic. *Nature* 392, 565-568.
- Barnes P.J. (1998a). Current issues for establishing inhaled corticosteroids as the anti-inflammatory agents of choice in asthma. *J. Allergy Clin. Immunol.* 101, S427-433.
- Barnes P.J. (1998b). Anti-inflammatory actions of glucocorticoids: molecular mechanisms. *Clin. Sci. (Colch)* 94, 557-572.
- Bosse R. and Vestweber D. (1994). Only simultaneous blocking of the L- and P-selectin completely inhibits neutrophil migration into mouse peritoneum. *Eur. J. Immunol.* 24, 3019-3024.
- Bousquet J., Chanez P., Lacoste J.Y., White R., Vic P., Godard P. and Michel F.B. (1992). Asthma: a disease remodeling the airways. *Allergy* 47, 3-11.
- Boyce J.A., Friend D., Marsumoto R., Austen K.F. and Owen W.F. (1995). Differentiation *in vitro* of hybrid eosinophil/basophil granulocytes: autocrine function of an eosinophil developmental intermediate. *J. Exp. Med.* 182, 49-57.
- Bozza P.T., Yu W., Penrose J.F., Morgan E.S., Dvorak A.M. and Weller P.F. (1997). Eosinophil lipid bodies: specific, inducible intracellular sites for enhanced eicosanoid formation. *J. Exp. Med.* 186, 909-920.
- Bradding P., Roberts J.A., Britten K.M., Montefort S., Djukanovic R., Mueller R., Heusser C.H., Howarth P.H. and Holgate S.T. (1994). Interleukin-4, -5, and -6 and tumor necrosis factor-alpha in normal and asthmatic airways: evidence for the human mast cell as a source of these cytokines. *Am. J. Respir. Cell. Mol. Biol.* 10, 471-480.
- Brassart D., Kolodziejczyk E., Granato D., Woltz A., Pavillard M., Perotti B., Frigeri L.G., Liu F.-T., Borel Y. and Neeser J.-R. (1992). An intestinal galactose-specific lectin mediates the binding of murine IgE to mouse intestinal epithelial cells. *Eur. J. Biochem.* 203, 393-396.
- Broide D.H., Humber D. and Sriramarao P. (1998a). Inhibition of eosinophil rolling and recruitment in P-selectin and intracellular adhesion molecule-1-deficient mice. *Blood* 91, 2847-2856.
- Broide D.H., Sullivan S., Gifford T. and Sriramarao P. (1998b). Inhibition of pulmonary eosinophilia in P-selectin- and ICAM-1-deficient mice. *Am. J. Respir. Cell. Mol. Biol.* 18, 218-225.
- Brujinzeel-Koomen C., Storz E., Menz G. and Brujinzeel P. (1992). Skin eosinophilia in patients with allergic and nonallergic asthma and atopic dermatitis. *J. Allergy Clin. Immunol.* 89, 52-59.
- Bundy D.A. (1994). Immunoepidemiology of intestinal helminthic infections. 1: The global burden of intestinal nematode disease. *Trans. R. Soc. Trop. Med. Hyg.* 88, 259-261.
- Busse W., Nelson H., Wolfe J., Kalberg C., Yancey S.W. and Rickard K.A. (1999). Comparison of inhaled salmeterol and oral zafirlukast in patients with asthma. *J. Allergy Clin. Immunol.* 103, 1075-1080.
- Butterworth A.E. (1984). Cell mediated damage to helminths. *Adv. Parasitol.* 23, 143-235.
- Caldenhoven E., van Dijk T., Raaijmakers J.A., Lammers J.W., Koenderman L. and De Groot R.P. (1995). Activation of the STAT3/acute phase response factor transcription factor by interleukin-5. *J. Biol. Chem.* 270, 25778-25784.
- Capron M. and Joseph M. (1991). The low affinity receptor for IgE on eosinophils and platelets. In CD23: A novel multifunctional regulator of the immune system that binds IgE. Gordon J. (ed). *Monogr. Allergy*. Vol. 29. Karger, Basel. pp 63-75.
- Capron M., Bazin H., Torpier G., Joseph M. and Capron A. (1981). Evidence for IgE-dependent cytotoxicity by rat eosinophils. *J. Immunol.* 126, 1764-1768.
- Carlos T.M. and Harlan J.M. (1994). Leukocyte-endothelial adhesion molecules. *Blood* 84, 2068-2101.
- Chand N., Harrison J.E., Rooney S., Pillar J., Jakubicki R., Nolan K., Diamantis W. and Sofia R.D. (1992). Anti-IL-5 monoclonal antibody inhibits allergic late phase bronchial eosinophilia in guinea pigs: a therapeutic approach. *Eur. J. Pharmacol.* 211, 121-123.
- Cheever A.W., Xu Y.H., Sher A. and Macedonia J.G. (1991). Analysis of egg granuloma formation in *Schistosoma japonicum*-infected mice

- treated with antibodies to interleukin-5 and gamma interferon. *Infect. Immun.* 59, 4071-4074.
- Chin J.E., Winterrowd G.E., Hatfield C.A., Brashler J.R., Griffin R.L., Vonderfecht S.L., Kolbasa K.P., Fidler S.F., Shull K.L., Krzesicki R.F., Ready K.A., Dunn C.J., Sly L.M., Staite N.D. and Richards I.M. (1998). Involvement of intercellular adhesion molecule-1 in the antigen-induced infiltration of eosinophils and lymphocytes into the airways in a murine model of pulmonary inflammation. *Am. J. Respir. Cell. Mol. Biol.* 18, 158-167.
- Coffman R.L., Seymour B.W., Hudak S., Jackson J. and Rennick D. (1989). Antibody to interleukin-5 inhibits helminth-induced eosinophilia in mice. *Science* 245, 308-310.
- Colgan S.P., Resnick M.B., Parkos C.A., Delp-Archer C., McGuirk D., Bacarra A.E., Weller P.F. and Madara J.L. (1994). IL-4 directly modulates function of a model human intestinal epithelium. *J. Immunol.* 153, 2122-2129.
- Collins P.D., Marleau S., Griffiths-Johnson D.A., Jose P.J. and Williams T.J. (1995). Cooperation between interleukin-5 and the chemokine eotaxin to induce eosinophil accumulation *in vivo*. *J. Exp. Med.* 182, 1169-1174.
- Das A.M., Flower R.J. and Perreti M. (1997). Eotaxin-induced eosinophil migration in the peritoneal cavity of ovalbumin-sensitized mice: mechanism of action. *J. Immunol.* 159, 1466-1473.
- Daugherty B.L., Siciliano S.J., DeMartino J.A., Malkowitz L., Sirotna A. and Springer M.S. (1996). Cloning, expression, and characterization of the human eosinophil eotaxin receptor. *J. Exp. Med.* 183, 2349-2354.
- De Andres B., Rakasz E., Hagen M., McCormik M.L., Mueller A.L., Elliot D., Metwali A., Sandor M., Britigan B.E., Weinstock J.V. and Lynch R.G. (1997). Lack of Fc-epsilon receptors on murine eosinophils: implications for the functional significance of elevated IgE and eosinophils in parasitic infections. *Blood* 89, 3826-3836.
