1	Gender differences in the renal changes induced by a prolonged high fat diet in rats with
2	altered renal development.
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#### Abstract 34

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36

The mechanisms involved in renal dysfunction induced by high fat diet (HFD) in subjects with altered renal development (ARDev) are understudied. The objective of this study is to

examine whether there are sex-dependent differences in the mechanisms involved in the 37

hypertension and deterioration of renal function in SD rats with prolonged HFD and ARDev. 38

39 The role of angiotensin II (Ang II) in the arterial pressure (AP) increments, the renal

hemodynamic sensitivity to Ang II, glomerular damage and changes in fat abdominal volume, 40

plasma adipokines levels, renal NADPHp67phox expression and renal infiltration of immune 41

cells were examined. Hypertension and deterioration of renal function were enhanced 42

(P < 0.05) in both sexes of rats with HFD and ARDev. The decrease (P < 0.05) of AP elicited by 43

candesartan in hypertensive rats was similar to that induced by the simultaneous 44

administration of candesartan and apocynin. The greater (P<0.05) renal vasoconstriction 45

induced by Ang II in both sexes of rats with HFD and ARDev, was accompanied by an 46

47 enhanced (P<0.05) infiltration of CD-3 cells and macrophages in the renal cortex and renal

medulla. The increments (P<0.05) in the renal expression of NADPHp67phox and 48

glomeruloesclerosis were greater (P<0.05) in males than in females with HFD and ARDev. 49

Our results suggest that the hypertension and deterioration of renal function induced by HFD 50

in rats with ARDev are Ang II-dependent, and mediated by increments in oxidative stress and 51

immune system activation. Sex-dependent increments in oxidative stress and glomerular 52

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damage may contribute to the deterioration of renal function in these rats.
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55 Key Words: Angiotensin II, High fat diet, Immune system, Oxidative stress, Renal development, Hypertension, Renal damage. 56

# 58 Key Points

- An early and prolonged high fat diet induces a greater increment of arterial pressure in
  male than in female rats with a reduced nephron endowment.
- Renal damage is enhanced by a high fat diet in both sexes of rats with altered renal
  development.
- Angiotensin II is involved in the hypertension and deterioration of renal function in rats
  with a prolonged high fat diet and reduced nephron endowment.
- The activation of oxidative stress and inflammatory pathways is greater in male than in
- 66 female rats with a prolonged high fat diet and reduced nephron endowment.

### 68 INTRODUCTION

The mechanisms involved in the sex-dependent differences in cardiovascular and renal 69 70 alterations, as a consequence of an altered renal development (ARDev), have been examined in several models of developmental programming [7, 31]. It has also been reported that a 71 72 decreased renal reserve and hypertension enhances the possibility to have a further 73 deterioration of renal function, when exposed to a second insult such as a prolonged increment of sodium [29] or proteins [26] in the diet. However, studies evaluating whether 74 there is a sexual dimorphism in the renal function changes secondary to a prolonged exposure 75 to a high fat diet (HFD) in subjects with an ARDev are limited [10, 12]. It is also unknown to 76 what extent glomerular damage is enhanced in both sexes, and whether the mechanisms 77 involved in the arterial pressure and renal alterations, as a consequence of the prolonged HFD 78 and an ARDev, are sex-dependent. 79

The main objective of this study was to examine whether there is a sexual dimorphism in the 80 81 mechanisms involved in the hypertension and deterioration of renal function observed in rats with ARDev and a prolonged HFD from an early age. Changes in fat abdominal volume 82 (FAV), plasma adipokines levels and glomerular damage were evaluated since they may 83 contribute to hypertension and renal dysfunction [8, 20, 22, 27, 28]. The involvement of 84 angiotensin II (Ang II) was examined by evaluating both, the AP response to candesartan 85 administration and the renal hemodynamic sensitivity to Ang II. It was also examined whether 86 the Ang II effects are associated to increments in oxidative stress and in the renal infiltration 87 of T lymphocytes and macrophages. To evaluate the contribution of these mechanisms is 88 89 important considering that obesity precipitates the appearance of renal dysfunction among patients with a reduced nephron endowment [24], and that the progressive prevalence of 90 91 overweight and obesity in children and adolescents is highly correlated with an increase of 92 kidney disease [30].

### 93 MATERIAL AND METHODS

Studies were performed in Sprague-Dawley (SD) rats purchased from the Laboratory Animals 94 Service of the University of Murcia. Rats were housed in a temperature (23°C) and humidity 95 (45-60%) controlled room on a 12/12 light-dark cycle, with ad libitum access to water and 96 food. Female SD rats (250 g b.w.) were placed with males, and day 1 of pregnancy was 97 98 marked as the morning that sperm was found in the vaginal smear. On postnatal day 1, litter size was fixed (8-10 pups) to ensure similar nourishment during the suckling period. Newborn 99 rats were treated from postnatal day 1 to postnatal day 14 with vehicle (isotonic saline) or an 100 101  $AT_1$  receptor antagonist (candesartan, 7 mg·kg-1·day-1) by oral administration. Previous studies performed at four months of age have shown that this treatment induces a 36-38% 102 reduction in nephron endowment [27], the development of hypertension and a progressive 103 deterioration of renal function [25, 28, 29]. A reduced nephron endowment and a decrease in 104 105 the renin-angiotensin system (RAS) activity are two common findings in several models of 106 developmental programming of cardiovascular and renal diseases [7, 31]. 107 After weaning, male and female rats were fed with either a normal (NFD) or a high (HFD) fat diet until the end of the experimental period. The calories in the NFD (Tekland 2014, Energy 108 density: 2,9 Kcal·gr-1) are from proteins (20%), fat (soybean oil) (13%) and carbohydrate 109 (67%). The calories in the HFD (Tekland TD.06414, Energy density: 5,1 Kcal·gr-1) are from 110 proteins (18,4%), fat (lard + soybean oil) (60,3%) and carbohydrate (21,3%). NaCl content is 111 similar in both diets (NFD: 0,3%; HFD: 0,4%). Therefore, four groups of male and female 112 113 rats were included to perform each experiment: rats treated with vehicle during the first two 114 weeks and fed a NFD (NFD group); rats treated with vehicle during the first two weeks and fed a HFD (HFD group); rats with ARDev and NFD (ARDev+NFD group); and rats with 115 ARDev and exposed to a prolonged HFD from weaning (ARDev+HFD group). One male and 116 117 one female rat from each dam were included in each group to perform the next experiments.

