REVIEW



Role of mitochondria in neonatal hypoxic-ischemic encephalopathy

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Summary. Neonatal hypoxic-ischemic encephalopathy, an important cause of death as well as long-term disability in survivors, is caused by oxygen and glucose deprivation, and limited blood flow. Following hypoxicischemic injury in the neonatal brain, three main biochemical damages (excitotoxicity, oxidative stress, and exacerbated inflammation) are triggered. Mitochondria are involved in all three cascades. Mitochondria are the nexus of metabolic pathways to offer most of the energy that our body needs. Hypoxicischemic injury affects the characteristics of mitochondria, including dynamics, permeability, and ATP production, which also feed back into the process of neonatal hypoxic-ischemic encephalopathy. Mitochondria can be a cellular hub in inflammation, which is another main response of the injured neonatal brain. Some treatments for neonatal hypoxic-ischemic encephalopathy affect the function of mitochondria or target mitochondria, including therapeutic hypothermia and erythropoietin. This review presents the main roles of mitochondria in neonatal hypoxic-ischemic encephalopathy and discusses some potential treatments directed at mitochondria, which may foster the development of new therapeutic strategies for this encephalopathy.

Key words: Neonatal hypoxic-ischemic encephalopathy, Mitochondria, HIE

Introduction

Neonatal hypoxic-ischemic encephalopathy (HIE), an important cause of death as well as long-term disability in survivors, is caused by oxygen and glucose deprivation, and limited blood flow (Fig. 1) (Verklan and Walden, 2015). Asphyxia during the perinatal period can induce HIE. The incidence of HIE was estimated as 1.5 (95% CI 1.3 to 1.7) per 1000 live births with population-

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based results (Kurinczuk et al., 2010). The mortality rate of HIE (deaths/live births) was reported as 0.32% (Velaphi and Pattinson, 2007). Queensland Clinical Guidelines updated the clinical guidelines for HIE in 2021, including assessment, diagnosis, and management (Queensland-Clinical-Guidelines, 2021). Hypothermia is currently the only proven therapy for moderate and severe HIE, which improves mortality and long-term outcomes for babies (Wassink et al., 2019; Queensland-Clinical-Guidelines, 2021). In addition to hypothermia, some agents that may be synergistic with hypothermia are proven to relieve patients with HIE at the molecular level (Ruegger et al., 2018; Gou et al., 2020; Wu et al., 2022).

Mitochondria are the nexus of metabolic pathways to offer most of the energy that our body needs. Following hypoxic-ischemic (HI) injury, the characteristics of mitochondria, including dynamics, permeability, and ATP production, are affected. These altered mitochondrial characteristics also feed back into the process of HIE (Thornton et al., 2018; Rodriguez et al., 2020). There are some good review articles about the roles of mitochondria in HIE (Thornton et al., 2018; Ham and Raju, 2017). Until July 04, 2023, the PubMed database offered 447 articles with two keywords ("hypoxic-ischemic encephalopathy" and mitochondri*), which indicates that researchers are still interested in this content. In this review, mitochondrial roles in HIE were reviewed to offer ideas for potential treatments.

Damaged HIE cascades

Following HI injury in the brain, three main damaged cascades (excitotoxicity, oxidative stress, and exacerbated inflammation) were triggered (Pedroza-Garcia et al., 2022). Initially, the adenosine triphosphate (ATP) deficit, caused by lack of blood and oxygen, induces Na⁺/K⁺ pump failure, causing abnormal intracellular accumulation of sodium, calcium, and water, which will lead to neuronal depolarization and glutamate release. The release of glutamate, an excitatory neurotransmitter, over activates ionotropic receptors, which will enhance excitotoxicity and aggravate HI brain injury (Iovino et al., 2020).



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After HI attack, reactive oxygen species (ROS) production rapidly increases and overwhelms the antioxidant defenses, which are immature in the neonatal brain (Zhao et al., 2016). During reperfusion, increased production of ROS will worsen the situation (Widgerow, 2014).

Inflammation is another main response of the injured brain. Microglia detect changes in the environment and will be activated. Activated microglia migrate to injured sites and produce proinflammatory cytokines, including tumor necrosis factor and chemokines to recruit additional cells and remove pathological agents (Kwon and Koh, 2020; Rodriguez-Gomez et al., 2020). Similar to microglia, astrocytes have pro-inflammatory and immunoregulatory subpopulations, which are involved in inflammation in HIE (Kwon et al., 2020).

Mitochondria are involved in all three cascades, they offer most of the energy, including ATP, we get to spend. Mitochondria account for the majority of oxygen consumption and are a potential source of ROS (Fuhrmann and Brune, 2017). Mitochondria can be a cellular hub for infection and inflammation (Andrieux et al., 2021).

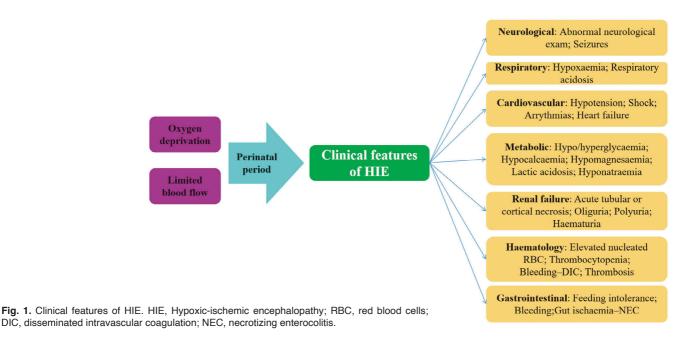
Oxidative phosphorylation

Under basal physiological conditions, glucose metabolizes into nicotinamide adenine dinucleotide (NADH), ATP, and two molecules of pyruvate, which are oxidized into acetyl CoA and an additional molecule of NADH and carbon dioxide (CO₂). The acetyl CoA is used in the citric acid cycle to offer CO₂, NADH, flavin adenine dinucleotide (FADH₂), and ATP. In the mitochondrial inner membrane, NADH and FADH₂ are used in oxidative phosphorylation (OXPHOS) to synthesize ATP through an electrochemical