- De Sanctis G.T., Wolyniec W.W., Green F.H., Qin S., Jiao A., Finn P.W., Noonan T., Joetham A.A., Gelfand E., Doerschuk C.M. and Drazen J.M. (1997). Reduction of allergic airway responses in P-selectin-deficient mice. *J. Appl. Physiol.* 83, 681-687.
- Dent L.A., Strath M., Mellor A.L. and Sanderson C.J. (1990). Eosinophilia in transgenic mice expressing interleukin 5. *J. Exp. Med.* 172, 1425-1431.
- Dent L.A., Daly C.M., Mayhofer G., Zimmerman T., Halett A., Bignold L.P., Creaney J. and Parsons J.C. (1999). Interleukin-5 transgenic mice show enhanced resistance to primary infection with *Nippostrongylus brasiliensis* but not primary infections with *Toxocara canis*. *Infect. Immun.* 67, 989-993.
- Dent L.A., Munro G.H., Piper K.P., Sanderson C.J., Finlay D.A., Dempster R.K., Bignold L.P., Harkin D.G. and Hagan P. (1997). Eosinophilic interleukin 5 (IL-5) transgenic mice: eosinophil activity and impaired clearance of *Schistosoma mansoni*. *Parasite Immunol.* 19, 291-300.
- Dessein A.J., Parker W.L., James S.L. and David J.R. (1981). IgE antibody and resistance to infection. I. Selective suppression of the IgE antibody response in rats the resistance and the eosinophil response to *Trichinella spiralis* infection. *J. Exp. Med.* 153, 423-436.
- Djukanovic R., Roche W.R., Wilson J.W., Beasley C.R.W., Twentyman O.P., Howarth P.H. and Holgate S.T. (1990). Mucosal inflammation in asthma. *Am. Rev. Respir. Dis.* 142, 434-457.
- Druilhe A., Cai Z., Haile S., Chouaib S. and Pretolani M. (1996). Fas-mediated apoptosis in cultured human eosinophils. *Blood* 87, 2822-2830.
- Dunne D.W., Butterworth A.E., Fulford A.J.C., Kariuki H.C., Langley J.G., Ouma J.H., Capron A., Pierce R.J. and Sturrock R.F. (1992). Immunity after treatment of human schistosomiasis: Association between IgE antibody to adult worm antigens and resistance to reinfection. *Eur. J. Immunol.* 22, 1483-1494.
- Dupuis R., Collins D.S., Koh Y.Y., Pollice M., Albertine K.H., Fish J.E. and Peters S.P. (1992). Effect of antigen dose on the recruitment of inflammatory cells to the lung by segmental antigen challenge. *J. Allergy Clin. Immunol.* 89, 850-857.
- Dvorak A.M., Dvorak H.F., Peters S.P., Shulman E.S., MacGlashan D.W. Jr., Pyne K., Harvey V.S., Galli S.J. and Lichtenstein L.M. (1983). Lipid bodies: cytoplasmic organelles important to arachidonate metabolism in macrophages and mast cells. *J. Immunol.* 131, 2965-2976.
- Dvorak A.M., Letourneau L., Login G.R., Weller P.F. and Ackerman S.J. (1988). Ultrastructural localization of the "charcot-Leyden" crystal protein to a distinct crystalloid-free granule population in mature human eosinophils. *Blood* 72, 150-158.
- Dvorak A.M., Ackerman S.J. and Weller P.F. (1991). Subcellular morphology and biochemistry of eosinophils. In: *Blood cell biochemistry*. Dvorak A.M. and Harris J.R. (eds) Plenum Press. pp 237-344.
- Egan R.W., Athwahl D., Chou C.C., Chapman R.W., Emtage S., Jenh C.H., Kung T.T., Mauser P.J., Murgolo N.J. and Bodmer M.W. (1997). Pulmonary biology of anti-interleukin 5 antibodies. *Mem. Inst. Oswaldo Cruz* 92 (Suppl 2), 69-73.
- Else K.J., Finkelman F.D., Maliszewski C.R. and Grecnis R.K. (1994). Cytokine-mediated regulation of chronic intestinal helminth infection. *J. Exp. Med.* 179, 347-351.
- Else K.J., Hulter L. and Grecnis R.K. (1992). Cellular immune responses to the murine nematode parasite *Trichuris muris* II: Differential induction of Th-cell subsets in resistant versus susceptible mice. *Immunology* 75, 232-237.
- Finkelman F.D., Katona I.M. and Urban Jr. J.F. (1988). IL-4 is required to generate and sustain *in vivo* IgE response. *J. Immunol.* 141, 2335-2341.
- Folkard S.G., Hogarth P.J., Taylor M.J. and Bianco A.E. (1996). Eosinophils are the major effector cells of immunity to microfilariiae in a mouse model of onchocerciasis. *Parasitology* 112, 323-329.
- Foster C.S., Rice B.A. and Dutt J.E. (1991). Immunopathology of atopic keratoconjunctivitis. *Ophthalmology* 98, 1190-1196.
- Foster P.S., Hogan S.P., Ramsay A.J., Matthaci K.I. and Young I.G. (1996). Interleukin-5 deficiency abolishes eosinophilia, airways hyperreactivity, and lung damage in a mouse asthma model. *J. Exp. Med.* 183, 195-201.
- Freeman Jr. G.L., Tominaga A., Takatsu K., Secor W.E. and Colley D.G. (1995). Elevated innate peripheral blood eosinophilia fails to augment irradiated cercarial vaccine-induced resistance to *Schistosoma mansoni* in IL-5 transgenic mice. *J. Parasitol.* 81, 1010-1011.
- Frigeri L.G. and Liu F-T. (1992). Surface expression of functional IgE binding protein and endogenous lectin on mast cells and macrophages. *J. Immunol.* 148, 861-867.
- Fryer A.D., Costello R.W., Yost B.L., Lobb R.R., Tedder T.F., Steeber D.A. and Bochner B.S. (1997). Antibody to VLA-4, but not to L-selectin, protects neuronal M2 muscarinic receptors in antigen-challenged guinea pig airways. *J. Clin. Invest.* 99, 2036-2044.
- Gerwin N., Gonzalo J.-A., Lloyd C., Coyle A.J., Reiss Y., Banu N., Wang B., Xu H., Avraham H., Engelhard B., Springer T.A. and Gutierrez-

- Ramos J.C. (1999). Prolonged eosinophil accumulation in allergic lung interstitium of ICAM-2-deficient mice results in extended hyperresponsiveness. *Immunity* 10, 9-19.
- Giembycz M.A. (1996). Phosphodiesterase 4 and tolerance to beta 2-adrenoceptor agonists in asthma. *Trends Pharmacol. Sci.* 17, 331-336.
- Giembycz M.A. and Lindsay M.A. (1999). Pharmacology of the eosinophil. *Pharmacol. Ther.* 51, 213-339.
- Giembycz M.A. and Dent G. (1992). Prospects for selective cyclic nucleotide phosphodiesterase inhibitors in the treatment of bronchial asthma. *Clin. Exp. Allergy* 22, 337-344.
- Gleich G.J., Olson G.M. and Herlich H. (1979). The effect of antiserum to eosinophils and susceptibility and acquired immunity of the guinea-pig to *Trichostrongylus colubriformis*. *Immunology* 37, 873-880.