- 118 Arterial pressure response to candesartan. The SAP response to a 3-day oral administration
- of candesartan (7 mg·kg-1·day-1) was examined at 4 months of age in each group of male
- 120 (NFD, n=6; HFD, n=8; ARDev+NFD, n=6; ARDev+HFD, n=6) and female (NFD, n=6;
- 121 HFD, n=6; ARDev+NFD, n=6; ARDev+HFD, n=7) rats. Systolic AP was measured before
- treatment and the third day of candesartan administration
- 123 <u>Arterial pressure response to apocynin and candesartan</u>. The SAP response to a 3 weeks'
- oral administration of apocynin (60 mg·kg-1·day-1 in drinking water) was examined at 14
- 125 weeks of age in each group of male (NFD, n=8; HFD, n=8; ARDev+NFD, n=8;
- 126 ARDev+HFD, n=7) and female (NFD, n=7; HFD, n=7; ARDev+NFD, n=9; ARDev+HFD,
- 127 n=9) rats. During the last three days, apocynin was simultaneously administered with
- 128 candesartan (7 mg·kg-1·day-1) to examine whether the Ang II effects on arterial pressure are
- 129 mediated by changes in oxidative stress and/or in inflammatory mediators. Systolic AP was
- 130 measured before apocynin administration (14 weeks of age), immediately before and the third
- 131 day of candesartan treatment.
- 132 Systolic AP was measured in rats by plethysmography (CODA, Kent Scientific Corporation,
- 133 CT) and under superficial anesthesia (isoflurane: 4% to induce; 2-2,5% to maintain) to avoid
- the stress during the inflation-deflation cycles in the tail. The average of at least 10
- 135 measurements was taken as SAP value. Previous studies showed that the SAP values obtained
- by plethysmography are highly correlated with those obtained in conscious freely moving ratswith intra-arterial catheters [29].
- 138 Renal hemodynamic response to acute Ang II infusion. After overnight fasting, 4 months old
- rats were instrumented to measure arterial pressure (PowerLab, ADInstruments) and to
- 140 perform renal function studies [25, 26, 28] in each group of male (NFD, n=8; HFD, n=8;
- 141 ARDev+NFD, n=7; ARDev+HFD, n=6) and female (NFD, n=8; HFD, n=6; ARDev+NFD,
- 142 n=6; ARDev+HFD, n=6) rats. Briefly, glomerular filtration rate (GFR) was calculated by the

[<sup>3</sup>H] inulin clearance. A transit-time flow probe (Transonic Systems) was implanted on the
left renal artery for renal blood flow (RBF) measurement. A 70-min stabilization period was
allowed before experiments began. Two 20-min basal clearance periods were followed by an
i.v. captopril (10 ng·kg-1·min-1) infusion that does not modify GFR but leads to an increment
of RBF [24]. An i.v. Ang II infusion (30 ng·kg-1·min-1) was started 30 min after captopril
administration began. Fifteen minutes after initiating Ang II infusion, two more 20-min

149 clearances were obtained.

150 *Fat abdominal volume, leptin and adiponectin levels, renal NADPHp67phox expression and* 

151 *immunohistopathology studies*. FAV was measured at 4 months of age as described [17], (n=8

in each experimental group). Rats were anesthetized by isoflurane (4% to induce and 2.5% to

153 maintain) for image acquisitions using the Albira CT system (Bruker Molecular Imaging,

154 Woodbridge, CT). Images were reconstructed using the filtered back projection algorithm via

the Albira Suite 5.0 Reconstructor using "standard" parameters. These settings produce a final

image with 125 μm isotropic voxels, deemed sufficient for abdominal cavity analysis.

157 Analysis was performed using the PMOD (PMOD Technologies LTD, Zurich, Switzerland)

158 software. Images were segmented in PMOD according to tissue density, first for total

abdominal volume and then for fat volume.

160 One day after FAV measurements, rats were again anesthetized (isoflurane) and blood

161 withdrawn by cardiac puncture. Plasma samples were stored at -80°C until analysis of leptin

and adiponectin concentrations. Right kidney was removed, weighed and stored at -80°C until

163 processed for Western blot analysis of NADPHp67phox expression. Left kidney was fixed in

- 164 10% neutral formalin (Panreac, Barcelona, Spain) for 24 hours, processed and paraffin-
- 165 embedded to examine the glomerular damage and for immune histopathologic analysis.
- 166 <u>Plasma *leptin*</u> and *adiponectin* concentrations were determined in duplicate by commercially
- available ELISA kits (R&D System, MN, USA) in each group of male (NFD, n=7; HFD, n=7;

- 168 ARDev+NFD, n=7; ARDev+HFD, n=7) and female (NFD, n=8; HFD, n=7; ARDev+NFD,
- 169 n=7; ARDev+HFD, n=7) rats.