transmembrane gradient that is maintained by the electron transfer chain (ETC) (Gilkerson et al., 2003). Electrons are passed to oxygen, the final electron carrier in aerobic cellular respiration, by Complex IV (cytochrome oxidase) through a series of redox reactions. Electrons may be leaked from the ETC to oxygen producing primarily the superoxide anion (O_2) (Turrens, 2003). Under normal conditions, 50-60% of total O_2 is spent on maintenance of ionic gradients (Erecinska and Silver., 1989) (Fig. 2).

During HI attack, oxygen and glucose deprivation and limited blood flow will induce cells to a bioenergetic crisis caused by anaerobic metabolism with limited glucose. Aerobic metabolism and anaerobic metabolism produce 32-36 ATP and 2 ATP with a single glucose (Erecinska et al., 1989; Goldhaber, 1997). Lack of oxygen, the final electron carrier in the ETC, blocks the transfer of electrons and reduces ionic gradients, which decreases the production of ATP and causes primary energy failure (Fig. 2). Compared with adults, the efficiency of glycolysis in newborns is lower, which leads to more severe primary energy failures. The main reason for this phenomenon is that glucose transporter proteins GLUT1 (glial) and GLUT3 (neuronal) at seven days are expressed at only about 10% of their adult levels (Vannucci et al., 2005).

As the duration of hypoxia increases, ionic gradients imbalance and the ATP decrease will become more severe. Theoretically, the production rate of ROS should decrease with a decrease in oxygen availability. Interestingly, detailed investigations confirmed that the production of ROS showed a burst under conditions of mild hypoxia (1-3% O_2) but not during severe hypoxia or anoxia (Schroedl et al., 2002; Guzy and Schumacker, 2006; Hernansanz-Agustin et al., 2014). ROS reduced gradually and disappeared about one hour later, however,



these ROS could also damage cells (Hernansanz-Agustin et al., 2014). Deactivation of mitochondrial complex I, switching to a Na⁺/H⁺ antiporter, is involved in ROS production to face acute hypoxia (Hernansanz-Agustin et al., 2017). In addition to this switching, complex I deactivation presented three functions: 1) reversed the reaction of the tricarboxylic acid cycle dehydrogenases, which are known to produce ROS by this reaction (Tretter and Adam-Vizi, 2004); 2) modified ubiquinone pool redox state; and 3) triggered the production of superoxide by complex III (Chandel et al., 1998).

After HI attack, reperfusion and restoration of blood flow send oxygen and glucose back to the neurons. ATP is generated gradually to restore the cell's energy resources. During primary energy failure, cytosolic Ca² is overloaded, which evokes an increase in intermitochondrial Ca^{2+} to reduce cytosolic Ca^{2+} (Zaidan and Sims, 1994; Puka-Sundvall et al., 2000). Calcium is an activation signal for mitochondrial phosphatases, which induces dephosphorylation of multiple mitochondrial proteins, including cytochrome c (Cytc) and cytochrome c oxidase (COX), to active their functions (Hopper et al., 2006; Huttemann et al., 2012; Kadenbach, 2020). In addition to the effect on the ETC transfer rate, dephosphorylation of COX also leads to the loss of allosteric inhibition by ATP (Huttemann et al., 2008). Activated oxidative phosphorylation proteins (OxPhos) could hyperpolarize mitochondrial membrane potential, which could be further exacerbated by the loss of allosteric inhibition by ATP (Huttemann et al., 2008). Hyperpolarization of mitochondrial membrane potential (exceeding 140 mV) causes an exponential increase in ROS generation (Starkov and Fiskum, 2003; Liu, 1999, 2010). Accelerated production of ROS overwhelms the antioxidant system, which is immature in neonatal brain

(Zhao et al., 2016). Before reperfusion injury becomes obvious, the stage is described as the latent phase (Fig. 2).

During primary energy failure, ATP-driven active transport loses its function, which destroys cell homeostasis, induces glutamate-induced excitotoxicity, and leads to the imbalance of Na⁺, Ca2⁺, and water (Hasselbach and Makinose, 1962; Mahmoud et al., 2019; Iovino et al., 2020; Chen and Lui, 2022). Intracellular calcium accumulation generates mitochondrial swelling, which is the major factor leading to mitochondriamediated cell death (Chapa-Dubocq et al., 2018). Calcium influx also activates nitric oxide synthase (NOS) to synthesize the free radical nitric oxide ('NO), which has high affinity for complex IV (Torres et al., 1998; Cooper and Giulivi, 2007). 'NO not only inhibits the activity of complex IV to block electron transfer but also reacts with O_2^{\bullet} to produce peroxynitrite (ONOO-), which is a mitochondrial and cellular toxin (Moller et al., 2019; Radi et al., 2002). Exceeding ROS levels are proven to cause significant damage to biological macromolecules, such as protein degeneration, lipid oxidation, and DNA degeneration (Vranic et al., 2018). Thereafter, a phase of secondary energy failure occurs, principally due to mitochondrial dysfunction. In newborn piglets, phosphorus magnetic resonance spectroscopy (³¹P-MRS) presented progressively declined cerebral bioenergetics and secondary energy failure after a few hours of reperfusion (Lorek et al., 1994).