- Goldhill J., Morris S., Finkelman F., Pineiro Carrero V. and Shea-Donohue T. (1997). Interleukin (IL)-4 alters neural control of mouse small intestinal longitudinal muscle. *Am. J. Physiol.* 272, G1135-G1140.
- Gonzalo J.A., Lloyd C.M., Kremer L., Finger E., Martinez-A.C., Siegelman M.H., Cybulsky M. and Gutierrez-Ramos J.C. (1996). Eosinophil recruitment to the lung in a murine model of allergic inflammation. The role of T cells, chemokines, and adhesion receptors. *J. Clin. Invest.* 98, 2332-2345.
- Gounni A.S., Lamkhioued B., Ochiai K., Tanaka Y., Delaporte E., Capron A., Kinet J.P. and Capron M. (1994). High-affinity IgE receptor on eosinophils is involved in defence against parasites. *Nature* 367, 183-186.
- Grabbe J., Haas N., Hamann K., Kolde G., Hakimi J. and Czarnetzki B. M. (1993). Demonstration of high-affinity IgE receptor on human Langerhans cells in normal and diseased skin. *Br. J. Dermatol.* 129, 120-123.
- Green D.R. and Ware C.F. (1997). Fas-ligand: privilege and peril. *Proc. Natl. Acad. Sci. USA* 94, 5986-5990.
- Grewe M., Bruijnzeel-Koomen C.A., Schopf E., Thepen T., Langeveld-Wildschut A.G., Ruzicka T. and Krutmann J. (1998). A role for Th1 and Th2 cells in the immunopathogenesis of atopic dermatitis. *Immunol. Today* 19, 359-361.
- Grove D.I., Mahmoud A.A. and Warren K.S. (1977). Eosinophils and resistance to *Trichinella spiralis*. *J. Exp. Med.* 145, 755-759.
- Gundel R.H., Wegner C.D. and Letts L.G. (1992). Antigen-induced acute and late phase responses in primates. *Am. Rev. Respir. Dis.* 146, 369-373.
- Hagan P. (1993). IgE and protective immunity to helminth infections. *Parasite Immunol.* 15, 1-4.
- Hagan P., Moore P.J., Adjukiewicz A.B., Greenwood B.M. and Wilkins H.A. (1985). *In vitro* antibody-dependent killing of schistosomula of *Schistosoma haematobium* by human eosinophils. *Parasite Immunol.* 7, 625-632.
- Hall L.R., Mehlotra R.K., Higgins A.W., Haxhiu M.A. and Pearlman E. (1998). An essential role for interleukin-5 and eosinophils in helminth-induced airway hyperresponsiveness. *Infect. Immun.* 66, 4425-4430.
- Hebestreit H., Dibbert B., Balatti I., Braun D., Schapowal A., Blaser K. and Simon H.U. (1998). Disruption of Fas receptor signaling by nitric oxide in eosinophils. *J. Exp. Med.* 187, 415-425.
- Hebestreit H., Yousefi S., Balatti I., Weber M., Cramer R., Simon D., Hartung K., Schapowal A., Blaser K. and Simon H.U. (1996). Expression and function of the Fas receptor on human blood and tissue eosinophils. *Eur. J. Immunol.* 26, 1775-1780.
- Henriques M.G.M.O., Miotla J.M., Cordeiro R.S.B., Wolitsky B.A., Wolley S.T. and Hellewell P.G. (1996). Selectins mediate eosinophil recruitment *in vivo*: A comparison with their role in neutrophil influx. *Blood* 87, 5297-5304.
- Her E., Frazer F., Austin K.F. and Owen W. (1991). Eosinophil hematopoietins antagonise the programmed cell death of eosinophils. *J. Clin. Invest.* 88, 1982-1987.
- Herndon F.J. and Kaye S.G. (1992). Depletion of eosinophils by anti-IL-5 monoclonal antibody treatment of mice infected with *Trichinella spiralis* does not alter parasite burden or immunologic resistance to reinfection. *J. Immunol.* 149, 3642-3647.
- Hirsch J.G. and Hirsch B.I. (1980). Paul Ehrlich and the discovery of the eosinophil. In: *The eosinophil in health and disease*. Mahmoud A.A.F. and Austen K.F. (eds). Grune & Stratton, Inc. New York. pp 3-23.
- Hisada T., Hellewell P.G., Teixeira M.M., Malm M.G., Salmon M., Huang T.J., Chung K.F. (1999). Alpha4 integrin-dependent eotaxin induction of bronchial hyperresponsiveness and eosinophil migration in interleukin-5 transgenic mice. *Am. J. Respir. Cell. Mol. Biol.* 20, 992-1000.
- Hokibara S., Takamoto M., Tominaga A., Takatsu K. and Sugane K. (1997). Marked eosinophilia in interleukin-5 transgenic mice fails to prevent *Trichinella spiralis* infection. *J. Parasitol.* 83, 1186-1189.
- Horii Y., Khan A.I. and Nawa Y. (1993). Persistent infection of *Strongyloides venezuelensis* and normal expulsion of *Nippostrongylus brasiliensis* in Mongolian gerbils, *Meriones unguiculatus*, with reference to the cellular responses in the intestinal mucosa. *Parasite Immunol.* 15, 175-179.
- Ishikawa N., Horii Y. and Nawa Y. (1993). Immune-mediated alteration of the terminal sugars of goblet cell mucins in the small intestine of *Nippostrongylus brasiliensis*-infected rats. *Immunology* 78, 303-307.
- Ishikawa N., Horii Y., and Nawa Y. (1994). Reconstitution by bone marrow grafting of the defective protective capacity at the migratory phase but not at the intestinal phase of *Nippostrongylus brasiliensis* infection in W/W^v mice. *Parasite Immunol.* 16, 181-186.
- Jarrett E.E.E. and Miller H.R.P. (1982). Production and activities of IgE in helminth infections. *Prog. Allergy* 31, 178-233.
- Jia G.Q., Gonzalo J.A., Hidalgo A., Wagner D., Cybulsky M. and Gutierrez-Ramos J.C. (1999). Selective eosinophil transendothelial migration triggered by eotaxin via modulation of Mac-1/ICAM-1 and VLA-4/VCAM-1 interactions. *Int. Immunol.* 11, 1-10.
- Jutila M.A., Wilson E. and Kurk S. (1997). Characterization of an adhesion molecule that mediates leukocyte rolling on 24 h cytokine- or lipopolysaccharide-stimulated bovine endothelial cells under flow conditions. *J. Exp. Med.* 186, 1701-1711.
- Kan H., Ogata T., Taniyama A., Migita M., Matsuda I. and Nawa Y. (1995). Extraordinarily high eosinophilia and elevated serum interleukin-5 level observed in a patient infected with *Paragonimus westermani*. *Pediatrics* 96, 351-354.
- Kansas G.S. (1996). Selectins and their ligands: current concepts and controversies. *Blood* 88, 3259-3287.
- Kanwar S., Bullard D.C., Hickey M.J., Smith C.W., Beaudet A.L., Wolitsky B.A. and Kubers P. (1997). The association between alpha4-integrin, P-selectin, and E-selectin in an allergic model of inflammation. *J. Exp. Med.* 185, 1077-1087.