Renal NADPHp67phox expression. Renal cortex and renal medulla were homogenized at 10% 170 (w/v) using a Dounce tissue grinder (Sigma, USA) in a homogenization buffer [20 mM 171 172 HEPES, pH 7.9, 1 mM MgCl2, 0.5 mM EDTA, 1 mM EGTA, 1 mM dithiothreitol (DTT), 0.5 mM phenylmethylsulphonyl fluoride (PMSF), 5 µg·ml-1 aprotinin, and 2.5 µg·ml-1 173 leupeptin] (n=5 rats per group). Homogenates were cleared by centrifugation at 1,000 g for 5 174 min at 4°C. Supernatants were removed and centrifuged at 15,000 g at 4°C for 40 min to 175 176 obtain the cytosolic fraction. Protein content was determined using the Bradford assay and protein extracts were stored at -80°C until use. Briefly, 50 µg of protein was separated on 4-177 178 20% SDS-PAGE gels (Bio-Rad) and transferred to a nitrocellulose membrane (Hybord ECL, Amersham). A mouse monoclonal antibody against NADPHp67phox (Santa Cruz 179 Biotechnology, sc-374510) and β-Actin (Sigma, A1978) were used. Bound antibody was 180 detected using a horseradish peroxidase-conjugated antibody (Santa Cruz, sc-516102). Bands 181 were visualized using an enhanced chemiluminescence system (ECL Plus, Amersham), and 182 images were captured on autoradiography film, scanned, and quantified with the Image J 183 program. Expression in male and female rats was examined on the same gels. 184 185 Glomerular damage. To examine the degree of glomerulosclerosis, 3 µm-thick sections from formalin-fixed and paraffin-embedded samples kidneys were stained with hematoxylin-eosin 186 187 (HE) and Masson's trichrome (Tric). Each glomerular profile was graded and assigned to 1 of 4 groups with respect to the degree of glomerular damage [27, 28] (6 rats per group). 188 Approximately 500 glomerular profiles were examined for each experimental group. 189 Immunohistopathology. To establish the distribution of T-cells and macrophages on the renal 190 191 cortex and renal medulla, an indirect immunohistochemical procedure was carried out on 3-192 µm-thick sections from formalin-fixed and paraffin-embedded samples (n=6 per group). After

deparaffination and rehydration, a demasking antigen procedure was performed (EDTA buffer 193 pH 9.0 at 98°C for 30 min for T-cells, and proteinase K, incubation at 37°C for 15 min for 194 195 macrophages). After peroxidase blocking, sections were incubated (4°C) overnight with the primary antibody (polyclonal rabbit anti-T CD3, dilution 1:500, Dako, A0452, Barcelona, 196 197 Spain) or monoclonal rat anti-CD68 (dilution 1:100, Merck, MAB1435, Madrid, Spain) and after that, incubated with a secondary anti-rabbit (Dako) or anti-rat (Vector IMPRESS, Vector 198 Labs., Madrid, Spain) biotynilated polymer for 20 min at 37°C. Immunoreaction was finally 199 revealed with 3,3'-diaminobencidine and counterstained with Harry's hematoxylin (Thermo 200 201 Scientific, Madrid, Spain). Positive reaction was identified by a dark-brown precipitated with membrane pattern. Average number of positive cells were determined by analysis of 10-202 random high power fields (X400) for each cell population. This analysis was performed by 203 using a Zeiss Axio Scope A10 (Carls Zeiss, Jenna, Germany) light microscope with a digital 204 205 camera and software system (AxioCam IcC3 and Axio Vision SE64, Zeiss).

# 206 Statistical analysis

Data in text, tables, and figures are given as means  $\pm$  SE. Normal distributions were tested by Shapiro-Wilk. Differences between experimental periods within one group were evaluated using one way ANOVA for repeated measures. Differences between groups and sex were assessed by two way ANOVA with interaction terms between groups and sex. Bonferroni correction was used for multiple comparisons. A p value < 0.05 was considered significant. Data were analysed using SPSS software (v 24.0; Chicago, USA).

213

#### 214 **RESULTS**

215 Body weight, FAV and plasma adipokines levels (table 1). Body weight was similar in rats

with NFD and in rats with ARDev+NFD, and enhanced (P<0.05) in both groups with a

217 prolonged exposure to a HFD (table 1). FAV was similarly enhanced (P<0.05) in both sexes

- with NFD. Leptin levels were greater (P < 0.05) in HFD and ARDev+HFD rats than in NFD
- and ARDev+NFD rats. Adiponectin concentration was similar among each group of HFD and
- 221 ARDev+HFD rats and their respective groups with NFD. However, plasma adiponectin levels
- were greater (P < 0.05) in female than in male rats with NFD, ARDev+NFD and
- 223 ARDev+HFD.
- 224 Systolic AP (SAP). Figure 1 shows that SAP was enhanced (P<0.05) in male rats with HFD
- $(148 \pm 2 \text{ mmHg})$ , and in male rats with ARDev exposed to either a NFD ( $154 \pm 2 \text{ mmHg}$ ), or
- a HFD (167  $\pm$  2 mmHg), with respect to the SAP found in male rats with NFD (124  $\pm$  1
- mmHg). Similar changes in SAP were found in females since it was greater (P < 0.05) in rats
- with HFD (136  $\pm$  2 mmHg), ARDev+NFD (135  $\pm$  2 mmHg), or ARDev+HFD (145  $\pm$  2
- mmHg) than in female rats with NFD ( $113 \pm 1 \text{ mmHg}$ ). It also can be seen in figure 1 that
- SAP was greater (P < 0.05) in both sexes of rats with ARDev+HFD than in those with only
- HFD or with ARDev+NFD. Sex-dependent differences in SAP were found in each group,
- being higher (P < 0.05) in males than in females.
- 233 The administration of candesartan alone at four months of age, induced a reduction of SAP in
- each group of rats (Figure 2). The fall of SAP was greater (P<0.05) in male rats with
- ARDev+HFD (55  $\pm$  3 mmHg) than in male rats with only HFD (39  $\pm$  3 mmHg) or with
- ARDev+NFD ( $41 \pm 4$  mmHg). However, no significant differences in the reduction of SAP
- elicited by candesartan were found between females with ARDev+HFD ( $46 \pm 6 \text{ mmHg}$ ) and
- female rats with only HFD  $(33 \pm 5 \text{ mmHg})$  or with ARDev+NFD  $(38 \pm 2 \text{ mmHg})$  (Figure 2).
- Apocynin administration only induced a decrease (P < 0.05) of SAP in both sexes of rats with
- 240 ARDev+NFD (males:  $24 \pm 1$  mmHg; females:  $18 \pm 5$  mmHg) and ARDev+HFD (males:  $23 \pm 1$
- 5 mmHg; females:  $21 \pm 2$  mmHg) (Figure 2). Blockade of AT<sub>1</sub> receptors in apocynin-treated
- rats induced a further decrease (P < 0.05) of SAP in rats with only HFD, rats with