Permeability

Two membranes, inner and outer, separate the mitochondrial matrix from the cytosol, nonetheless,

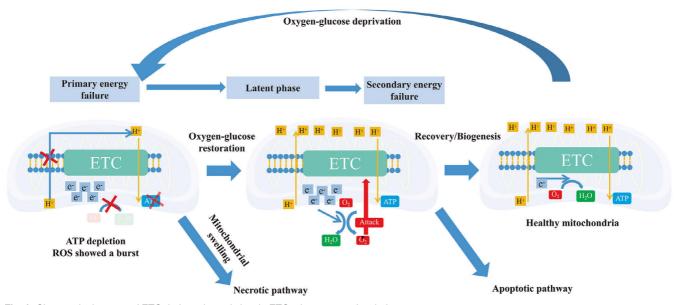


Fig. 2. Changes in the state of ETC during a hypoxic insult. ETC, electron transfer chain.

matrix and cytosol can exchange their contents through the mitochondrial permeability transition pore (mPTP). ETC complexes are all in an ion-impermeable inner membrane to maintain membrane potential, which is important for ATP synthesis. Switches of mPTP play an important physiological role, in maintaining healthy mitochondria homeostasis. Although the molecular characterization of the mPTP is not clear, some molecular compositions of mPTP have been proven, including the adenosine nucleotide translocate (ANT) family of proteins at the inner membrane and the Bcl-2 family of proteins at the outer membrane (Halestrap et al., 1997; Marzo et al., 1998) (Fig. 3). In immature rats, data suggested that mPTP opening occurred after HI attack (Puka-Sundvall et al., 2001). mPTP is regulated by many factors, including Ca²⁺, ROS, and Cyclophilin D (CypD) (Robichaux et al., 2022).

Mitochondrial Ca^{2+} uptake is important for cellular homeostasis. Mitochondrial membrane potential maintained by proton extrusion by ETC is the driving force for Ca^{2+} uptake into the mitochondria. The voltage-dependent anion channel (VDAC) protein family forms the main channel that regulates ions, including Ca^{2+} , and metabolite flux between the outer mitochondrial membrane and the cytosol (Sander et al., 2021). The mitochondrial calcium uniporter (MCU) complex in the mitochondrial inner membrane is identified as the main route for Ca^{2+} uptake from outside the inner membrane to the matrix (Baughman et al., 2011; De Stefani et al., 2011). Cooperation between VDAC and MCU introduces Ca^{2+} into the matrix. Meanwhile, excess intramitochondrial Ca²⁺ plays a crucial role in mPTP opening to maintain the normal level of Ca²⁺ in mitochondria (Haworth and Hunter, 1979). This phenomenon has an interesting name: " Ca^{2+} -induced Ca^{2+} release" (Fabiato and Fabiato, 1975). During primary energy failure, abnormal accumulation of cytosolic Ca^{2+} induces Ca^{2+} uptake into the mitochondria by retained mitochondrial membrane potential. Ca²⁺ overload will induce mPTP opening to cause mitochondrial depolarization and swelling of the organelles (Bernardi, 2013). Homo/hetero oligomers formed by BAX and BAK, of the Bcl-2 family containing at least one Bcl-2 homology (BH) domain, may be involved in Ca²⁺-dependent mPTP opening because mPTP sensitivity to matrix Ca²⁺ is regulated by BH3 mimetic inhibition of the anti-apoptotic Bcl-2 family members localized to mitochondria (Patel et al., 2021). BAX locates to the mitochondrial outer membrane; however, BAX works together with inner membrane proteins, the ANT family, to mediate mPTP opening (Marzo et al., 1998). Independent of Bax and Bak, Ca²⁺ can mediate mitochondrial inner membrane permeabilization to induce cell death, although the exact mechanism is not clear (Quarato et al., 2022).

Analogous to the phenomenon of " Ca^{2+} -induced Ca^{2+} release" is ROS-induced mPTP opening followed by a "burst phase" of ROS generated by the particular mitochondrion and lasting 5-10 s (Zorov et al., 2000). Hicks and Gebicki first established a quantitative relationship between ROS concentration and rate of lipid peroxidation (Hicks and Gebicki, 1978). For the

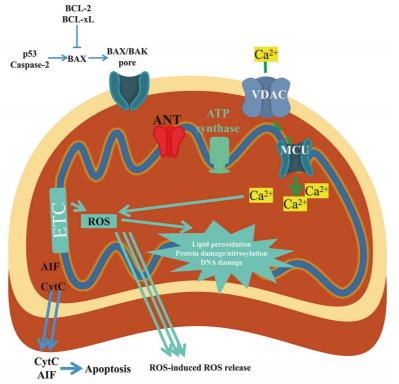


Fig. 3. Factors in regulation of mitochondrial permeability during hypoxic-ischemic attack. AIF, apoptosis-inducing factor; ANT, adenosine nucleotide translocate; BAX, BCL2-associated X; BAK, BCL2 antagonist/killer; BCL2, B-cell lymphoma 2; BCL-xL, B-cell lymphoma-extra-large; CytC, Cytochrome c; ETC, electron transfer chain; MCU, mitochondrial calcium uniporter; ROS, reactive oxygen species; VDAC, Voltage-dependent anion channel.

phospholipid bilayer, ROS oxidizes the head groups first, followed by the lipid tails (Yusupov et al., 2017b). Oxidation of the lipids alters the structural and dynamic properties of the membrane, which can lead to an increase in the permeability and efflux of contents held within (Chen and Yu, 1994; Yusupov et al., 2017a). ROS not only oxidizes lipids but also targets mPTP regulator proteins via posttranslational modifications (PTMs) to mediate mPTP opening. For example, ROS modified the ANT family and VDAC proteins allowing activation of the mPTP pore with a lower concentration of Ca²⁺ (Chang et al., 2014). ROS can sensitize mitochondria to Ca²⁺-dependent mPTP opening.