- Kemp J.P., Minkwitz M.C., Bonuccelli C.M. and Warren M.S. (1999). Therapeutic effect of zafirlukast as monotherapy in steroid-naive patients with severe persistent asthma. *Chest* 115, 336-342.

- Klementsson H. (1992). Eosinophils and the pathophysiology of allergic rhinitis. *Clin. Exp. Allergy* 22, 1058-1064.
- Kopf M., Brombacher F., Hodgkin P., Ramsay A.J., Milbourne E.A., Dai W.J., Ovington K.S., Bhem C.A., Köhler G., Young I.G. and Matthaei K.I. (1996). IL-5 deficient mice have a developmental defect in CD5+ B-1 cells and lack eosinophilia but have normal antibody and cytotoxic T cell responses. *Immunity* 4, 15-24.
- Korenaga M., Hitoshi Y., Takatsu K. and Tada I. (1994). Regulatory effect of anti-interleukin-5 monoclonal antibody on intestinal worm burden in a primary infection with *Strongyloides venezuelensis* in mice. *Int. J. Parasitol.* 24, 951-957.
- Korenaga M., Hitoshi Y., Yamaguchi N., Sato Y., Takatsu K. and Tada I. (1991). The role of interleukin-5 in protective immunity to *Strongyloides venezuelensis* infection in mice. *Immunology* 72, 502-507.
- Korenaga M., Nawa Y. and Tada I. (1986). IgE response in *Strongyloides ratti*-infected rats with special reference to the life cycle of the parasite. *Z. Parasitenkd* 72, 213-220.
- Kraneveld A.D., Van Ark I., Van Der Linde H.J., Fattah D., Nijkamp F.P. and Van Oosterhout A.J. (1997). Antibody to very late activation antigen 4 prevents interleukin-5-induced airway hyper-responsiveness and eosinophil infiltration in the airways of guinea pigs. *J. Allergy Clin. Immunol.* 100, 242-250.
- Kuiter L.M., Angus R.M., Barnes N.C., Barnes P.J., Bone M.F., Chung K.F., Fairfax A.J., Higenbotham T.W., O'Connor B.J. and Piotrowska B. (1995) Effect of a novel potent platelet-activating factor antagonist, modipafant, in clinical asthma. *Am J Respir Crit Care Med* 151, 1331-5.
- Kung T.T., Stelts D.M., Zurcher J.A., Adams G.K. 3rd, Egan R.W., Kreutner W., Watnick A.S., Jones H. and Chapman R.W. (1995). Involvement of IL-5 in a murine model of allergic pulmonary inflammation: prophylactic and therapeutic effect of an anti-IL-5 antibody. *Am. J. Respir. Cell. Mol. Biol.* 13, 360-365.
- Kurniawan A., Yazdanbakhsh M., van Ree R., Aalberse R., Selkirk M.E., Partono F. and Maizels R. M. (1993). Differential expression of IgE and IgG4 specific antibody responses in asymptomatic and chronic human filariasis. *J. Immunol.* 150, 3941-3950.
- Laberge S., Rabb R., Issekutz T.B. and Martin J.G. (1995). Role of VLA-4 and LFA-1 in allergen-induced airway hyperresponsiveness and lung inflammation in the rat. *Am. J. Respir. Crit. Care Med.* 151, 822-829.
- Labow M.A., Norton C.R., Rumberger J.M., Lombard-Gillooly K.M., Shuster D.J., Hubbard J., Bertko R., Knaack P.A., Terry R.W., Harbison M.L., Kontgen F., Stewart C.L., McIntyre K.W., Will P.C., Burns D.K. and Wolitzky B.A. (1994). Characterization of E-selectin-deficient mice: Demonstration of overlapping function of the endothelial selectins. *Immunity* 1, 709-720.
- Laitinen L.A., Laitinen A. and Haahela T. (1993) Airway mucosal inflammation even in patients with newly diagnosed asthma. *Am. Ver. Respir. Dis.* 147, 697-704.
- Lake A.M., Bloch K.J., Sinclair K.J. and Walker W.A. (1980). Anaphylactic release of intestinal goblet cell mucus. *Immunology* 39, 173-178.
- Lange A.M., Yutanawiboonchai W., Scott P. and Abraham D. (1994). IL-4- and IL-5-dependent protective immunity to *Onchocerca volvulus* infective larvae in BALB/cBYJ mice. *J. Immunol.* 153, 205-211.
- Larsh J.E. and Race G.J. (1975). Allergic inflammation as a hypothesis for the expulsion of worms from tissue: A review. *Exp. Parasitol.* 37, 251-266.
- Lebowitz M.D. and Spinaci S. (1993). The epidemiology of asthma. *Eur. Respir. Rev.* 3, 415-423.
- Lefort J., Nahori M.A., Ruffie C., Vargaftig B.B. and Pretolani M. (1996). *in vivo* neutralization of eosinophil-derived major basic protein inhibits antigen-induced bronchial hyperreactivity in sensitized guinea pigs. *J. Clin. Invest.* 97, 1117-1121.
- Ley K. and Tedder T.F. (1995). Leukocyte interactions with vascular endothelium. New insights into selectin-mediated attachment and rolling. *J. Immunol.* 155, 525-528.
- Lobb R.R. and Hemler M.E. (1994). The pathophysiologic role of alpha4 integrins *in vivo*. *J. Clin. Invest.* 94, 1722-1728.
- Love R.J., Ogilvie B.M. and McLaren D.J. (1976). The immune mechanism which expels the intestinal stage of *Trichinella spiralis* from rats. *Immunology* 30, 7-15.
- Macari D.M., Teixeira M.M., Ansari T., Jeffery P.K. and Hellewell P.G. (1998). Priming and induction of eosinophil trafficking in guinea-pig cutaneous inflammation by tumour necrosis factor alpha. *Br. J. Pharmacol.* 125, 1228-1235.
- Macari D.M., Teixeira M.M. and Hellewell P.G. (1996). Priming of eosinophil recruitment *in vivo* by LPS pretreatment. *Immunology* 157, 1684-1692.
- Madden K.B., Urban Jr. J.F., Ziltener H.J., Schrader J.W., Filkelman F.D. and Katona I.M. (1991). Antibodies to IL-3 and IL-4 suppress helminth-induced intestinal mastocytosis. *J. Immunol.* 147, 1387-1391.
- Magnussen P., Makunde W., Simonsen P.E., Meyrowitsch D. and Jakubowski K. (1995). Chronic pulmonary disorders, including tropical pulmonary eosinophilia, in villages with endemic lymphatic filariasis in Tanga region and in Tanga town, Tanzania. *Trans. R. Soc. Trop. Med. Hyg.* 89, 406-409.
- Malmstrom K., Rodriguez-Gomez G., Guerra J., Villaran C., Pineiro A., Wei L.X., Seidenberg B.C. and Reiss T.F. (1999). Oral montelukast, inhaled beclomethasone, and placebo for chronic asthma. A randomized, controlled trial. Montelukast/Beclomethasone Study Group. *Ann. Intern. Med.* 130, 487-495.
- Mansfield L.S. and Gamble H.R. (1995). Alveolar mastocytosis and eosinophilia in lambs with naturally acquired nematode infections of *Protostrongylus rufescens* and *Haemonchus contortus*. *Vet. Immunol. Immunopathol.* 49, 251-262.