- 243 ARDev+NFD and rats with ARDev+HFD. The decrease in SAP elicited by the simultaneous
- administration of apocynin+candesartan in HFD rats (males:  $28 \pm 2$  mmHg; females:  $30 \pm 7$
- 245 mmHg), ARDev+NFD rats (males:  $34 \pm 4$  mmHg; females:  $31 \pm 4$  mmHg), and
- ARDev+HFD (males:  $42 \pm 3$  mmHg; females:  $34 \pm 3$  mmHg) was not significantly different
- to that induced by candesartan alone (Figure 2).
- 248 Renal hemodynamic response to acute Ang II infusion. Basal renal hemodynamic was
- significantly affected in male and female rats with ARDev+HFD, with respect to the GFR and
- 250 RBF found in rats with NFD (Table 2). Acute Ang II infusion induced a decrease of RBF and
- 251 GFR in each experimental group (Table 2). However, GFR decreased to lower levels in male
- and female rats with ARDev+HFD than in those with only HFD or in rats with ARDev+NFD.
- 253 The Ang II-induced increment in RVR was greater (P<0.05) in both sexes of rats with
- ARDev+HFD (males:  $212 \pm 23\%$ ; females:  $163 \pm 31\%$ ) than in HFD (males:  $106 \pm 14\%$ ;
- 255 females:  $97 \pm 15\%$ ) or ARDev+NFD (males:  $104 \pm 14\%$ ; females:  $90 \pm 7\%$ ) rats.
- 256 <u>The expression of NADPHp67phox</u> in the renal cortex and renal medulla from each group is
- shown in Figure 3, with respect to the expression found in male rats with NFD. The
- expression of NADPHp67phox was similar in rats with NFD and in those only exposed to a
- 259 HFD from weaning. The ARDev led to an increment (P<0.05) of NADPHp67phox in the
- renal cortex and renal medulla in both sexes but its overexpression was greater (P < 0.05) in
- rats with HFD than in those with NFD. A sex-dependent difference in the renal
- 262 NADPHp67phox expression was found in rats with HFD and ARDev+NFD, being greater
- (P<0.05) in females than in males. However, the overexpression of NADPHp67phox in the
- renal cortex and renal medulla of rats with ARDev+HFD was greater (P < 0.05) in males than in females (Figure 3).
- 266 Infiltration of T-cells and macrophages in the renal cortex and renal medulla. Figure 4 shows
- the count of CD3 T-cell in each group of rats. The number of CD3 T-cell was enhanced

268	(P < 0.05) in the renal cortex of both sexes in rats with ARDev but the number of these cells
269	was greater (P<0.05) in ARDev+HFD than in ARDev+NFD rats. The infiltration of CD3 T-
270	cells in the renal cortex was greater (P<0.05) in male than in female rats with ARDev+HFD.
271	The number of CD3 T-cell was significantly elevated in the renal medulla of male rats with
272	ARDev but no differences were found between rats with ARDev+NFD or ARDev+HFD
273	(Figure 4). However, the infiltration of CD3 T-cell in the renal medulla was similar in each
274	group of female rats. Figure 5 shows representative images of the T-CD3 lymphocyte
275	infiltrate in the renal cortex of rats with NFD and rats with ARDev+HFD.
276	Figure 6 shows the infiltration of macrophages in the renal cortex and renal medulla in each
277	group. When compared to the number of macrophages in NFD rats, it was unchanged in the
278	renal cortex and renal medulla in both sexes of HFD rats and only increased (P<0.05) in the
279	renal medulla of male ARDev+NFD rats. The infiltration of macrophages was enhanced
280	(P < 0.05) in the renal cortex of both sexes in ARDev+HFD rats being greater $(P < 0.05)$ in male
281	than in female rats. The number of macrophages was elevated ( $P < 0.05$ ) in the renal medulla
282	of male but not of female ARDev+HFD rats (Figure 6).
283	<u>Glomerulosclerosis index</u> in each group of rats is shown in table 3. The degree of glomerular
284	damage was slightly elevated in both sexes of rats exposed to a HFD from weaning.
285	Glomerular damage was enhanced (P<0.05) in both sexes of rats with ARDev+NFD but the
286	increment was greater (P<0.05) in males than in females. A further sex-dependent increase
287	(P<0.05) of glomerular damage was found in both sexes of rats with ARDev+HFD, with
288	respect to that found in ARDev+NFD or HFD rats. Figure 7 shows representative images of
289	glomeruli with grade 0, 1, 2 and 3 of damage.
290	

# 291 **DISCUSSION**

This study reports new findings showing that the prolonged exposure to a HFD early in life induces a higher increase of arterial pressure and a further deterioration of renal function in both sexes of rats with ARDev. The results obtained suggest that the hypertension and renal
hemodynamic changes induced by HFD in both sexes of rats with an ARDev are secondary to
the Ang II effects, to greater increases in oxidative stress and immune system activation and
to an enhanced glomerular damage. The increments in arterial pressure, oxidative stress and
glomerular damage are sex-dependent, being greater in males than in females.

