

# Galectin-7: will the lectin's activity establish clinical correlations in head and neck squamous cell and basal cell carcinomas?

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**Summary.** The human lectin galectin-7 (Gal-7; p53-induced gene-1) has anti- and pro-malignant features in different *in vitro* models. We tried to clarify relation of its expression to cellular and clinical parameters in head and neck squamous and basal cell carcinomas. Using a non-cross-reactive antibody, immunohistochemical staining in squamous cell epithelia (epidermis, epithelium of oropharynx and larynx) (n = 57), squamous cell carcinomas (n = 47) and lymph node metastases (n = 25), as well as basal cell carcinomas (n = 10) were studied. This monitoring was flanked by processing to assess the level of differentiation (cytokeratins 10 and 14), proliferation (Ki67) and basal lamina formation (collagen IV). The results were correlated with clinical and pathological findings (grading, TNM-staging, extracapsular spread, angio- and lymphangiogenesis, perineural invasion, recurrence and survival). Gal-7 resides in all layers of epithelia with cytoplasmic and nuclear localization in normal specimens. Basal cell carcinomas were devoid of the Gal-7 respective signal. Squamous cell carcinomas were positive, presenting different staining profiles. Intense staining was predominantly found in squamous cell cancers with high degrees of differentiation and keratinization. Fittingly, poor level of differentiation (P = 0.0009), absence of keratinization (P = 0.0105) and significant discontinuity or absence of collagen IV expression in the peritumoral basal lamina (P = 0.0024)

was found in Gal-7-negative tumors. Gal-7 presence was not related to gender, primary tumor site, T-stage, N-stage, clinical stage, extracapsular spread, angio- and lymphangiogenesis, perineural spread or treatment outcome at a statistically significant level. Immunohistochemical analysis revealed a positive correlation for differentiation and keratinization to Gal-7 presence in squamous cell carcinomas. Absence of Gal-7 expression was detected in basal cell carcinomas. These clinical data delineate Gal-7 influence on differentiation *in vivo*, without evidence for a role in dissemination reported for lymphoma.

**Key words:** Carcinoma, Collagen IV, Galectin, Keratinization, Lectin

## Introduction

The malignant process is known to be associated with aberrant glycosylation. Because the emerging concept of the sugar code ascribes a role as biochemical signals to glycan epitopes of glycoconjugates from normal and tumor cells, these changes may not serve just as phenotypic markers. They also convey new properties to the cells which can be decoded by tissue lectins (Gabius, 1997a, 2006). In fact, these glycan-binding proteins are capable of "reading" even rather subtle modifications in glycan structures, such as the presence of core fucosylation or alterations in epitope density, and translate them into responses, affecting e. g. cell adhesion, growth or migration (Villalobo et al., 2006; Wu et al., 2006; André et al., 2007a). Homing especially

in on spatially accessible branch-end  $\beta$ -galactosides, the members of the galectin family belong to these endogenous effectors (Kasai and Hirabayashi, 1996; Gabius, 1997b; Cooper, 2002). The recent finding that a tumor suppressor modulates in an orchestrated manner the expression of a galectin and glycan tailoring of its ligands for acquisition of susceptibility to anoikis underscores the effectiveness of such interactions in tumor biology (André et al., 2007b). In addition to sensing changes in glycan profiles, these endogenous lectins are known to exert activities also in the cytoplasm and nucleus by virtue of peptide binding, for example, regulating transcriptional activity, transformation or apoptosis (Rotblat et al., 2004; Wang et al., 2004; Smetana et al., 2005). This background explains our interest in galectins and their presence in tumors.

The homodimeric galectin-7 (Gal-7) was initially detected in studies aimed at identifying markers associated with the normal keratinocyte phenotype, its expression was sensitive to SV40 transformation and linked to p53-related induction of apoptosis in epidermis and human DLD-1 colon carcinoma cells (Madsen et al., 1995; Magnaldo et al., 1995; Polyak et al., 1997; Bernerd et al., 1999; Saussez and Kiss, 2006). *In vitro*, the lectin inhibits growth of neuroblastoma cells and induces apoptosis in activated T cells, but, in stark contrast, is associated with an aggressive phenotype in murine 164T2 lymphoma, characterized by increased matrix metalloproteinase-9 expression (Kopitz et al., 2003; Moisan et al., 2003; Sturm et al., 2004; Demers et al., 2007). Proteomic profiling raised evidence for a relation to differentiation in bladder squamous cell carcinomas and, conversely, tumorigenesis in buccal squamous cell cancer (Østergaard et al., 1997; Chen et al., 2004). Differential display of mRNA populations to chemical carcinogenesis was described in rat mammary gland but not colon (Lu et al., 1997). Thus, Gal-7 activities and relations to disease progression in models appear to indicate contextual functionality, making predictions for clinical correlations on the basis of *in vitro* data difficult. In view of the same aim, the immunohistochemical analysis of the anti-apoptotic Gal-3 in breast cancer has recently revealed that *in vitro* activities cannot simply be extrapolated to the clinical situation (Moisa et al., 2007). The question is thus open to define associations between lectin expression and clinical parameters in tumor specimens. Toward this end, we analyzed head and neck squamous cell and basal cell carcinomas, using an antibody preparation non-cross-reactive to other members of the galectin family.