CypD, peptidyl-prolyl cis/trans isomerase, is an activator of mPTP opening because deletion of CypD reduces the sensitivity of mitochondria to Ca²⁺dependent mPTP opening (Crompton et al., 1998; Woodfield et al., 1998). Cyclophilin A (CsA) interacts with CypD to inhibit mPTP (Kim et al., 2014; Haleckova and Benek, 2022). Compared with adult mice, neonatal mice had a considerably higher induction threshold and lower sensitivity to CsA (Wang et al., 2009). CypDrelated mPTP opening is critical for the development of brain injury in the adult, whereas in neonatal HI brain injury, mitochondrial permeabilization appears to be primarily Bax-dependent (Wang et al., 2009, 2010). In neuronal mitochondria from postnatal day 1 rat pups, Ca²⁺-induced mPTP exhibited limited sensitivity to CsA (Brustovetsky and Dubinsky, 2000).

mPTP opening mainly happens during reperfusion and is a major cause of reperfusion injury (Halestrap, 2009; Morciano et al., 2017). Due to mPTP opening, mitochondria release accumulative ROS and Ca^{2+} , which induce damage in biological macromolecules, excitotoxicity, and inflammatory response. Except for ROS and Ca^{2+} , the efflux of apoptosis-inducing factors, including apoptosis-inducing factor (AIF) and cytochrome c (CytC), will activate apoptosis and cause neuron loss in the immature brain after HI (Kratimenos et al., 2017; Li et al., 2020) (Fig. 3).

Mitochondrial dynamics

Mitochondria are highly dynamic organelles that constantly undergo events of fission, fusion, biogenesis, mitophagy, and cytoskeleton-based transport. The master mediator of mitochondrial fission is dynamin-related protein 1 (DRP1) (Mears and Ramachandran, 2022). The main components involved in mitochondrial fusion are also members of the dynamin-like family of GTPases-Mitofusins 1 and 2 (MFN1/2) in the outer membrane, and optic atrophy 1 (OPA1) protein in the inner membrane (Dorn, 2020). Master regulators of biogenesis are the transcription factors TFAM, NRF1/2, and PGC1 α (Ventura-Clapier et al., 2008). PINK1 (PTEN-induced putative kinase 1), PARKIN (Parkin RBR E3 ubiquitinprotein ligase), and FUNDC1 (FUN14 domain containing 1) have important roles in mitophagy (Sica et al., 2016) (Fig. 4).

Dynamic regulation helps mitochondria face different environments (Popov, 2020; Yapa et al., 2021). Fusion lets mitochondria share their contents, including

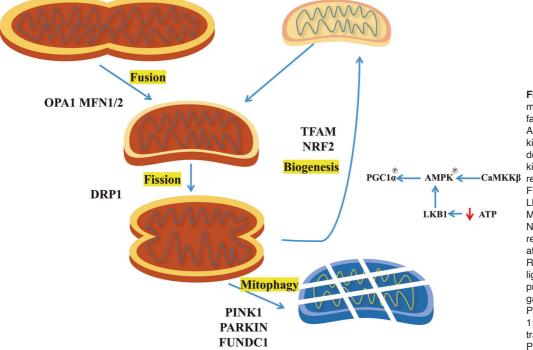


Fig. 4. Dynamics of mitochondria and related factors during a hypoxic insult. AMPK, AMP-activated protein kinase; CaMKKß, Calmodulindependent protein kinase kinase-β; DRP1, Dynamin--CaMKKβ related protein 1; FUNDC1, FUN14 domain containing 1; LKB1, Liver kinase B1; MFN1/2. Mitofusins 1 and 2: NRF2, Nuclear factor E2related factor 2; OPA1, Optic atrophy 1: PARKIN, Parkin RBR E3 ubiquitin-protein ligase; PGC-1a, Peroxisome proliferator-activated receptorgamma coactivator-1a; PINK1, PTEN-induced putative kinase 1: TFAM. Mitochondrial transcription factor A; ®, Phosphorylation.

mitochondrial DNA (mtDNA), proteins, and lipids. Meanwhile, minor defects caused by low-level mtDNA mutation or accumulating ROS can be fused with more complex mitochondrial networks to correct the defects (Youle et al., 2012). If the area is damaged, fission helps mitochondria clear dysfunctional parts. The fate of mitochondria following fission is based on mitochondrial conditions, including ROS, membrane potential, and mtDNA mutation. Healthy mitochondria act as a "seed", which is used as a substrate for biogenesis. Damaged mitochondria trigger the accumulation and dimerization of PINK1 to initiate mitophagy (Youle and van der Bliek, 2012; Jones and Thornton, 2022).

In neurons (Baburamani et al., 2015; Li et al., 2016), astrocytes (Quintana et al., 2019), and microglia (Zhou et al., 2019), HI attack can increase mitochondrial fission. Few studies about the mitochondrial morphology of the immature brain after HI in vivo have been reported, however, rapid induction of mitochondrial fragmentation after neonatal HI in immature rats was observed (Demarest et al., 2016). HI-induced fragmentation of mitochondria is sex-specific: fission was more prominent in males compared with female ipsilateral sections (Demarest et al., 2016). Consistent with energy failure, fission also presents two phases: primary fission was observed immediately after HI attack, followed by a secondary phase of fission at 24h following recovery (Nair et al., 2022). Mitophagy also has two phases: it was upregulated immediately after HI followed by a second wave after seven days (Nair et al., 2022). A prospective multicentric study reported that the neonatal plasma level of PARKIN (a protein involved in mitophagy) is related to HI insult and is reliable, also at birth (Tarocco et al., 2022). After lethal oxygen-glucose deprivation, overload of the Ca²⁺-stimulated activity of calpains mediated the degradation of cytoskeletal protein, causing abnormal mitochondrial distribution (Novorolsky et al., 2020).