Previous studies have examined the AP increments after a prolonged HFD in experimental 299 models of developmental programming [10, 12, 18]. However, studies evaluating the renal 300 changes in both sexes of subjects with an ARDev and a prolonged HFD are very limited. The 301 302 mechanisms involved in the cardiovascular and renal function alterations in these subjects are also understudied. Our study shows that the prolonged exposure to a HFD induced an 303 increment of SAP in both sexes of rats with an ARDev, but SAP was greater in male that in 304 female rats (Figure 1). This different AP increment may be partly involved in the sex-305 dependent glomerular damage found in rats with ARDev+HFD. These results are in contrast 306 307 with those reported in two different models of developmental programming with reduced nephron endowment [10, 12]. The differences in the AP increment elicited by the prolonged 308 HFD may be related to the content of fat in the diet and/or to the degree of ARDev in the 309 experimental models used. As also occur in infants born small for their gestational age [30], 310 with smaller kidneys [21], the results of this study suggest that a faster body weight gain early 311 in life leads to a further AP increment in subjects with an ARDev. 312

The increment in AP elicited by a prolonged HFD in rats with ARDev is mainly secondary to the Ang II effects because a three days' infusion of candesartan reduces SAP to the levels found in normotensive rats. The involvement of oxidative stress and inflammatory mediators in the systemic vascular Ang II effects was examined by evaluating the AP changes in response to apocynin administration since it has anti-oxidative and anti-inflammatory effects [15]. The increment in oxidative stress was confirmed by the sex-dependent elevation of

NADPHp67phox expression in the renal cortex and renal medulla of rats with ARDev+HFD. 319 The activation of inflammatory pathways is suggested by the enhanced infiltration of T-CD3 320 321 cells and macrophages in the renal cortex of both sexes and the elevated infiltration of these inflammatory cells in the renal medulla of male but not female rats with ARDev and HFD. An 322 323 increase in plasma and renal oxidized proteins has also been shown in rats with ARDev [25]. 324 The results obtained show that apocynin induces a reduction in SAP in rats with ARDev, and that the administration of candesartan alone in these rats leads to a decrease in SAP that is 325 similar to that elicited by the simultaneous apocynin and candesartan treatment. Collectively, 326 these results suggest that vascular Ang II effects in both sexes of rats with ARDev and a HFD 327 are partly mediated by the activation of oxidative and inflammatory pathways. The 328 contribution of oxidative stress to the hypertension elicited by a HFD in rats with ARDev 329 confirms the results obtained in male offspring of rats exposed to dexamethasone [16] during 330 gestation period. However, this is the first study showing that the increment in oxidative stress 331 332 elicited by a HFD in subjects with ARDev is greater in males than in females. Prolonged HFD in our study induced an increase in FAV and in leptin levels that were similar 333 in males and females, indicating that FAV may be contributing to the higher AP by increasing 334 leptin levels [8]. A similar increment in plasma leptin in both sexes after a prolonged HFD 335 has been reported to occur in rats with intrauterine growth restriction (IUGR) [12] and in Dahl 336 SS rats [6]. Furthermore, similarly to female rats, women show greater adiponectin levels 337 [20]. The SAP response to candesartan in rats with HFD may be related to the increments in 338 339 FAV and leptin since RAS activity is enhanced in adipose tissue of rats with HFD [4], and an 340 increase in leptin levels is associated with an elevated AT1 receptors expression [33]. This study examined to what extent renal function is more susceptible to a prolonged 341 342 exposure to a HFD in rats with ARDev and whether the mechanisms involved in the greater 343 renal susceptibility are sex-dependent. The hypothesis that renal function would be more

susceptible to a prolonged HFD was supported by studies performed in rats exposed to high 344 sodium [29] or high protein [26] diets. The results obtained show that basal renal 345 346 hemodynamic was deteriorated in both sexes of rats with ARDev and a prolonged HFD. Our results are in contrast with those showing that the exposure to a high fat high sugar diet 347 348 induces a decrease of GFR only in males with IUGR secondary to a maternal placental 349 insufficiency [12]. Differences in basal renal function between both studies may be explained by the degree of ARDev in the experimental models used and the % of fat in the diet (45% vs. 350 60%). Our results suggest that the prolonged HFD from an early age reduces even more renal 351 352 functional reserve in subjects with ARDev and exposes the glomerulus to elevated renal perfusion pressures, with the consequent glomerular damage, the impairment of renal function 353 and exacerbation of hypertension. A greater renal vulnerability to overweight and obesity has 354 been reported in patients with a reduced nephron number [24]. 355 The results obtained show that the Ang II-induced increment in RVR is significantly 356 357 enhanced in rats with ARDev and a prolonged exposure to a HFD, when compared to the rise 358 in RVR found in rats with only a HFD or in rats with ARDev+NFD. To the best of our knowledge, the important increment in the renal hemodynamic sensitivity to Ang II, as a 359 360 consequence of the prolonged HFD from an early age in both sexes of subjects with ARDev, has not been previously reported. The renal hemodynamic changes before and during Ang II 361 infusion could be explained by the coexistence of a progressive reduction in the number of 362 undamaged nephrons, due to glomerular sclerosis, with significant increments in leptin, renal 363 364 oxidative stress and pro-inflammatory cytokines derived from the elevated infiltration of 365 lymphocytes and macrophages in the kidney. The enhanced renal sensitivity to Ang II may then contribute to a further increase of glomerular damage [19]. Elevated leptin levels may 366

367 contribute to these hemodynamic changes because the kidney expresses abundant leptin

368 receptors, and patients with high leptin levels are more prone to develop nephropathy [32].

369 The renal hemodynamic effects of leptin seem to be mediated by an increase in sympathetic370 activity [8] and reactive oxygen species (ROS) production [3].