- Matsumoto K., Schleimer R.P., Saito H., Iikura Y. and Bochner B.S. (1995). Induction of apoptosis in human eosinophils by anti-Fc ϵ 1 antibody treatment *in vitro*. *Blood* 86, 1437-1443.
- Matthaei K.I., Foster P.S. and Young I.G. (1997). The role of interleukin 5 (IL-5) *in vivo*: Studies with IL-5 deficient mice. *Mem. Inst. Oswaldo Cruz Suppl.* 2, 63-68.
- Matthews A.N., Friend D.S., Zimmermann N., Sarafi M.N., Luster A.D., Pearlman E., Wert S.E. and Rothenberg M.E. (1998). Eotaxin is required for the baseline level of tissue eosinophils. *Proc. Natl. Acad. Sci. USA* 95, 6273-6878.
- Mausner P.J., Pitman A.M., Fernandez X., Foran S.K., Adams G.K. 3rd, Kreutner W., Egan R.W. and Chapman R.W. (1995). Effects of an antibody to interleukin-5 in a monkey model of asthma. *Am. J. Respir. Crit. Care Med.* 152, 467-472.
- Mausner P.J., Pitman A., Witt A., Fernandez X., Zurcher J., Kung T., Jones H., Watnick A.S., Egan R.W., Kreutner W. and Chapman R.W. (1993). Inhibitory effect of the TRFK-5 anti-IL-5 antibody in a guinea pig model of asthma. *Am. Ver. Respir. Dis.* 148, 1623-1627.
- McGee D.W. and Vitkus S.J.D. (1996). IL-4 enhances IEC-6 intestinal epithelial cell proliferation yet has no effect on IL-6 secretion. *Clin.*

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- Exp. Immunol. 105, 274-277.
- Milne A.A. and Piper P.J. (1994). The effects of two anti-CD18 antibodies on antigen-induced airway hyperresponsiveness and leukocyte accumulation in the guinea pig. *Am. J. Respir. Cell. Mo. Biol.* 11, 337-43.
- Milne A.A. and Piper P.J. (1995). Role of the VLA-4 integrin in leukocyte recruitment and bronchial hyperresponsiveness in the guinea-pig. *Eur. J. Pharmacol.* 282, 243-249.
- Milne A.A.Y., Teixeira M.M., Hellewell P.G. and Piper P.J. (1995). Induction of leukocyte recruitment and bronchial hyperresponsiveness in the guinea pig by aerosol administration of interleukin-2. *Int. Arch. Allergy Immunol.* 108, 60-67.
- Mogbel R. (1980). Histopathological changes in rats following primary, secondary and repeated infections with *Strongyloides ratti*, with special reference to tissue eosinophils. *Parasite Immunol.* 2, 11-17.
- Morley J. (1993). Immunopharmacology of asthma. *Immunol. Today* 14, 317-322.
- Mosmann T.R. and Coffman R.L. (1989). Th1 and Th2 cells: Different patterns of lymphokine secretion lead to different functional properties. *Annu. Rev. Immunol.* 7, 145-173.
- Mosmann T.R. and Sad S. (1996). The expanding universe of T-cell subsets: Th1, Th2 and more. *Immunol. Today* 17, 138-146.
- Mui A.L., Wakao H., O'Farrell A.M., Harada N. and Miyajima A. (1995). Interleukin-3, granulocyte-macrophage colony stimulating factor and interleukin-5 transduce signals through two STAT5 homologs. *EMBO J.* 14, 1166-1175.
- Murakami M. and Honjo T. (1995). Involvement of B1 Cells in mucosal immunity and autoimmunity. *Immunol. Today* 16, 534-539.
- Murphy P.M. (1996). Chemokine receptors: structure, function and role in microbial pathogenesis. *Cyt. Growth Factor Rev.* 7, 47-64.
- Murray M. (1972). Immediate hypersensitivity effector mechanisms II. *in vivo* reaction. In: *Immunity to animal parasites*. Soulsby E.J.L. (ed). Academic Press. New York. pp 155-190.
- Musoke A.J., Williams J.F. and Leid R.W. (1978). Immunological response of the rat to infection with *Taenia taeniaeformis*. VI The role of immediate hypersensitivity in resistance to reinfection. *Immunology* 34, 565-570.
- Nagata S. (1997). Apoptosis by death factor. *Cell* 88, 355-365.
- Nakajima H., Sano H., Nishimura T., Yoshida S. and Iwamoto I. (1994). Role of vascular cell adhesion molecule 1/very late activation antigen 4 and intercellular adhesion molecule 1/lymphocyte function-associated antigen 1 interactions in antigen-induced eosinophil and T cell recruitment into the tissue. *J. Exp. Med.* 179, 1145-1154.
- Nawa Y., Kiyota M., Korenaga M. and Kotani M. (1985). Defective protective capacity of W/W^v mice against *Strongyloides ratti* infection and its reconstitution with bone marrow cells. *Parasite Immunol.* 7, 429-438.
- Negrão-Corrêa D. (1997). The intestinal IgE response to infection with *Trichinella spiralis* in rats. Tese de Doutorado, Universidade de Cornell USA.
- Negrão-Corrêa D., Adams L.S. and Bell R.G. (1996). Intestinal transport and catabolism of IgE. A major blood-independent pathway of IgE dissemination during a *Trichinella spiralis* infection of rats. *J. Immunol.* 157, 4037-4044.
- Negrão-Corrêa D., Adams L.S. and Bell R.G. (1999). Variability of the intestinal immunoglobulin E response of rats to infection with *Trichinella spiralis*, *Heligmosomoides polygyrus* or *Nippostrongylus brasiliensis*. *Parasite Immunol.* 21, 287-297.
- Noonan T.C., Gundel R.H., Desai S.N., Stearns C., Barton R.W., Rothlein R., Letts L.G. and Piper P.J. (1991). The effects of an anti-CD18 antibody (R15.7) in antigen-induced airway hyperresponsiveness (AH) and cell influx in guinea pigs. *Agents Actions* 34, 211-213.
- O'Byrne P.M., Dolovich J. and Hargreave F.E. (1987). Late asthmatic responses. *Am. Ver. Respir. Dis.* 136, 740-751.
- Ogata N., Kouro T., Yamada A., Koike M., Hanai N., Ishikawa T. and Takatsu K. (1998). JAK2 and JAK1 constitutively associate with an interleukin-5 (IL-5) receptor alpha and beta subunit, respectively, and are activated upon IL-5 stimulation. *Blood* 91, 2264-2271.
- Ovington K.S. and Behm C.A. (1997). The enigmatic eosinophil: investigation of the biological role of eosinophils in parasitic helminth infection. *Mem. Inst. Oswaldo Cruz Suppl* 2, 93-104.
- Ovington K.S., McKie K., Matthaai K.I., Young I.G. and Behm C.A. (1998). Regulation of primary *Strongyloides ratti* infections in mice: a role for interleukin-5. *Immunology* 95, 488-493.
- Pazdrak K., Schreiber D., Forsythe P., Justement L. and Alam R. (1995). The intracellular signal transduction mechanism of interleukin 5 in eosinophils: the involvement of lyn tyrosine kinase and the Ras-Raf-1-MEK-microtubule-associated protein kinase pathway. *J. Exp. Med.* 181, 1827-1834.