Discussion

Although HIE is not a mitochondrial disease, the roles of mitochondria should be taken seriously. In this review, mitochondrial roles, including oxidative phosphorylation, mPTP, and dynamics, during and after deprivation of oxygen and glucose were reviewed. Mitochondrial functions are affected and altered by oxygen-glucose deprivation (OGD) after which dysfunctional mitochondria feedback to cells and cause further cell damage.

There are some treatments for HIE with effects on the function of mitochondria or target mitochondria. Moderate-severe HIE in term and near-term infants is treated by therapeutic hypothermia (TH), the only mandated therapy approved for term HIE, to reduce death and improve prognosis (Jacobs et al., 2013; Labat et al., 2022). TH helps cells to restore metabolic and cellular energy state homeostasis, in which mitochondria play an important role. Zhou et al. reported that mild hypothermia alleviates OGD-induced apoptosis by inhibiting ROS generation and improving mitochondrial dysfunction (Zhou et al., 2022). Sosunov et al. suggested that TH inhibited calcium-induced mitochondrial mPTP to protect neurons and reduce HI-induced injury (Sosunov et al., 2022).

Except for TH, there are no other approved therapies for HIE, however, many drugs have been tried in HIE. These drugs affect mitochondrial functions (Thornton et al., 2018; Rodriguez et al., 2020). Erythropoietin and melatonin can promote mitochondrial biogenesis (Carraway et al., 2010; Niu et al., 2020). Melatonin, a broad-spectrum antioxidant, also restores mitochondrial fusion/fission dynamics affected by OGD and enhances mitophagy (Kang et al., 2016; Nasoni et al., 2021). SkQR1 (Silachev et al., 2018) and MitoSNO (Kim et al., 2018) are other antioxidants that specifically abolish mitochondrial ROS to protect neurons. The rescue of mitochondrial function due to 2-Iminobiotin treatment by inhibition of neuronal and inducible isoforms of NOS is currently under development to reduce brain cell damage after HI injury (Zitta et al., 2017; Albrecht et al., 2019). Thiamine, in particular, has a critical role in oxidative phosphorylation and the TCA cycle in mitochondria; moreover, this micronutrient can regulate mitochondrial transition pore opening (Bâ, 2017; Sechi et al., 2022). An energy source supply alternative to glucose with normalization of aberrant cellular energy metabolism and decreased oxidative stress in ketogenic diets suggests a therapeutic role. Combined treatment with ketone bodies and thiamine may have a synergistic effect, further fostering the normalization of aberrant cellular energy metabolism (Sechi and Sechi, 2023).

Great efforts provided new visions of mitochondrial functions in HIE. These efforts attracted academic institutions and industries to provide more and more potential therapies for HIE. We will see the approved treatment in the near future.

Consent for publication. Not applicable.

Authors' contributions. KWJ collected and analyzed related articles, and then wrote the manuscript. LC collected related articles and drew figures. KWJ supervised data and manuscript. All authors read and approved the final manuscript.

References

Ethics approval and consent to participate. Not applicable.

Availability of data and materials. Data from patients are available from the corresponding author after discussion with the Institutional Review Board.

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Albrecht M., Zitta K., Groenendaal F., van Bel F. and Peeters-Scholte C. (2019). Neuroprotective strategies following perinatal hypoxia-

ischemia: Taking aim at NOS. Free Radic. Biol. Med. 142, 123-131.