An enhanced oxidative stress may contribute to the renal hemodynamic effects of Ang II 371 because it stimulates NADPH oxidase [1] and the blockade of RAS activity delay the 372 progression of renal disease by promoting anti-oxidative effects [2]. The increments of 373 immune cells may be Ang II-dependent because their receptors are present on the surface of 374 monocytes [11]. The elevated oxidative stress probably contributes to the generation of 375 cytokines that would recruit additional inflammatory cells, and to the progressive elevations 376 of RVR and AP in both sexes of rats with ARDev and HFD [9]. The greater increment in the 377 infiltration of T-lymphocytes and macrophages in the kidney of these male rats may also 378 contribute to the sex-dependent oxidative stress increment since an enhanced infiltration of 379 immune cells create oxidative stress, and the prevention of their infiltration not only reduces 380 oxidative stress but also ameliorates the hypertension [5]. Accumulation of ROS may provide 381 382 the initial stimulus for attracting immune cells [14] and their secretion of pro-inflammatory cytokines would then lead to a further ROS production and a vicious circle with a progressive 383 deterioration of renal function. The possible increase of cytokines secreted by the elevated 384 infiltration of CD-3 cells and macrophages and the increment in oxidative stress may also be 385 involved in the greater glomerular damage in rats with ARDev and a prolonged HFD [11]. 386 Contrary to what was expected from those obtained in other experimental model [6], the 387 results in this study suggest that there are no sex-dependent differences in the deterioration of 388 389 renal function as a consequence of a prolonged exposure to a HFD in subjects with ARDev 390 during nephrogenic period. The absence of sex-dependent differences in the renal hemodynamic changes occurs despite glomerular damage and the increments in oxidative 391 392 stress and infiltration of immune cells are greater in male than in female rats. This difference 393 in the renal T-cells infiltration has also been shown in rats with Ang II hypertension [23] and

may contribute to the sex-dependent differences in renal NADPHp67phox expression and APlevels found in rats with ARDev and a prolonged HFD [13].

396 The new data reported in this study contribute to improve our understanding of the mechanisms involved in the hypertension and renal dysfunction secondary to the prolonged 397 exposure to a HFD in both sexes of subjects with ARDev. These results strongly suggest that 398 399 high calorie diets should be especially prevented during childhood in patients with ARDev since it may lead to a cardiovascular and renal dysfunction by increasing the vascular Ang II 400 401 effects and immune system activation, and inducing a sex-dependent elevation in oxidative 402 stress and glomerular damage. Considering that the intake of high calorie diets is frequent during childhood, these results may help to improve the treatment of the cardiovascular and 403 renal consequences of a prolonged exposure to a HFD from an early age in both sexes of 404 subjects with ARDev. 405

407 **DECLARATIONS** 

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412

# 413 Conflicts of interest/Competing interests

414 They are no disclosures.

415

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416 Ethics approval
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The procuration of animals, the husbandry and the experiments conform to the European Convention for the Protection of Vertebrate Animals used for Experimental and other Scientific Purposes. Approval was obtained from the ethics committee of the University of Murcia, Spain.

421

### 422 Consent for publication

423 All authors revised the manuscript and approved the version to be published.

424

# 425 Availability of data and material

426 All raw data that support the findings of this study will be available without restriction from

427 the corresponding author on reasonable request.

428

# 429 Authors' contributions

- Moreno JM, Salazar FJ and Llinas MT contributed to the conception and design of the work.

- All authors contributed to the acquisition, analysis or interpretation of data, and agree to be

432 accountable for all aspects of the work in ensuring that questions related to the accuracy or

- 433 integrity of any part of the work are appropriately investigated and resolved. The authors
- 434 declare that all data were generated in-house and that no paper mill was used.

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- 553 Figure legends.
- **Figure 1**. Systolic arterial pressure in male and female rats with normal (NFD) or high (HFD)

fat diet, and in rats with altered renal development and normal (ARDev+NFD) or high

556 (ARDev+HFD) fat diet. Rats were exposed to NFD or HFD from weaning to the end of the

experiment. Values are shown as mean  $\pm$  SEM. Differences between groups and sex were

assessed by two way ANOVA with Bonferroni's multiple comparisons analysis. \* P < 0.05 vs.

559 NFD rats; # P<0.05 vs. HFD rats;  $\blacklozenge$  P<0.05 vs. ARDev+NFD rats. + P<0.05 vs. respective

- 560 male rats;  $\phi$  P<0.05 sex-dependent differences.
- 561

Figure 2. Systolic arterial pressure changes elicited by 3-day oral administration of 562 candesartan (7 mg·kg-1·day-1), or 3 weeks oral administration of apocynin (60 mg·kg-1·day-563 1), without or with simultaneous candesartan treatment. Arterial pressure changes were 564 examined in rats with normal (NFD) or high (HFD) fat diet, and in rats with altered renal 565 566 development and normal (ARDev+NFD) or high (ARDev+HFD) fat diet. Rats were exposed to NFD or HFD from weaning to the end of the experiment. Values are shown as mean  $\pm$ 567 SEM. Differences with respect to basal period and vs. apocynin treatment were assessed by 568 one-way ANOVA for repeated measures. Differences between groups and sex were evaluated 569 using two-way ANOVA with Bonferroni's multiple comparisons analysis.  $\delta$  P<0.05 vs. basal 570 period. # P<0.05 vs. HFD rats;  $\blacklozenge$  P<0.05 vs. ARDev+NFD rats;  $\lambda$  P<0.05 vs apocynin 571 572 treatment alone.