- Pazdrak K., Stafford S. and Alam R. (1995). The activation of the Jak-STAT 1 signaling pathway by IL-5 in eosinophils. *J. Immunol.* 155, 397-402.
- Peled A., Gonzalo J.A., Lloyd C. and Gutierrez-Ramos J.C. (1998). The chemotactic cytokine eotaxin acts as a granulocyte-macrophage colony-stimulating factor during lung inflammation. *Blood* 91, 1909-1916.
- Ponath P.D., Qin S., Post T.W., Wang J., Wu L., Gerard N.P., Newman W., Gerard C. and Mackay C.R. (1996). Molecular cloning and characterization of a human eotaxin receptor expressed selectively on eosinophils. *J. Exp. Med.* 183, 2437-2448.
- Power C.A. and Wells T.N.C. (1996). Cloning and characterization of human chemokine receptors. *TIPS* 17, 209-213.
- Pretolani M., Ruffie C., Joseph D., Campos M.G., Church M.K., Lefort J. and Vargaftig B.B. (1994). Role of eosinophil activation in the bronchial reactivity of allergic guinea pigs. *Am. J. Respir. Crit. Care Med.* 149, 1167-1174.
- Pritchard D.I., Quinnell R.J. and Wash E.A. (1995). Immunity in humans to *Necator americanus*: IgE, parasite weight and fecundity. *Parasite Immunol.* 17, 71-75.
- Ramaswamy K., Hakimi J. and Bell R.G. (1994). Evidence for an interleukin 4-inducible immunoglobulin E uptake and transport mechanism in the intestine. *J. Exp. Med.* 180, 1793-1803.
- Ramaswamy K., Negrão-Corrêa D. and Bell R.G. (1996). Local intestinal immune response to infections with *Trichinella spiralis*: real time, continuous assay of cytokines in the intestinal afferent and efferent thoracic duct lymph of rats. *J. Immunol.* 156, 4328-4337.
- Richards I.M., Kolbasa K.P., Winterrowd G.E., Hatfield C.A., Vonderfecht S.L., Fidler S.F., Griffin R.L., Brashler J.R., Krzesicki R.F., Lane C.L., Anderson D.C., Sly L.M., Staite N.D. and Chin J.E. (1996). Role of intercellular adhesion molecule-1 in antigen-induced lung inflammation in brown Norway rats. *Am. J. Physiol.* 27, L267-276.
- Robinson D.S., Hamid Q. and Ying S. (1992). Predominant Th2-like bronchoalveolar T-lymphocyte population in atopic asthma. *N. Engl. J. Med.* 326, 298-304.
- Romagnani S. (1994). Lymphokine production by human T cells in disease states. *Annu. Rev. Immunol.* 12, 227-257.

- Rothenberg M.E., Owen W.F. Jr, Silberstein D.S., Soberman R.J., Austen K.F. and Stevens R.L. (1987). Eosinophils cocultured with endothelial cells have increased survival and functional properties. *Science* 237, 645-647.
- Rothenberg M.E., MacLean J.A., Pearlman E., Luster A.D. and Leder P. (1997). Targeted disruption of the chemokine eotaxin partially reduces antigen-induced tissue eosinophilia. *J. Exp. Med.* 185, 785-790.
- Ruitenbergh E.J., Elgersma A. and Kruizinga W. (1979). Intestinal mast cell and globule leukocytes: Role of the thymus on their presence and proliferation during a *Trichinella spiralis* infection in the rat. *Int. Arch. Allergy Appl. Immunol.* 60, 302-309.
- Sagara H., Matsuda H., Wada N., Yagita H., Fukuda T., Okumura K., Makino S. and Ra C. (1997). A monoclonal antibody against very late activation antigen-4 inhibits eosinophil accumulation and late asthmatic response in a guinea pig model of asthma. *Int. Arch. Allergy Immunol.* 112, 287-294.
- Sanderson C.J. (1992). Interleukin-5, eosinophils, and disease. *Blood* 79, 3101-3109.
- Sanz M.J., Hartnell A., Chisholm P., Williams C., Davies D., Weg V.B., Feldmann M., Bolanowski M.A., Lobb R.R. and Nourshargh S. (1997). Tumor necrosis factor alpha-induced eosinophil accumulation in rat skin is dependent on alpha4 integrin/vascular cell adhesion molecule-1 adhesion pathways. *Blood* 15, 4144-4152.
- Sanz M.J., Marinova-Mutafchieva L., Green P., Lobb R.R., Feldmann M. and Nourshargh S. (1998). IL-4-induced eosinophil accumulation in rat skin is dependent on endogenous TNF-alpha and alpha 4 integrin/VCAM-1 adhesion pathways. *J. Immunol.* 160, 5637-5645.
- Sasaki O., Sugaya H., Ishida K. and Yoshimura K. (1993). Ablation of eosinophils with anti-IL-5 antibody enhances the survival of intracranial worms of *Angiostrongylus cantonensis* in the mouse. *Parasite Immunol* 15, 349-354.
- Satoh T., Chen Q.J., Sasaki G., Yokozeki H., Katayama I and Nishioka K. (1997). Cyclophosphamide-induced blood and tissue eosinophilia in contact sensitivity. Mechanisms of hapten-induced eosinophil recruitment into the skin. *Eur. J. Immunol.* 27, 85-91.
- Schneider T., Issekutz T.B. and Issekutz A.C. (1999). The role of alpha4 (CD49d) and beta2 (CD18) integrins in eosinophil and neutrophil migration to allergic lung inflammation in the Brown Norway rat. *Am. J. Respir. Cell. Mol. Biol.* 20, 448-457.
- Schwartz L.B. (1994). The molecular and cell biology of mast cells and basophils. In: *Molecular and cellular biology of the allergic response*. Levinson A.I. and Paterson Y. (eds). Marcel Dekker. New York. pp 281-330.
- Sher A., Coffman R.L., Hiery S. and Cheever A.W. (1990). Ablation of eosinophil and IgE responses with anti-IL-5 or anti-IL-4 antibodies fails to affect immunity against *Schistosoma mansoni* in the mouse. *J. Immunol.* 145, 3911-3916.
- Shin E.H., Osada Y., Chai J.Y., Matsumoto N., Takatsu K. and Kojima S. (1997). Protective roles of eosinophils in *Nippostrongylus brasiliensis* infection. *Int. Arch. Allergy Immunol.* 114 (Suppl 1), 45-50.
- Simon H.U. and Alam R. (1999). Regulation of eosinophil apoptosis: transduction of survival and death signals. *Int. Arch. Allergy Immunol.* 118, 7-14.
- Simon H.U., Yousefi S., Schranz C., Schapowal A., Bachert C. and Blaser K. (1997). Direct demonstration of delayed eosinophil apoptosis as a mechanism causing tissue eosinophilia. *J. Immunol.* 158, 3902-3908.
- Smith H. (1992). Asthma, inflammation, eosinophils and bronchial hyperresponsiveness. *Clin. Exper. Allergy* 22, 187-197.
- Springer T.A. (1994). Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. *Cell* 76, 301-314.
- Spry C.J.F. (1988). Eosinophils. A comprehensive review, and guide to the scientific and medical literature. Oxford University Press. Oxford, England.