## Materials and methods

### Tissue processing

Samples of human tissue were obtained with the explicit informed consent of patients according to the Helsinki Declaration during surgical procedures for head

and neck squamous cell carcinomas (Department of Otorhinolaryngology and Head and Neck Surgery, First Faculty of Medicine and Faculty Hospital Motol, Charles University in Prague) and basal cell carcinomas (Department of Dermatovenerology, First Faculty of Medicine and General Teaching Hospital, Charles University in Prague). Each sample was divided to two parts. First part was routinely embedded to paraffin and used for histopathologic inspection, second was prepared for preparation of frozen sections. These samples (Table 1) were frozen in liquid nitrogen using Tissue-Tek (Sakura-Finetek Europe B.V., Zoeterwoude, The Netherlands). 7- $\mu$ m-thick frozen sections were prepared by a Cryocut-E microtome (Reichert-Jung, Vienna, Austria). Tissue-Tek was removed by rinsing in phosphate-buffered saline (pH 7.2; PBS) immediately before starting immunohistochemical processing. Sections were routinely fixed with 2% (w/v) paraformaldehyde in PBS. Carbohydrate-free bovine serum albumin (BSA; Sigma, Prague, Czech Republic) was used to block non-specific protein-protein interactions.

Sections from paraffin embedded tissue were routinely analyzed after the staining by hematoxylin and eosin. Parameters such as differentiation grading, extracapsular spread, angio- and lymphangiogenesis, perineural invasion were evaluated as described (Bryne et al., 1989; Ravasz et al., 1993).

### Immunohistochemical processing

After recombinant production Gal-7 was purified using affinity chromatography as a crucial step, and purity was ascertained by one- and two-dimensional gel electrophoresis, gel filtration and mass spectrometry (Kopitz et al., 2003; André et al., 2004). The quality-controlled protein was used as antigen in rabbits, and the resulting polyclonal antibodies were thoroughly checked for any cross-reactivity against other members of the galectin family by Western blotting and ELISA, especially the proto-type proteins, then removing any traces by chromatographic affinity depletion (Kayser et al., 2003; Lohr et al., 2007). Double labeling using commercial antibodies was performed to characterize

**Table 1.** Number of tissue samples.

| Tissue   | Number of donors |
|--|------------------|
| Human epidermis  | 10               |
| Basal cell carcinomas  | 10               |
| Squamous cell epithelia (oral cavity, oropharynx, larynx, hypopharynx)                           | 47               |
| Primary squamous cell cancer (oral cavity, oropharynx, larynx, hypopharynx)                      | 47               |
| Lymph node metastases of squamous cell carcinomas (oral cavity, oropharynx, larynx, hypopharynx) | 25               |

cell characteristics. Cytokeratins 10 and 14 were detected by mouse monoclonal antibodies (Dako, Brno, Czech Republic; SIGMA, Prague, Czech Republic), as were the nuclear Ki67 antigen of proliferating cells (Dako, Prague, Czech Republic) and collagen IV (Sigma, Prague, Czech Republic). Commercially available ExtrAvidin-tetramethylrhodamine isothiocyanate (TRITC) (Sigma, Prague, Czech Republic) and fluorescein isothiocyanate (FITC)-labeled swine-anti mouse and swine-anti rabbit immunoglobulins (SwAM-FITC, SwAR-FITC, ALSEVA, Prague, Czech Republic; GoAM-TRITC: Sigma, Prague, Czech Republic) were used as second-step reagents. To exclude a false-positive reaction by non-specific binding of immunoglobulins via Fc receptors, an antibody specific for CD1a (Immunotech, Prague, Czech Republic) not present on epithelial cells was tested in parallel. This reagent replaced the first-step markers during routine processing in a control section. Finally, the specimens were mounted to Vectashield (Vector Laboratories, Burlingame, CA, USA.) and then visually inspected and analyzed using an Eclipse 90i fluorescence microscope (Nikon, Prague, Czech Republic) equipped with respective filter blocks, a high-resolution CCD camera (Cool-1300Q; Vossküller, Osnabrück, Germany) and a computer-assisted image analyser (LUCIA 5.10) (Laboratory Imaging, Prague, Czech Republic). All sections were carefully examined by two independent observers, who were completely blinded with respect to clinical features of the patients. In each case, at least 500 cells within randomly selected and defined area sections on each slide were counted. For statistical analysis, cut-off points were chosen to classify tumors to be intensely or faintly positive for Gal-7 staining. A cut-off point of lower 1/3 of the intensity profile value (arbitrary units) between intensity of the background signal and the intensity profile value in corresponding non-malignant control epithelia within the tumor cell population was arbitrarily set to determine the range of faintly positive cells. Cut-offs were defined prior to relating clinical parameters to results of histochemical staining. Ki67-positive cells were counted per 1000 cells, in this way percentages of Ki67-positive cells in different samples were determined. Mean average of Ki67-positive cells for tumors with identical Gal-7 cytoplasmatic staining profiles were calculated.