- Andrieux P., Chevillard C., Cunha-Neto E. and Nunes J.P.S. (2021). Mitochondria as a Cellular Hub in Infection and Inflammation. Int. J. Mol. Sci. 22, 11338.
- Bâ A. (2017). Alcohol and thiamine deficiency trigger differential mitochondrial transition pore opening mediating cellular death. Apoptosis 22, 741-752.
- Baburamani A.A., Hurling C., Stolp H., Sobotka K., Gressens P., Hagberg H. and Thornton C. (2015). Mitochondrial Optic Atrophy (OPA) 1 processing is altered in response to neonatal hypoxicischemic brain injury. Int. J. Mol. Sci. 16, 22509-22526.
- Baughman J.M., Perocchi F., Girgis H.S., Plovanich M., Belcher-Timme C.A., Sancak Y., Bao X.R., Strittmatter L., Goldberger O., Bogorad R.L., Koteliansky V. and Mootha V.K. (2011). Integrative genomics identifies MCU as an essential component of the mitochondrial calcium uniporter. Nature 476, 341-345.
- Bernardi P. (2013). The mitochondrial permeability transition pore: a mystery solved? Front. Physiol. 4, 95.
- Brustovetsky N. and Dubinsky J.M. (2000). Limitations of cyclosporin A inhibition of the permeability transition in CNS mitochondria. J. Neurosci. 20, 8229-8237.
- Carraway M.S., Suliman H.B., Jones W.S., Chen C.W., Babiker A. and Piantadosi C.A. (2010). Erythropoietin activates mitochondrial biogenesis and couples red cell mass to mitochondrial mass in the heart. Circ. Res. 106, 1722-1730.
- Chandel N.S., Maltepe E., Goldwasser E., Mathieu C.E., Simon M.C. and Schumacker P.T. (1998). Mitochondrial reactive oxygen species trigger hypoxia-induced transcription. Proc. Natl. Acad. Sci. USA 95, 11715-11720.
- Chang A.H., Sancheti H., Garcia J., Kaplowitz N., Cadenas E. and Han D. (2014). Respiratory substrates regulate S-nitrosylation of mitochondrial proteins through a thiol-dependent pathway. Chem. Res. Toxicol. 27, 794-804.
- Chapa-Dubocq X., Makarov V. and Javadov S. (2018). Simple kinetic model of mitochondrial swelling in cardiac cells. J. Cell. Physiol. 233, 5310-5321.
- Chen I. and Lui F. (2022). Physiology, active transport. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing 2023.
- Chen J.J. and Yu B.P. (1994). Alterations in mitochondrial membrane fluidity by lipid peroxidation products. Free Radic. Biol. Med. 17, 411-418.
- Cooper C.E. and Giulivi C. (2007). Nitric oxide regulation of mitochondrial oxygen consumption II: Molecular mechanism and tissue physiology. Am. J. Physiol. Cell Physiol. 292, C1993-2003.
- Crompton M., Virji S. and Ward J.M. (1998). Cyclophilin-D binds strongly to complexes of the voltage-dependent anion channel and the adenine nucleotide translocase to form the permeability transition pore. Eur. J. Biochem. 258, 729-735.
- De Stefani D., Raffaello A., Teardo E., Szabo I. and Rizzuto R. (2011). A forty-kilodalton protein of the inner membrane is the mitochondrial calcium uniporter. Nature 476, 336-340.
- Demarest T.G., Waite E.L., Kristian T., Puche A.C., Waddell J., McKenna M.C. and Fiskum G. (2016). Sex-dependent mitophagy and neuronal death following rat neonatal hypoxia-ischemia. Neuroscience 335, 103-113.
- Dorn G.W. 2nd. (2020). Mitofusins as mitochondrial anchors and tethers. J. Mol. Cell. Cardiol. 142, 146-153.
- Erecinska M. and Silver I.A. (1989). ATP and brain function. J. Cereb. Blood Flow Metab. 9, 2-19.

- Fabiato A. and Fabiato F. (1975). Contractions induced by a calciumtriggered release of calcium from the sarcoplasmic reticulum of single skinned cardiac cells. J. Physiol. 249, 469-495.
- Fuhrmann D.C. and Brune B. (2017). Mitochondrial composition and function under the control of hypoxia. Redox Biol. 12, 208-215.
- Gilkerson R.W., Selker J.M. and Capaldi R.A. (2003). The cristal membrane of mitochondria is the principal site of oxidative phosphorylation. FEBS Lett. 546, 355-358.
- Goldhaber J.I. (1997). Metabolism in normal and ischemic myocardium. In: The myocardium. 2nd. Langer G.A. (ed). Academic Press. San Diego.
- Gou Z., Su X., Hu X., Zhou Y., Huang L., Fan Y., Li J. and Lu L. (2020). Melatonin improves hypoxic-ischemic brain damage through the Akt/Nrf2/Gpx4 signaling pathway. Brain Res. Bull. 163, 40-48.
- Guzy R.D. and Schumacker P.T. (2006). Oxygen sensing by mitochondria at complex III: the paradox of increased reactive oxygen species during hypoxia. Exp. Physiol. 91, 807-819.
- Haleckova A. and Benek O. (2022). Small-molecule inhibitors of cyclophilin D as potential therapeutics in mitochondria-related diseases. Med. Res. Rev. 42, 1822-1855.
- Halestrap A.P. (2009). What is the mitochondrial permeability transition pore? J. Mol. Cell. Cardiol. 46, 821-831.
- Halestrap A.P., Woodfield K.Y. and Connern C.P. (1997). Oxidative stress, thiol reagents, and membrane potential modulate the mitochondrial permeability transition by affecting nucleotide binding to the adenine nucleotide translocase. J. Biol. Chem. 272, 3346-3354.
- Ham P.B. 3rd. and Raju R. (2017). Mitochondrial function in hypoxic ischemic injury and influence of aging. Prog. Neurobiol. 157, 92-116.
- Hasselbach W. and Makinose M. (1962). ATP and active transport. Biochem. Biophys. Res. Commun. 7, 132-136.
- Haworth R.A. and Hunter D.R. (1979). The Ca²⁺-induced membrane transition in mitochondria. II. Nature of the Ca²⁺ trigger site. Arch. Biochem. Biophys. 195, 460-467.
- Hernansanz-Agustin P., Izquierdo-Alvarez A., Sanchez-Gomez F.J., Ramos E., Villa-Pina T., Lamas S., Bogdanova A. and Martinez-Ruiz A. (2014). Acute hypoxia produces a superoxide burst in cells. Free Radic. Biol. Med. 71, 146-156.
- Hernansanz-Agustin P., Ramos E., Navarro E., Parada E., Sanchez-Lopez N., Pelaez-Aguado L., Cabrera-Garcia J.D., Tello D., Buendia I., Marina A., Egea J., Lopez M.G., Bogdanova A. and Martinez-Ruiz A. (2017). Mitochondrial complex I deactivation is related to superoxide production in acute hypoxia. Redox Biol. 12, 1040-1051.
- Hicks M. and Gebicki J.M. (1978). A quantitative relationship between permeability and the degree of peroxidation in ufasome membranes. Biochem. Biophys. Res. Commun. 80, 704-708.
- Hopper R.K., Carroll S., Aponte A.M., Johnson D.T., French S., Shen R.F., Witzmann F.A., Harris R.A. and Balaban R.S. (2006). Mitochondrial matrix phosphoproteome: effect of extra mitochondrial calcium. Biochemistry 45, 2524-2536.
- Huttemann M., Helling S., Sanderson T.H., Sinkler C., Samavati L., Mahapatra G., Varughese A., Lu G., Liu J., Ramzan R., Vogt S., Grossman L.I., Doan J.W., Marcus K. and Lee I. (2012). Regulation of mitochondrial respiration and apoptosis through cell signaling: cytochrome c oxidase and cytochrome c in ischemia/reperfusion injury and inflammation. Biochim. Biophys. Acta 1817, 598-609.
- Huttemann M., Lee I., Pecinova A., Pecina P., Przyklenk K. and Doan J.W. (2008). Regulation of oxidative phosphorylation, the mitochondrial membrane potential, and their role in human disease.