- Spry C.J.F. (1993). The idiopathic hypereosinophilic syndrome. In: *Eosinophils: Biological clinical aspects*. Gleich G.J. and Kay A.B. (eds). CRC Press. FL. pp 403-420.
- Sriramarao P., von Andrian U.H., Butcher E.C., Bourdon M.A., Broide D.H. (1994). L-selectin and very late antigen-4 integrin promote eosinophil rolling at physiological shear rates *in vivo*. *J. Immunol.* 153, 4238-4246.
- Stamation P., Hamid Q., Taha R., Yu W., Issekutz T.B., Rokach J., Khanapure S.P. and Powel W.S. (1998). 5-Oxo-ETE induces pulmonary eosinophilia in an integrin-dependent manner in Brown Norway rats. *J. Clin. Invest.* 102, 2165-2172.
- Strath M., Dent L. and Sanderson C. (1992). Infection of IL5 transgenic mice with *Mesocostoides corti* induces very high levels of IL5 but depressed production of eosinophils. *Exp. Hematol.* 20, 229-234.
- Sugane K., Kusama Y., Takamoto M., Tominaga A. and Takatsu K. (1996). Eosinophilia, IL-5 level and recovery of larvae in IL-5 transgenic mice infected with *Toxocara canis*. *J. Helminthol.* 70, 153-158.
- Sugaya H., Aoki M., Yoshida T., Takatsu K. and Yoshimura K. (1997). Eosinophilia and intracranial worm recovery in interleukin-5 transgenic and interleukin-5 receptor alpha chain-knockout mice infected with *Angiostrongylus cantonensis*. *Parasitol. Res.* 83, 583-590.
- Sun J., Elwood W., Haczk A., Barnes P.J., Hellewell P.G. and Chung K.F. (1994). Contribution of intercellular-adhesion molecule-1 in allergen-induced airway hyperresponsiveness and inflammation in sensitised brown-Norway rats. *Int. Arch. Allergy Immunol.* 104, 291-295.
- Symon F.A., Lawrence M.B., Williamson M.L., Walsh G.M., Watson S.R. and Wardlaw A.J. (1996). Functional and structural characterization of the eosinophil P-selectin ligand. *J. Immunol.* 157, 1711-1719.
- Takamoto M., Ovington K.S., Behm C.A., Sugane K., Young I.G. and Matthaai K.I. (1997). Eosinophilia, parasite burden and lung damage in *Toxocara canis* infection in C57Bl/6 mice genetically deficient in IL-5. *Immunology* 90, 511-517.
- Teixeira M.M. (1998). Eosinophil-active chemokines - assessment of *in vivo* activity. *Braz. J. Med. Biol. Res.* 31, 19-24.
- Teixeira M.M. and Hellewell P.G. (1994). Effect of a 5-lipoxygenase inhibitor, ZM 230487, on cutaneous allergic inflammation in the guinea pig. *Br. J. Pharmacol.* 111, 1205-1211.
- Teixeira M.M. and Hellewell P.G. (1996). Effects of dexamethasone and cyclosporin A on the accumulation of eosinophils in acute inflammation in the guinea pig. *Br. J. Pharmacol.* 118, 317-324.
- Teixeira M.M. and Hellewell P.G. (1997). The selectin binding polysaccharide fucoidin inhibits eosinophil recruitment *in vivo*. *Br. J. Pharmacol.* 120, 1059-1066.
- Teixeira M.M. and Hellewell P.G. (1998). Contribution of endothelial selectins and alpha4 integrins in allergic and non-allergic inflammatory reactions in skin. *J. Immunol.* 161, 2516-2513.
- Teixeira M.M., Williams T.J. and Hellewell P.G. (1994a). Effect of phosphodiesterase isoenzyme inhibitors on cutaneous inflammation in the guinea pig. *Br. J. Pharmacol.* 112, 332-340.

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- Teixeira M.M., Reynia S., Robinson M., Shock A., Williams T.J., Williams F.M., Rossi A.G. and Hellewell, P.G. (1994b). Role of CD11/CD18 in mediating cutaneous inflammation in the guinea pig. *Br. J. Pharmacol.* 111, 811-818
- Teixeira M.M., Giembycz M.A., Lindsay M.A. and Hellewell P.G. (1997a). Pertussis toxin reveals distinct early signalling events in platelet-activating factor-, leukotriene B₄- and C5a-induced eosinophil homotypic aggregation *in vitro* and recruitment *in vivo*. *Blood* 89, 4566-4573.
- Teixeira M.M., Wells T.N.C., Lukacs N., Proudfoot A.E.I., Kunkel S., Williams T.J. and Hellewell P.G. (1997b). Chemokine-induced eosinophil recruitment *in vivo*: evidence of a role for endogenous eotaxin. *J. Clin. Invest.* 100, 1657-1666.
- Teixeira M.M., Gristwood R.W., Nicola C. and Hellewell P.G. (1997c). Phosphodiesterase (PDE)4 inhibitors: anti-inflammatory drugs of the future?. *Trends Pharmacol. Sci.* 18, 164-171.
- Teixeira M.M., Miotla J.M., Cooper N., Gristwood R.W. and Hellewell P.G. (1997). A comparison of the inhibitory activity of selective PDE4 inhibitors of eosinophil recruitment in guinea pig skin. *Memorias do Inst. Oswaldo Cruz* 92 (Suppl. II), 193-196.
- Teixeira M.M., Das A.M., Miotla J.M., Perretti M. and Hellewell P.G. (1998). Investigation into the role of lipocortin-1 in the inhibitory action of dexamethasone on eosinophil recruitment in cutaneous inflammatory reactions in the mouse. *Br. J. Pharmacol.* 123, 538-544.
- Teixeira M.M., Williams T.J. and Hellewell P.G. (1995). Mechanisms and pharmacological modulation of eosinophil accumulation *in vivo*. *Trends Pharmacol. Sci.* 16, 418-423.
- Terada N., Sagara H., Yagita H., Okumura K., Makino S., Konno A., Yamashita T., Togawa K. and Ra C. (1996). The effect of anti-VLA-4 monoclonal antibody on eosinophil accumulation and leukotriene production in nasal mucosa. *Acta Oto-Laryngol.* 116, 883-887.
- Truong M.J., Gruart V., Liu F.T., Prin L., Capron A. and Capron M. (1993a). IgE-binding molecules (Mac-2/epsilon BP) expressed by human eosinophils. Implication in IgE-dependent eosinophil cytotoxicity. *Eur. J. Immunol.* 12, 3230-3235.
- Truong M.J., Gruart V., Kusnierz J.P., Papin J.P., Loiseau S., Capron A. and Capron M. (1993b). Human neutrophils express immunoglobulin E (IgE)-binding proteins (Mac-2/epsilon BP) of the S-type lectin family: role in IgE-dependent activation. *J. Exp. Med.* 177, 243-248.
- Tsuyuki S., Bertrand C., Erard F., Trifillieff A., Tsuyuki J., Wesp M., Anderson G.P. and Coyle A.J. (1995). Activation of the Fas receptor on lung eosinophils leads to apoptosis and the resolution of eosinophilic inflammation of the airways. *J. Clin. Invest.* 96, 2924-2931.