#### *Statistical analysis*

The Chi-squared test was used to set Gal-7 parameters in relation to the different clinicopathological parameters, except for the Ki67 status treated with the Mann-Whitney U test. Overall survival and disease-free survival were calculated using the standard method, data sets being analyzed by using the Gehan-generalised Wilcoxon test. Statistica 6.0 software (StatSoft, Prague, Czech Republic) was run in all statistical analyses. Overall survival was computed from the date of surgery to the documented date of the last follow-up or death,

whereas disease-free survival was considered to cover the period from the date of surgery to the date of recurrence.

## **Results**

### *Normal epithelia in situ*

Application of the non-cross-reactive anti-Gal-7 antibody preparation to fixed sections of squamous cell epithelia of the epidermis and mucosal coverings (oral cavity, oropharynx, larynx and hypopharynx) detected lectin presence from the basal region to the most superficial layer (Fig. 1). Cytoplasmic and also nuclear presence, the latter most prominently in nucleoli, were seen in both basal and suprabasal layers. These observations extend the evidence for nuclear presence of galectins from proto-type Gal-1 and chimera-type Gal-3 (Smetana et al., 2005) and, most recently, Gal-2 (Dvořánková et al., 2008). As a measure of cell differentiation, cytokeratin-14 was present in cells of the basal layer in epithelia of all specimens, whereas cytokeratin-10 was encountered in keratinized epithelia of epidermis and tongue only. The typical nuclear expression of the proliferation marker Ki67 was observed in the basal layer and to a restricted extent in the surrounding suprabasal layers. A subpopulation of Ki67-positive cells represented about 5% of the cells in the basal layer. As assessed by monitoring collagen IV presence, the basal lamina was well established and continuous in the studied epithelia.

### *Basal cell carcinomas*

As also shown in Fig. 1, a qualitative difference was seen for negative tumor cells compared to the positive surrounding non-transformed epithelium. Cytokeratin-10 was not detected in studied tumors, and the presence of nuclear Ki67 antigen was observed in about 5-15% of cells, predominantly in peripheral parts of tumor nodules. Continuous collagen IV staining appeared around the studied tumors (not shown).

### *Squamous cell carcinomas*

The staining profile for Gal-7 was not uniform in the different specimens of primary and metastatic squamous cell carcinomas. Four different patterns could be discerned (Fig. 1). Intense staining with homogeneous distribution in all tumor cells (intense and homogeneous) was present in 32% of primary tumors and 12% of regional lymph node metastases. Gal-7 presence confined to the central parts of the tumor, mostly to regions of formation of keratin pearls (intense heterogeneous pattern), was observed in 25.5% of primary tumors and 32% of regional lymph node metastases. Faint but homogeneous staining throughout the entire tumor cell population applied to 25.5% of the primary tumors and 20% of the regional lymph node

metastases. 17% of primary tumors and 36% of regional lymph node metastases did not show immunohistochemical positivity. A clear-cut difference between primary tumors and corresponding lymph node metastases could not reliably be described. Nuclear positivity of tumor cells concerned only cases with intense staining, irrespective of presenting homogeneous or heterogeneous profiles. Regarding the markers for cytodifferentiation, cytokeratin-14 was spotted in all primary carcinomas and regional lymph node metastases, cytokeratin-10 in keratinized carcinomas only (Fig. 2). Gal-7 presence correlated with keratinization ( $P = 0.0105$ ). Ki67-expressing cells were in the peripheral regions of tumor nodules. Studied tumors differed among each other in the size of the Ki67-positive cell population from 5 to 60% (Fig. 2, Table 2). There was no apparent correlation of the

proliferation status with Gal-7 presence ( $P = 0.1376$ ). In general, peritumoral basement membranes were covered with a continuous layer of collagen IV in the tumors with intense and homogeneous staining for Gal-7. The other Gal-7 staining profiles showed variability for the appearance of the collagen IV layer, ranging from major defects to even complete absence in tumors lacking Gal-7 ( $P = 0.0024$ ) (Fig. 2, Table 2). In contrast, intense and homogeneous Gal-7 staining correlated with the level of differentiation (grading) ( $P = 0.0009$ ). When Gal-7 staining was set in relation with other factors, i.e. gender ( $P = 0.3781$ ), primary tumor site ( $P = 0.2703$ ), T-stage ( $P = 0.6222$ ), N-stage ( $P = 0.1065$ ), clinical stage ( $P = 0.5127$ ), extracapsular spread ( $P = 0.5998$ ), angioinvasion ( $P = 0.6443$ ) and lymphangiogenesis ( $P = 0.3781$ ), perineural spread ( $P = 0.1306$ ) and treatment outcome, no statistically significant association turned