J. Bioenerg. Biomembr. 40, 445-456.

- Iovino L., Tremblay M.E. and Civiero L. (2020). Glutamate-induced excitotoxicity in Parkinson's disease: The role of glial cells. J. Pharmacol. Sci. 144, 151-164.
- Jacobs S.E., Berg M., Hunt R., Tarnow-Mordi W.O., Inder T.E. and Davis P.G. (2013). Cooling for newborns with hypoxic ischaemic encephalopathy. Cochrane Database Syst. Rev. 2013, CD003311.
- Jones A. and Thornton C. (2022). Mitochondrial dynamics in the neonatal brain - a potential target following injury? Biosci. Rep. 42, BSR20211696.
- Kadenbach B. (2020). Regulation of cytochrome c oxidase contributes to health and optimal life. World J. Biol. Chem. 11, 52-61.
- Kang J.W., Hong J.M. and Lee S.M. (2016). Melatonin enhances mitophagy and mitochondrial biogenesis in rats with carbon tetrachloride-induced liver fibrosis. J. Pineal Res. 60, 383-393.
- Kim S.Y., Shim M.S., Kim K.Y., Weinreb R.N., Wheeler L.A. and Ju W.K. (2014). Inhibition of cyclophilin D by cyclosporin A promotes retinal ganglion cell survival by preventing mitochondrial alteration in ischemic injury. Cell Death Dis. 5, e1105.
- Kim M., Stepanova A., Niatsetskaya Z., Sosunov S., Arndt S., Murphy M.P., Galkin A. and Ten V.S. (2018). Attenuation of oxidative damage by targeting mitochondrial complex I in neonatal hypoxicischemic brain injury. Free Radic. Biol. Med. 124, 517-524.
- Kratimenos P., Koutroulis I., Agarwal B., Theocharis S. and Delivoria-Papadopoulos M. (2017). Effect of Src kinase inhibition on cytochrome c, Smac/DIABLO and Apoptosis Inducing Factor (AIF) following cerebral hypoxia-ischemia in newborn piglets. Sci. Rep. 7, 16664.
- Kurinczuk J.J., White-Koning M. and Badawi N. (2010). Epidemiology of neonatal encephalopathy and hypoxic-ischaemic encephalopathy. Early Hum. Dev. 86, 329-338.
- Kwon H.S. and Koh S.H. (2020). Neuroinflammation in neurodegenerative disorders: the roles of microglia and astrocytes. Transl. Neurodegener. 9, 42.
- Labat J., Brocard C., Belaroussi Y., Bar C., Gotchac J., Chateil J.F. and Brissaud O. (2022). Hypothermia for neonatal hypoxic-ischemic encephalopathy: Retrospective descriptive study of features associated with poor outcome. Arch. Pediatr. 30, 93-99.
- Li Y., Wang M. and Wang S. (2016). Effect of inhibiting mitochondrial fission on energy metabolism in rat hippocampal neurons during ischemia/reperfusion injury. Neurol. Res. 38, 1027-1034.
- Li T., Li K., Zhang S., Wang Y., Xu Y., Cronin S.J.F., Sun Y., Zhang Y., Xie C., Rodriguez J., Zhou K., Hagberg H., Mallard C., Wang X., Penninger J.M., Kroemer G., Blomgren K. and Zhu C. (2020). Overexpression of apoptosis inducing factor aggravates hypoxicischemic brain injury in neonatal mice. Cell Death Dis. 11, 77.
- Liu S.S. (1999). Cooperation of a "reactive oxygen cycle" with the Q cycle and the proton cycle in the respiratory chain--superoxide generating and cycling mechanisms in mitochondria. J. Bioenerg. Biomembr. 31, 367-376.
- Liu S.S. (2010). Mitochondrial Q cycle-derived superoxide and chemiosmotic bioenergetics. Ann. N.Y. Acad. Sci. 1201, 84-95.
- Lorek A., Takei Y., Cady E.B., Wyatt J.S., Penrice J., Edwards A.D., Peebles D., Wylezinska M., Owen-Reece H., Kirkbride V., Cooper C.E., Aldridge R.F., Roth S.C., Brown G., Delpy D.T. and Reynolds E.O.R. (1994). Delayed ("secondary") cerebral energy failure after acute hypoxia-ischemia in the newborn piglet: continuous 48-hour studies by phosphorus magnetic resonance spectroscopy. Pediatr. Res. 36, 699-706.