573

**Figure 3.** NADPHp67phox expression in the renal cortex and renal medulla from rats with normal (NFD) or a high (HFD) fat diet, and in rats with altered renal development and normal (ARDev+NFD) or high (ARDev+HFD) fat diet. Representative blots for NADPHp67phox and  $\beta$ -actin levels are shown. Values are shown as mean  $\pm$  SEM. Differences between groups and sex were assessed by two way ANOVA with Bonferroni's multiple comparisons analysis. \* P<0.05 vs. NFD rats; # P<0.05 vs. HFD rats;  $\blacklozenge$  P<0.05 vs. ARDev+NFD rats. + P<0.05 vs. respective male rats;  $\blacklozenge$  P<0.05 sex-dependent differences.

581	Figure 4. Number of T-CD3 lymphocytes cells per field in the renal cortex and renal medulla
582	of male and female rats with normal (NFD) or a high (HFD) fat diet, and in rats with altered
583	renal development and normal (ARDev+NFD) or high (ARDev+HFD) fat diet. Rats were
584	exposed to NFD or HFD from weaning to the end of the experiment. Values are shown as
585	mean $\pm$ SEM. Differences between groups and sex were evaluated using two-way ANOVA
586	with Bonferroni's multiple comparisons analysis. * P<0.05 vs. NFD rats; # P<0.05 vs. HFD
587	rats; $\bullet P < 0.05$ vs. ARDev+NFD rats. + P < 0.05 vs. respective male rats.
588 589	Figure 5. Representative images of the T-CD3 lymphocyte infiltrate (arrowheads) in the renal
590	cortex of male and female rats with normal fat diet (NFD) and male and female rats with
591	altered renal development and high fat diet (ARDev+HFD). Indirect ABC-
592	immunohistochemical anti T-CD3. Scale bar: 50 micrometers.
593 594	Figure 6. Number of macrophages cells per field in the renal cortex and renal medulla of rats
595	with normal (NFD) or a high (HFD) fat diet, and of rats with altered renal development and
596	normal (ARDev+NFD) or high (ARDev+HFD) fat diet. Rats were exposed to NFD or HFD
597	from weaning to the end of the experiment. Values are shown as mean $\pm$ SEM. Differences
598	between groups and sex were evaluated using two-way ANOVA with Bonferroni's multiple
599	comparisons analysis. * P<0.05 vs. NFD rats; # P<0.05 vs. HFD rats; • P<0.05 vs.
600	ARDev+NFD rats. + P<0.05 vs. respective male rats. $\phi$ P<0.05sex-dependent differences.
601 602	Figure 7. Representative images of different degrees of glomerular sclerosis. Profiles without
603	pathological changes were assigned to grade 0; grade 1 (moderated changes) was composed
604	of glomerular profiles with an increase of mesangial matrix and thickening of glomerular
605	basement membrane (arrows); grade 2 (advanced changes) consisted of sclerotic glomerular
606	profiles (arrows); and grade 3 was global sclerosis (*). Scale bar = 50 micrometers.

Table 1. Body weight (BW, grams), fat abdominal volume (FAV) (cm<sup>3</sup>·100cm<sup>3</sup>-1) and
plasma levels of leptin (ng·ml-1) and adiponectin (ng·ml-1) in rats treated with normal (NFD)
or high (HFD) fat diet, and in rats with altered renal development and normal (ARDev+NFD)
or high (ARDev+HFD) fat diet. Rats were exposed to NFD or HFD from weaning to 4
months of age.

612	MALES	NFD	HFD	ARDev+NFD	ARDev+HFD
613	BW	$450\pm7$	$538 \pm 5*$	$458\pm15\#$	510 ± 12*♦
614	FAV	$7,9\pm0,5$	$10,1 \pm 0,4*$	$5,7 \pm 0,5$ *#	11,1 ± 0,5 <b>*</b> ♦
615	Leptin	$1,\!69\pm0,\!37$	$4,89 \pm 0,65*$	$1,24 \pm 0,16 \#$	3,52 ± 0,55* ♦
616	Adiponectin	$5{,}33 \pm 0{,}45$	$5{,}79 \pm 0{,}40$	$4,\!90 \pm 0,\!52$	$5,\!64 \pm 0,\!33$
617					
618	FEMALES	NFD	HFD	ARDev+NFD	ARDev+HFD
619	BW	$273\pm6\text{+}\phi$	$298\pm4\text{*+}\phi$	$262 \pm 5\#+\phi$	$296 \pm 10 \blacklozenge + \phi$
620	FAV	$6,\!4 \pm 0,\!6 +$	$9,7 \pm 0,4*$	$4,5 \pm 0,5*$	$9,8\pm0,6^{*} \blacklozenge$
621	Leptin	$2,11 \pm 0,34$	$3,35 \pm 0,29*+$	$0,\!80 \pm 0,\!24 \#$	3,15 ± 0,69 ♦
622	Adiponectin	$8,56 \pm 0,44 +$	$6{,}66 \pm 0{,}58$	$8,77 \pm 0,49 +$	$8,16 \pm 0,82 +$
623					

Values are presented as mean ± SE. Differences between groups and sex were evaluated using
two-way ANOVA with Bonferroni's multiple comparisons analysis. \* P<0.05 vs. NFD group.</li>
# P<0.05 vs. HFD. ♦ P<0.05 vs. ARDev+NFD. + P<0.05 vs. respective male rats. ♦ P<0.05</li>

627 sex-dependent differences.

Table 2. Changes in renal plasma flow (RPF) (ml·min-1·g kw-1) and glomerular filtration
rate (GFR) (ml·min-1· g kw) in response to Ang II infusion, in rats with normal (NFD) or
high (HFD) fat diet, and in rats with altered renal development and normal (ARDev+NFD) or
high (ARDev+HFD) fat diet. Rats were exposed to NFD or HFD from weaning to 4 months
of age.