**Table 2.** Comparison of clinical and histopathological parameters with Gal-7 presence.

| PRIMARY CARCINOMAS              |                        | INTENSE SIGNAL<br>27 | FAINT/NO SIGNAL<br>20 | P value |
|---------------------------------|------------------------|----------------------|-----------------------|---------|
| Site                            | larynx/hypopharynx     | 13                   | 7                     | 0.2703  |
|                                 | oropharynx/oral cavity | 14                   | 13                    |         |
| Gender                          | male                   | 24                   | 16                    | 0.3781  |
|                                 | female                 | 3                    | 4                     |         |
| T-stage                         | T 1+2                  | 8                    | 9                     | 0.6222  |
|                                 | T 3+4                  | 19                   | 11                    |         |
| N-stage                         | N 0                    | 5                    | 5                     | 0.1065  |
|                                 | N 1-3                  | 22                   | 15                    |         |
| Clinical stage                  | CS 1+2                 | 1                    | 1                     | 0.5127  |
|                                 | CS 3+4                 | 26                   | 19                    |         |
| Grading                         | G1+G2                  | 22                   | 8                     | 0.0009  |
|                                 | G3+G4                  | 5                    | 12                    |         |
| Keratinization                  | keratinized            | 15                   | 4                     | 0.0105  |
|                                 | non-keratinized        | 12                   | 16                    |         |
| Extracapsular spread (ECS)      | ECS -                  | 21                   | 17                    | 0.5998  |
|                                 | ECS +                  | 6                    | 3                     |         |
| Lymphangiogenesis               | lymphangiogenesis -    | 22                   | 18                    | 0.3781  |
|                                 | lymphangiogenesis +    | 5                    | 2                     |         |
| Angioinvasion                   | angioinvasion -        | 22                   | 16                    | 0.6443  |
|                                 | angioinvasion +        | 5                    | 4                     |         |
| Perineural spread               | perin. spr. -          | 26                   | 18                    | 0.1306  |
|                                 | perin. spr. +          | 1                    | 2                     |         |
| Outcome                         | local recidive         | 2                    | 2                     | 0.1376  |
|                                 | regional recidive      | 3                    | 2                     |         |
|                                 | distant metastases     | 3                    | 2                     |         |
|                                 | no evidence of disease | 20                   | 16                    |         |
| Basal lamina (Col IV) formation | well surrounded        | 7                    | 4                     | 0.0024  |
|                                 | poorly formed          | 19                   | 16                    |         |
| Ki 67-positive population       | mean                   | 30%                  | 38%                   |         |

Clinical and histopathological parameters (TNM staging, extracapsular spread, grading, keratinization, angio- and lymphangiogenesis, perineural spread), pattern of basal lamina (collagen IV) formation and proliferation (Ki-67) vs. Gal-7 presence in primary head and neck squamous cell carcinomas. Intense signal corresponds to tumors with either intense homogeneous or intense inhomogeneous Gal-7-dependent staining. Faint/No signal corresponds to tumors with either weak homogeneous staining or no detectable Gal-7. Mean average of Ki-67-positive cells counted per 1000 tumor cells for tumors with identical staining pattern were calculated.



up (Table 2). Also, no statistically significant differences of survival among the studied patient groups suffering from squamous cell carcinoma were observed relative to Gal-7 (Fig. 3).