- Uber C.L., Roth R.L. and Levy D.A. (1980). Expulsion of *Nippostrongylus brasiliensis* by mice deficient in mast cells. *Nature* 287, 226-228.
- Urban Jr J.F., Katona I.M., Paul W.E. and Finkelman F.D. (1991). Interleukin 4 is important in protective immunity to a gastrointestinal nematode infection in mice. *Proc. Natl. Acad. Sci. USA* 88, 5513-5517.
- Urban Jr J.F., Madden K.B., Svetic A., Cheever A., Trotta P.P., Gause W.C., Katona I.M. and Finkelman F.D. (1992). The importance of Th2 cytokines in protective immunity to nematodes. *Immunol. Rev.* 127, 205-220.
- Urban Jr. J.F., Maliszewski C.R., Madden K.B., Katona I.M. and Finkelman F.D. (1995). IL-4 treatment can cure established gastrointestinal nematode infection in immunocompetent and immunodeficient mice. *J. Immunol.* 154, 4675-4684.
- van der Bruggen T., Caldenhoven E., Kanters D., Coffey P., Raaijmakers J.A., Lammers J.W. and Koenderman L. (1995). Interleukin-5 signaling in human eosinophils involves JAK2 tyrosine kinase and STAT 1. *Blood* 85, 1442-1448.
- Varki A. (1994). Selectin ligands. *Proc. Natl. Acad. Sci. USA* 91, 7390-7397.
- Verwaerde C., Joseph M., Capron M., Pierce R.J., Damonville M., Velge F., Aurault C. and Capron A. (1987). Functional properties of a rat monoclonal IgE antibody specific for *Schistosoma mansoni*. *J. Immunol.* 138, 4441-4446.
- Wakelin D. and Wilson M.M. (1979). *Trichinella spiralis*: immunity and inflammation in the expulsion of transplanted adult worms from mice. *Exp. Parasitol.* 48, 305-312.
- Walker C., Bauer W., Braun R.K., Menz G., Braun P., Schwarz F., Hansel T.T. and Villiger B. (1994). Activated T cells and cytokines in bronchoalveolar lavages from patients with various lung diseases associated with eosinophilia. *Am. J. Respir. Crit. Care Med.* 150, 1038-1048.
- Wang C.H. and Bell R.G. (1992). Characterization of cellular and molecular immune effectors against *Trichinella spiralis* newborn larvae *in vivo*. *Cell. Mol. Biol.* 38, 311-325.
- Wang C.H., Korenaga M., Greenwood A. and Bell R.G. (1990). T-helper subset function in the gut of rats: differential stimulation of eosinophils, mucosal mast cells and antibody-forming cells by OX8-OX22- and OX8 - OX22+ cells. *Immunology* 71, 166-175.
- Wardlaw A.J., Dunnette S., Gleich G.J., Collins J.V. and Kay A.B. (1988). Eosinophils and mast cells in bronchoalveolar lavage in subjects with mild asthma. Relationship to bronchial hyperreactivity. *Am. Rev. Respir. Dis.* 137, 62-69.
- Warren K.S., Bundy D.A.P., Anderson R.M., Davies A.R., Henderson D.A., Jamison D.T., Prescott N. and Senft A (1990). Helminth infections. In: Disease and disease control in developing countries, Chap 15. Jamison D.T. and Mosely W.H. (eds). World Bank, Washington, DC. pp 260-276.
- Watanabe N., Katakura K., Kobayashi A., Okumura K. and Ovary Z. (1988). Protective immunity and eosinophilia in IgE-deficient SJA/9 mice infected with *Nippostrongylus brasiliensis* and *Trichinella spiralis*. *Proc. Natl. Acad. Sci. USA* 85, 4460-4462.
- Watanabe N., Nawa Y., Okamoto K. and Kobayashi A. (1994). Expulsion of *Hymenolepis nana* from mice with congenital deficiencies of IgE production or of mast cell development. *Parasite Immunol.* 16, 137-144.
- Weg V.B., Williams T.J., Lobb R.R. and Nourshargh S. (1993). A monoclonal antibody recognizing very late activation antigen-4 inhibits eosinophil accumulation *in vivo*. *J. Exp. Med.* 177, 561-566.
- Wegner C.D., Gundel R.H., Reilly P., Haynes N., Letts L.G. and Rothlein R. (1990). Intercellular adhesion molecule-1 (ICAM-1) in the pathogenesis of asthma. *Science* 247, 456-459.
- Weller P.F. (1991). The immunobiology of eosinophils. *N. Engl. J. Med.* 324, 1110-1118.
- Weller P.F. and Lim K. (1997). Human eosinophil-lymphocyte interactions. *Mem. Inst. Oswaldo Cruz (Suppl II)*, 173-182.
- Wilder J.A., Collie D.D., Wilson B.S., Bice D.E., Richard Lyons C. and Lipscomb M.F. (1999). Dissociation of airway hyperresponsiveness from immunoglobulin E and airway eosinophilia in a murine model of allergic asthma. *Am. J. Respir. Cell. Mol. Biol.* 20, 1326-1334.
- Witte O.N. (1990). Steel locus defines new multipotent growth factor. *Cell* 63, 5-6.

- Wolyniec W.W., De Sanctis G.T., Nabozny G., Torcellini C., Haynes N., Joetham A., Gelfand E.W., Drazen J.M. and Noonan T.C. (1998). Reduction of antigen-induced airway hyperreactivity and eosinophilia in ICAM-1-deficient mice. *Am. J. Respir. Cell. Mol. Biol.* 18, 777-785.
- Yagi T., Sato A., Hayakawa H. and Ide K. (1997). Failure of aged rats to accumulate eosinophils in allergic inflammation of the airway. *J. Allergy Clin. Immunol.* 99, 38-47.
- Yamaguchi Y., Suda T., Ohta S., Tominaga K., Miura Y. and Kasahara T. (1991). Analysis of the survival of mature human eosinophils: interleukin-5 prevents apoptosis in mature human eosinophils. *Blood* 78, 2542-2547.
- Yang Y., Loy J., Ryseck R.P., Carrasco D. and Bravo R. (1998). Antigen-induced eosinophilic lung inflammation develops in mice deficient in chemokine eotaxin. *Blood* 92, 3912-3923.
- Ying S., Durham S.R., Barkans J., Masuyama K., Jacobson M., Rak S., Lowhagen O., Moqbel R., Kay A.B. and Hamid Q.A. (1993). T cells are the principal source of interleukin-5 mRNA in allergen-induced rhinitis. *Am. J. Respir. Cell. Mol. Biol.* 9, 356-60.
- Yoshida T., Ikuta K., Sugaya H., Maki K., Takagi M., Kanazawa H., Sunaga S., Kinashi T., Yoshimura K., Miyazaki J., Takaki S. and Takatsu K. (1996). Defective B-1 cell development and impaired immunity against *Angiostrongylus cantonensis* in IL-5R alpha-deficient mice. *Immunity* 4, 483-494.
- Yousefi S., Hoessli D.C., Blaser K., Mills G.B. and Simon H.U. (1996). Requirement of Lyn and Syk tyrosine kinases for the prevention of apoptosis by cytokines in human eosinophils. *J. Exp. Med.* 183, 1407-1414.

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