635			HFD	ARDev+NFD	ARDev+HFD
	RBF				
636	Basal	$3,\!55\pm0,\!47$	$2{,}96\pm0{,}27$	$2,\!55\pm0,\!27$	$1,81 \pm 0,17*$
637	Ang II	$2{,}51\pm0{,}32\delta$	$1,63 \pm 0,15*\delta$	$1{,}43\pm0{,}13{*}\delta$	$0,75\pm0,15*\delta$
638 639	GFR				
640	Basal	$1,10 \pm 0,09$	$1,\!19\pm0,\!08$	$0,\!80 \pm 0,\!04$ *#	0,53 ± 0,07*#♦
641 642	Ang II	$0,86 \pm 0,06\delta$	$0,72\pm0,04\delta$	$0{,}55\pm0{,}05{*}\delta$	0,19 ± 0,05*δ#♦
643	FEMALES	NFD	HFD	ARDev+NFD	ARDev+HFD
643 644	<b>FEMALES</b> RBF	NFD	HFD	ARDev+NFD	ARDev+HFD
643 644 645	FEMALES RBF Basal	NFD 4,03 ± 0,52	HFD 3,43 ± 0,14	ARDev+NFD 4,67 ± 0,41+	ARDev+HFD 1,92 ± 0,15* ♦
643 644 645 646	FEMALES RBF Basal Ang II	NFD $4,03 \pm 0,52$ $2,79 \pm 0,358$	HFD $3,43 \pm 0,14$ $1,97 \pm 0,16\delta$	ARDev+NFD $4,67 \pm 0,41+$ $2,82 \pm 0,28+\delta$	ARDev+HFD 1,92 ± 0,15* ♦ 0,93 ± 0,13*8 ♦
643 644 645 646 647	FEMALES RBF Basal Ang II	NFD $4,03 \pm 0,52$ $2,79 \pm 0,35\delta$	HFD $3,43 \pm 0,14$ $1,97 \pm 0,16\delta$	ARDev+NFD $4,67 \pm 0,41+$ $2,82 \pm 0,28+\delta$	ARDev+HFD 1,92 ± 0,15* ♦ 0,93 ± 0,13*δ ♦
643 644 645 646 647 648	FEMALES RBF Basal Ang II GFR	NFD $4,03 \pm 0.52$ $2,79 \pm 0.35\delta$	HFD $3,43 \pm 0,14$ $1,97 \pm 0,16\delta$	ARDev+NFD 4,67 $\pm$ 0,41+ 2,82 $\pm$ 0,28+ $\delta$	ARDev+HFD 1,92 ± 0,15* ♦ 0,93 ± 0,13*δ ♦
643 644 645 646 647 648 649	FEMALES RBF Basal Ang II GFR Basal	NFD $4,03 \pm 0,52$ $2,79 \pm 0,35\delta$ $1,16 \pm 0,07$	HFD $3,43 \pm 0,14$ $1,97 \pm 0,16\delta$ $1,21 \pm 0,11$	ARDev+NFD 4,67 $\pm$ 0,41+ 2,82 $\pm$ 0,28+ $\delta$ 1,09 $\pm$ 0,04+	ARDev+HFD $1,92 \pm 0,15^* \blacklozenge$ $0,93 \pm 0,13^* \& \blacklozenge$ $0,62 \pm 0,04^* \# \blacklozenge$

Values are presented as mean  $\pm$  SEM. Differences with respect to basal period were assessed

by one-way ANOVA for repeated measures. Differences between groups and sex were

evaluated using two-way ANOVA with Bonferroni's multiple comparisons analysis.

654 δ P<0.05 vs. Basal; \* P<0.05 vs. NFD rats; # P<0.05 vs. HFD rats;  $\blacklozenge$  P<0.05 vs.

ARDev+NFD rats. + P < 0.05 vs. respective male rats.

658	MALES	% Grade 0	% Grade 1	% Grade 2	% Grade 3
659	NFD	$90{,}6\pm0{,}7$	$9,0 \pm 0,3$	$0,5\pm0,5$	$0,0\pm0,0$
660	HFD	$67,2 \pm 9,3*$	$30,2 \pm 9,0$	$2,3 \pm 0,8$	$0,3 \pm 0,3$
661	ARDev+NFD	23,8 ± 3,3*#	62,0 ± 2,6*#	$12,8 \pm 3,9$	$1,4 \pm 0,6$
662	ARDev+HFD	$17,8 \pm 5,2$ *#	$49,8 \pm 8,5*$	$22,8 \pm 8,4*$ #	9,6 ± 3,3*#♦
663					
664	FEMALES				
665	NFD	$93,8 \pm 0,3$	$6,2 \pm 0,3$	$0,0\pm0,0$	$0,0\pm0,0$
666	HFD	$65,3 \pm 7,1*$	$33,3 \pm 6,7*$	$1,2 \pm 0,5$	$0,2 \pm 0,2$
667	ARDev+NFD	$55,8 \pm 6,7*+\phi$	$36,8 \pm 5,6*+$	4,8 ± 3,9*#	$2,6 \pm 0,9$
668	ARDev+HFD	$45,0 \pm 1,6*+\phi$	$39,0 \pm 2,2*$	$7,6 \pm 0,8$ *#+ $\phi$	8,4 ± 2,0*#♦
669					

**Table 3**. Percent of glomeruli with degrees of glomerulosclerosis Index

Grade 0 indicates normal; grade 1, moderated changes: grade 2, advanced changes; grade 3,

global sclerosis. Data are expressed as percent of area. Values are presented as mean  $\pm$  SE.

672 Differences between groups and sex were evaluated using two-way ANOVA with

Bonferroni's multiple comparisons analysis. \* P<0.05 vs. NFD rats; # P<0.05 vs. HFD rats;

• P<0.05 vs. ARDev+NFD rats. + P<0.05 vs. respective male rats;  $\phi$  P<0.05 sex-dependent

675 differences.