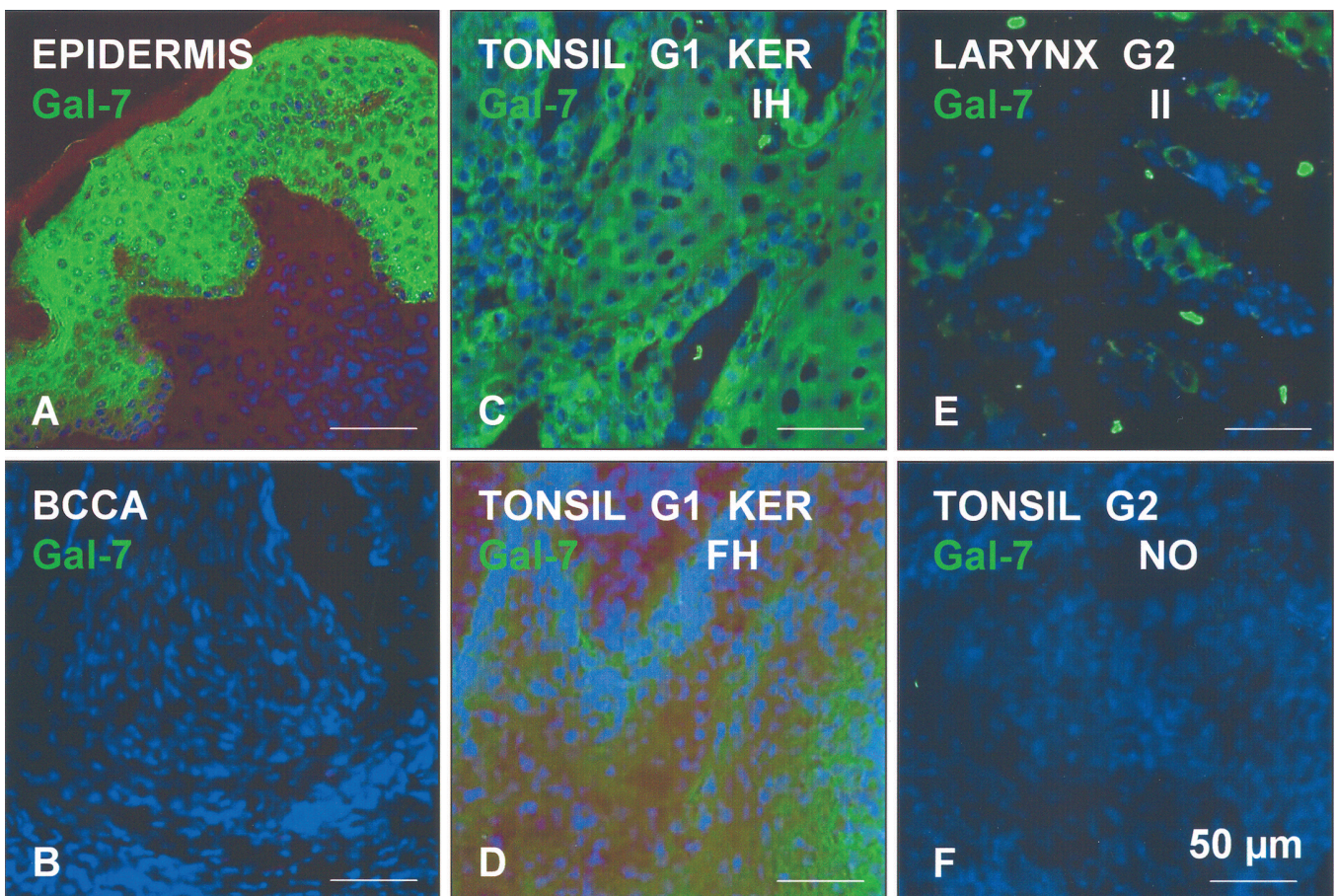
## Discussion

Using stratification, a prognostic correlation has been reported for stage IV hypopharyngeal squamous cell carcinoma patients. In this group, Gal-7 therefore has the potential to identify patients at risk of recurrence and with dismal prognosis (Saussez et al., 2006). Of note, staining characteristics for Gal-7 differed from those of Gal-1 belonging to the same subgroup, a strong argument for non-overlapping functionalities in the galectin network (Saussez et al., 2008). Also, associated with a feature of tumor progression, Gal-7 presence was correlated to muscle-infiltrating growth in urothelial cancer (Langbein et al., 2007), whereas Gal-7 monitoring in progression of thyroid cancer appeared to reflect a dual role, with anti- and promalignant features

at different stages (Rorive et al., 2002). These results, revealing a tumor-type- and also stage-of-tumorigenesis-related activity profile in the case of Gal-7, have a bearing on considerations to devise new treatment modalities based on modulating endogenous galectin expression.

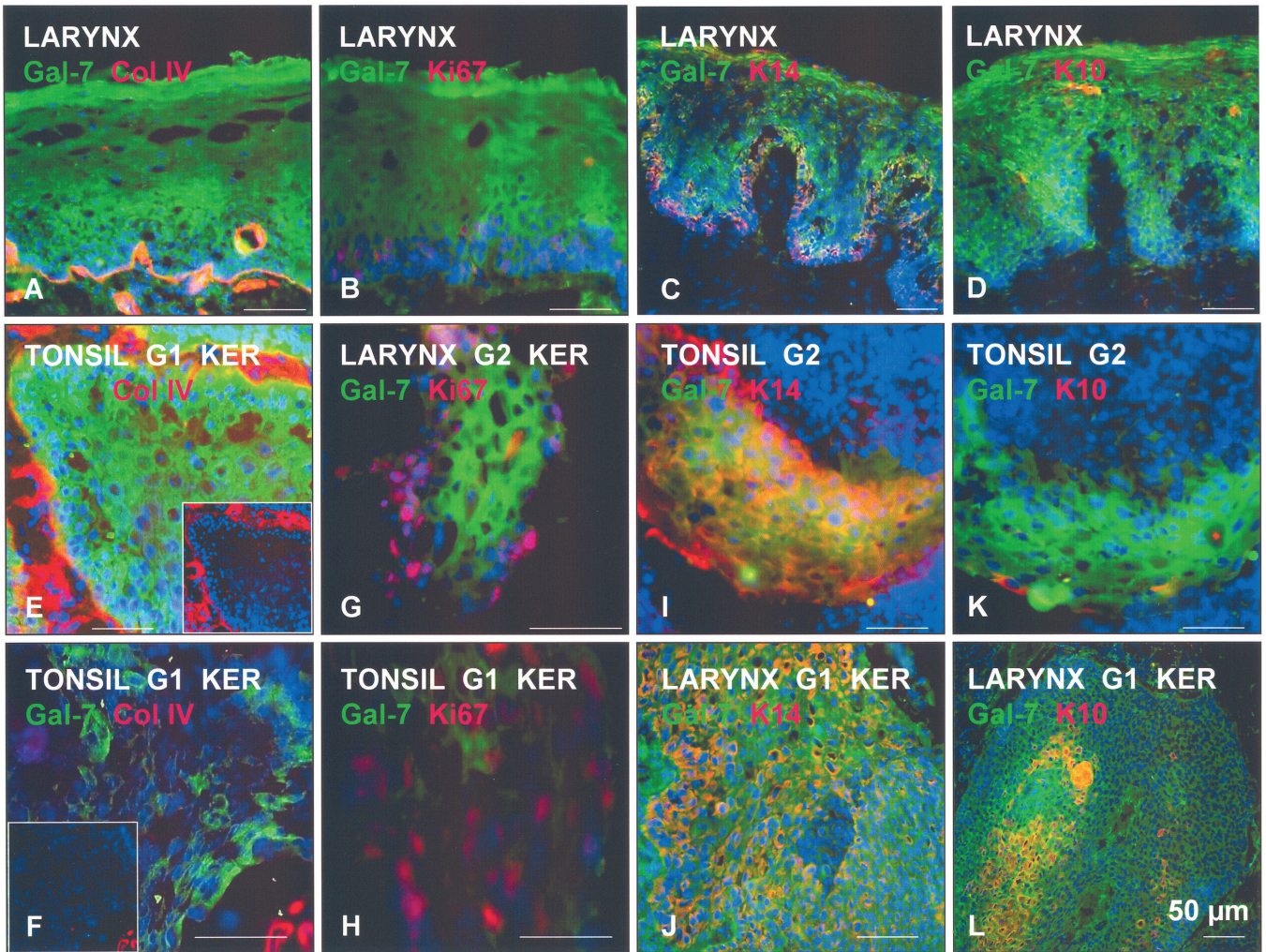
Absence of the signal for Gal-7 in basal cell carcinoma was also observed earlier (Magnaldo et al., 1998; Chovanec et al., 2005). Explanation of this phenomenon is only hypothetical but it can be related to the low level of differentiation of tumor epithelial cells. They express keratin 19, marker typical for epidermal stem cells and  $\alpha$ 2,6-linked sialic acid, marker of poorly differentiated epithelial cells (Holíková et al., 2002; Dvořánková et al., 2005). In harmony with these observations, cells of basal cell carcinoma were never recognized by labeled galectin-3, feature typical for suprabasal cells of squamous epithelia (Plzák et al., 2001).

Following its description as a marker associated with the normal keratinocyte phenotype and as p53-induced

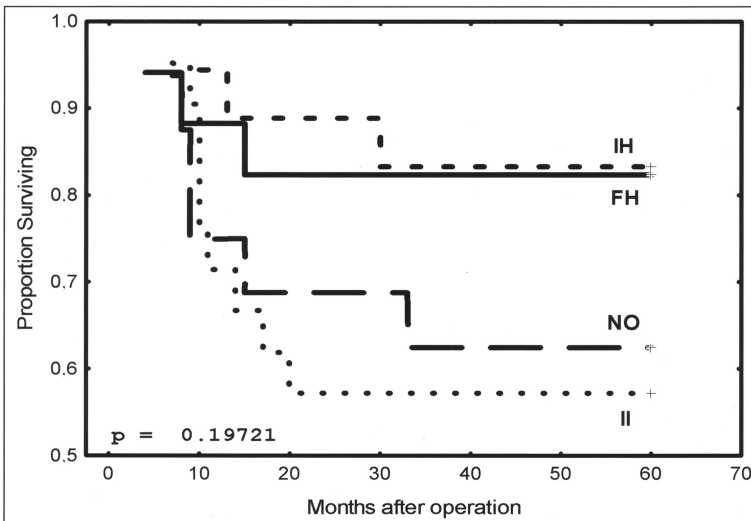


**Fig. 1.** Immunohistochemical detection of Gal-7 in epidermis (A), basal cell carcinoma (B) and head and neck squamous cell carcinoma (C: intense and homogeneous/IH, D: faint and homogeneous/FH, E: intense and inhomogeneous/II, F: no expression/NO). Grading (G1-G3), keratinized (KER). Nuclei were counterstained by DAPI.





**Fig. 2.** Immunohistochemical detection of Gal-7, a marker of proliferation (Ki67), collagen IV (basement membrane) and keratins (K10, K14) in squamous cell epithelia (A-D) and head and neck squamous cell carcinomas (E- L). Grading (G1-G3), keratinized (KER). Nuclei were counterstained by DAPI.



**Fig. 3.** Kaplan-Meier graph of overall survival of patients suffering from head and neck squamous carcinoma and Gal-7-associated parameters. Phenotype of Gal-7 localization: IH: intense and homogeneous, FH: faint and homogeneous, II: intense and inhomogeneous, NO: no expression.

gene product in DLD-1 colon cancer cells, cell biological data had indicated differential regulation of Gal-7 in squamous cell carcinomas of different origin, a differential response to chemical carcinogenesis in rat models, and anti- or pro-malignancy activities in different human tumor models. To decide on clinical correlations in patient material we studied tissue sections immunohistochemically and disclosed a correlation to increased status of differentiation and keratinization in head and neck squamous cell carcinomas.

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## References

- André S., Kaltner H., Furuike T., Nishimura S.I. and Gabius H.J. (2004). Persubstituted cyclodextrin-based glycoclusters as inhibitors of protein-carbohydrate recognition using purified plant and mammalian lectins and wild-type and lectin-gene-transfected tumor cells as targets. *Bioconjugate Chem.* 15, 87-98.
- André S., Kořár T., Schubert R., Unverzagt C., Kojima S. and Gabius H.J. (2007a). Substitutions in the N glycan core as regulators of biorecognition: the case of core-fucose and bisecting GlcNAc moieties. *Biochemistry* 46, 6984-6995.
- André S., Sanchez-Ruderisch H., Nakagawa H., Buchholz M., Kopitz J., Forberich P., Kemmner W., Böck C., Deguchi K., Detjen K.M., Wiedenmann B., von Knebel Doeberitz M., Gress T.M., Nishimura S.I., Rosewicz S. and Gabius H.J. (2007b). Tumor suppressor p16<sup>INK4a</sup> - modulator of glycomic profile and galectin-1 expression to increase susceptibility to carbohydrate-dependent induction of anoikis in pancreatic carcinoma cells. *FEBS J.* 274, 3233-3256.
- Bernernd F., Sarasin A. and Magnaldo T. (1999). Galectin-7 overexpression is associated with the apoptotic process in UVB-induced sunburn keratinocytes. *Proc. Natl. Acad. Sci. USA* 96, 11329-11334.
- Bryne M., Koppang H.S., Lilleng R., Stene T., Bang G. and Dabelsteen E. (1989). New malignancy grading is a better prognostic indicator than Broders' grading in oral squamous cell carcinomas. *J. Oral. Pathol. Med.* 18, 432-437.
- Chen J., He Q.Y., Yuen A.P.W. and Chiu J.F. (2004). Proteomics of buccal squamous cell carcinoma: the involvement of multiple pathways in tumorigenesis. *Proteomics* 4, 2465-2475.
- Chovanec M., Smetana K. Jr., Plzák J., Betka J., Plzáková Z., Štork J., Hrdličková E., Kuwabara I., Dvořánková B., Liu F.T., Kaltner H., André S. and Gabius H.J. (2005). Detection of new diagnostic markers in pathology by focus on growth-regulatory endogenous lectins. The case study of galectin-7 in squamous epithelia. *Prague Med. Rep.* 106, 209-216.
- Cooper D.N.W. (2002). Galectinomics: finding themes in complexity. *Biochim. Biophys. Acta* 1572, 209-231.
- Demers M., Biron-Pain K., Hébert J., Lamarre A., Magnaldo T. and St-Pierre Y. (2007). Galectin-7 in lymphoma: elevated expression in human lymphoid malignancies and decreased lymphoma dissemination by antisense strategies in experimental model. *Cancer Res.* 67, 2824-2829.
- Dvořánková B., Smetana K. Jr., Chovanec M., Lacina L., Štork J., Plzáková Z., Galovičová M. and Gabius H.J. (2005). Transient expression of keratin K19 is induced in originally negative interfollicular epidermal cells by adhesion of suspended cells. *Int. J. Mol. Med.* 16, 525-531.
- Dvořánková B., Lacina L., Smetana K. Jr., Lensch M., Manning J.C., André S. and Gabius H.J. (2008). Human galectin-2: nuclear presence in vitro and its modulation by quiescence/stress factors. *Histol. Histopathol.* 23, 167-178.
- Gabius H.J. (1997a). Concepts of tumor lectinology. *Cancer. Invest.* 15, 454-464.
- Gabius H.J. (1997b). Animal lectins. *Eur. J. Biochem.* 243, 543-576.
- Gabius H.J. (2006). Cell surface glycans: the why and how of their functionality as biochemical signals in lectin-mediated information transfer. *Crit. Rev. Immunol.* 26, 43-79.
- Holíčková Z., Hrdličková-Cela E., Plzák J., Smetana K. Jr., Betka J., Dvořánková B., Esner M., Wasano K., André S., Kaltner H., Motlík J., Hercogová J., Kodet R. and Gabius H.-J. (2002). Defining the glycophenotype of squamous epithelia by plant and mammalian lectins. Differentiation-dependent expression of -2,6- and -2,3-linked N-acetylneuraminic acid in squamous epithelia and carcinomas and its differential effect on binding of the endogenous lectins galectins-1 and -3. *APMIS.* 110, 845-856.
- Kasai K.I. and Hirabayashi J. (1996). Galectins: a family of animal lectins that decipher glyco-codes. *J. Biochem.* 119, 1-8.
- Kayser K., Hoefft D., Hufnagl P., Caselitz J., Zick Y., André S., Kaltner H. and Gabius H.J. (2003). Combined analysis of tumor growth pattern and expression of endogenous lectins as prognostic tool in primary testicular cancer and its lung metastases. *Histol. Histopathol.* 18, 771-779.
- Kopitz J., André S., von Reitzenstein C., Versluis K., Kaltner H., Pieters R.J., Wasano K., Kuwabara I., Liu F.T., Cantz M., Heck A.J.R. and Gabius H.J. (2003). Homodimeric galectin-7 (p53 induced gene 1) is a negative growth regulator for human neuroblastoma cells. *Oncogene* 22, 6277-6288.
- Langbein S., Brade J., Badawi J.K., Hatzinger M., Kaltner H., Lensch M., Specht K., André S., Brinck U., Alken P. and Gabius H.J. (2007). Gene-expression signature of adhesion/growth-regulatory tissue lectins (galectins) in transitional cell cancer and its prognostic relevance. *Histopathology* 51, 681-690.
- Lohr M., Lensch M., André S., Kaltner H., Siebert H.C., Smetana K. Jr., Sinowatz F. and Gabius H.J. (2007). Murine homodimeric adhesion/growth-regulatory galectins-1, -2, and -7: comparative profiling of gene/promoter sequences by database mining, of expression by RT-PCR/immunohistochemistry and of contact sites for carbohydrate ligands by computational chemistry. *Folia Biol. (Praha).* 53, 109-128.
- Lu J., Pei H., Kaeck M. and Thompson H.J. (1997). Gene expression changes associated with chemically induced rat mammary carcinogenesis. *Mol. Carcinogen.* 20, 204-215.
- Madsen P., Rasmussen H., Flint T., Gromov P., Kruse A., Honore B., Vorum H. and Celis J. (1995). Cloning, expression, and chromosome mapping of human galectin-7. *J. Biol. Chem.* 270, 5823-5829.



- Magnaldo T., Bernerd F. and Darmon M. (1995). Galectin-7, a human 14-kDa S-lectin, specifically expressed in keratinocytes and sensitive to retinoic acid. *Dev. Biol.* 168, 259-271.
- Magnaldo T., Fowles D. and Darmon M. (1998). Galectin-7, a marker of all types of stratified epithelia. *Differentiation* 63, 159-168.
- Moisa A., Fritz P., Eck A., Wehner H.D., Mürdter T., Simon W. and Gabius H.J. (2007). Growth/adhesion-regulatory tissue lectin galectin-3: stromal presence but not cytoplasmic/nuclear expression in tumor cells as negative prognostic factor in breast cancer. *Anticancer Res.* 27, 2131-2140.
- Moisan S., Demers M., Mercier J., Magnaldo T., Potworowski E.F. and St-Pierre Y. (2003). Upregulation of galectin-7 in murine lymphoma cells is associated with progression toward an aggressive phenotype. *Leukemia* 17, 751-759.
- Østergaard M., Rasmussen H.H., Nielsen H.V., Vorum H., Ørntoft T.F., Wolf H. and Celis J.E. (1997). Proteome profiling of bladder squamous cell carcinomas: identification of markers that define their degree of differentiation. *Cancer Res.* 57, 4111-4117.
- Plzák J., Smetana K. Jr., Hrdliáková E., Kodet R., Holíková Z., Liu F.T., Dvořáková B., Kaltner H., Betka J. and Gabius H.J. (2001). Expression of galectin-3-reactive ligands in squamous cancer and normal epithelial cells as a marker of differentiation. *Int. J. Oncol.* 19, 59-64.
- Polyak K., Xia Y., Zweier J.L., Kinzler K.W. and Vogelstein B. (1997). A model for p53-induced apoptosis. *Nature* 389, 300-305.
- Ravasz L.A., Hordijk G.J., Slootweg P.J., Smit F. and Tweel I.V. (1993). Uni- and multivariate analysis of eight indications for post-operative radiotherapy and their significance for local-regional cure in advanced head and neck cancer: *J. Laryngol. Otol.* 107, 437-440.
- Rorive S., Eddafali B., Fernandez S., Decaestecker C., André S., Kaltner H., Kuwabara I., Liu F.T., Gabius H.J., Kiss R. and Salmon I. (2002). Changes in galectin-7 and cytokeratin-19 expression during the progression of malignancy in thyroid tumors: diagnostic and biological implications. *Mod. Pathol.* 15, 1294-1301.
- Rotblat B., Niv H., André S., Kaltner H., Gabius H.J. and Kloog Y. (2004). Galectin-1(L11A) predicted from a computed galectin-1 farnesyl-binding pocket selectively inhibits Ras-GTP. *Cancer Res.* 64, 3112-3118.
- Saussez S., Cucu D.R., Decaestecker C., Chevalier D., Kaltner H., André S., Wacreniez A., Toubeau G., Camby I., Gabius H.J. and Kiss R. (2006). Galectin 7 (p53-induced gene 1): a new prognostic predictor of recurrence and survival in stage IV hypopharyngeal cancer. *Ann. Surg. Oncol.* 13, 999-1009.
- Saussez S. and Kiss R. (2006). Galectin-7. *Cell. Mol. Life Sci.* 63, 687-697.
- Saussez S., Lorfèvre F., Decaestecker C., Chevalier D., Kaltner H., André S., Toubeau G., Gabius H.J. and Leroy X. (2008). Increased expression and altered intracellular distribution of adhesion/growth-regulatory lectins galectins-1 and -7 during tumour progression in hypopharyngeal and laryngeal squamous cell carcinomas. *Histopathology* 52, 483-493.
- Smetana K. Jr., Dvořáková B., Chovanec M., Bouáek J., Klíma J., Motlík J., Lensch M., Kaltner H., André S. and Gabius H.J. (2005). Nuclear presence of adhesion/growth-regulatory galectins in normal/malignant cells of squamous epithelial origin. *Histochem. Cell Biol.* 125, 171-182.
- Sturm A., Lensch M., André S., Kaltner H., Wiedenmann B., Rosewicz S., Dignass A.U. and Gabius H.J. (2004). Human galectin-2: novel inducer of T cell apoptosis with distinct profile of caspase activation. *J. Immunol.* 173, 3825-3837.
- Villalobo A., Nogales-González A. and Gabius H.J. (2006). A guide to signaling pathways connecting protein-glycan interaction with the emerging versatile effector functionality of mammalian lectins. *Trends Glycosci. Glycotechnol.* 18, 1-37.
- Wang J.L., Gray R.M., Haudek K.C. and Patterson R.J. (2004). Nucleocytoplasmic lectins. *Biochim. Biophys. Acta* 1673, 75-93.
- Wu A.M., Singh T., Wu J.H., Lensch M., André S. and Gabius H.J. (2006). Interaction profile of galectin-5 with free saccharides and mammalian glycoproteins: probing its fine specificity and the effect of naturally clustered ligand presentation. *Glycobiology* 16, 524-537.

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