- Mahmoud S., Gharagozloo M., Simard C. and Gris D. (2019). Astrocytes maintain glutamate homeostasis in the CNS by controlling the balance between glutamate uptake and release. Cells 8, 184.
- Marzo I., Brenner C., Zamzami N., Jurgensmeier J.M., Susin S.A., Vieira H.L., Prevost M.C., Xie Z., Matsuyama S., Reed J.C. and Kroemer G. (1998). Bax and adenine nucleotide translocator cooperate in the mitochondrial control of apoptosis. Science 281, 2027-2031.
- Mears J.A. and Ramachandran R. (2022). Drp1 and the cytoskeleton: mechanistic nexus in mitochondrial division. Curr. Opin. Physiol. 29, 100574.
- Moller M.N., Rios N., Trujillo M., Radi R., Denicola A. and Alvarez B. (2019). Detection and quantification of nitric oxide-derived oxidants in biological systems. J. Biol. Chem. 294, 14776-802.
- Morciano G., Bonora M., Campo G., Aquila G., Rizzo P., Giorgi C., Wieckowski M.R. and Pinton P. (2017). Mechanistic Role of mPTP in Ischemia-Reperfusion Injury. Adv. Exp. Med. Biol. 982, 169-189.
- Nair S., Leverin A.L., Rocha-Ferreira E., Sobotka K.S., Thornton C., Mallard C. and Hagberg H. (2022). Induction of mitochondrial fragmentation and mitophagy after neonatal hypoxia-ischemia. Cells 11, 1193.
- Nasoni M.G., Carloni S., Canonico B., Burattini S., Cesarini E., Papa S., Pagliarini M., Ambrogini P., Balduini W. and Luchetti F. (2021). Melatonin reshapes the mitochondrial network and promotes intercellular mitochondrial transfer via tunneling nanotubes after ischemic-like injury in hippocampal HT22 cells. J. Pineal Res. 71, e12747.
- Niu Y.J., Zhou W., Nie Z.W., Shin K.T. and Cui X.S. (2020). Melatonin enhances mitochondrial biogenesis and protects against rotenoneinduced mitochondrial deficiency in early porcine embryos. J. Pineal Res. 68, e12627.
- Novorolsky R.J., Nichols M., Kim J.S., Pavlov E.V., J J.W., Wilson J.J. and Robertson G.S. (2020). The cell-permeable mitochondrial calcium uniporter inhibitor Ru265 preserves cortical neuron respiration after lethal oxygen glucose deprivation and reduces hypoxic/ischemic brain injury. J. Cereb. Blood Flow Metab. 40, 1172-1181.
- Patel P., Mendoza A., Robichaux D.J., Wang M.C., Wehrens X.H.T. and Karch J. (2021). Inhibition of the anti-apoptotic Bcl-2 family by BH3 mimetics sensitize the mitochondrial permeability transition pore through bax and bak. Front. Cell Dev. Biol. 9, 765973.
- Pedroza-Garcia K.A., Calderon-Vallejo D. and Quintanar J.L. (2022). Neonatal hypoxic-ischemic encephalopathy: Perspectives of neuroprotective and neuroregenerative treatments. Neuropediatrics 53, 402-417.
- Popov L.D. (2020). Mitochondrial biogenesis: An update. J. Cell Mol. Med. 24, 4892-4899.
- Puka-Sundvall M., Gajkowska B., Cholewinski M., Blomgren K., Lazarewicz J.W. and Hagberg H. (2000). Subcellular distribution of calcium and ultrastructural changes after cerebral hypoxia-ischemia in immature rats. Brain Res. Dev. Brain Res. 125, 31-41.
- Puka-Sundvall M., Gilland E. and Hagberg H. (2001). Cerebral hypoxiaischemia in immature rats: involvement of mitochondrial permeability transition? Dev. Neurosci. 23, 192-197.
- Quarato G., Llambi F., Guy C.S., Min J., Actis M., Sun H., Narina S., Pruett-Miller S.M., Peng J., Rankovic Z. and Green D.R. (2022). Ca²⁺-mediated mitochondrial inner membrane permeabilization induces cell death independently of Bax and Bak. Cell Death Differ.

29, 1318-1334.

- Queensland-Clinical-Guidelines. (2021). Hypoxic ischaemic encephalopathy (HIE).
- Quintana D.D., Garcia J.A., Sarkar S.N., Jun S., Engler-Chiurazzi E.B., Russell A.E., Cavendish J.Z. and Simpkins J.W. (2019). Hypoxiareoxygenation of primary astrocytes results in a redistribution of mitochondrial size and mitophagy. Mitochondrion 47, 244-255.
- Radi R., Cassina A. and Hodara R. (2002). Nitric oxide and peroxynitrite interactions with mitochondria. Biol. Chem. 383, 401-409.
- Robichaux D.J., Harata M., Murphy E. and Karch J. (2022). Mitochondrial permeability transition pore-dependent necrosis. J. Mol. Cell Cardiol. 174, 47-55.
- Rodriguez M., Valez V., Cimarra C., Blasina F. and Radi R. (2020). Hypoxic-ischemic encephalopathy and mitochondrial dysfunction: Facts, unknowns, and challenges. Antioxid. Redox Signal. 33, 247-262.
- Rodriguez-Gomez J.A., Kavanagh E., Engskog-Vlachos P., Engskog M.K.R., Herrera A.J., Espinosa-Oliva A.M., Joseph B., Hajji N., Venero J.L. and Burguillos M.A. (2020). Microglia: Agents of the CNS pro-inflammatory response. Cells 9, 1717.
- Ruegger C.M., Davis P.G. and Cheong J.L. (2018). Xenon as an adjuvant to therapeutic hypothermia in near-term and term newborns with hypoxic-ischaemic encephalopathy. Cochrane Database Syst. Rev. 8, CD012753.
- Sander P., Gudermann T. and Schredelseker J. (2021). A calcium guard in the outer membrane: Is VDAC a regulated gatekeeper of mitochondrial calcium uptake? Int. J. Mol. Sci. 22, 946.
- Schroedl C., McClintock D.S., Budinger G.R. and Chandel N.S. (2002). Hypoxic but not anoxic stabilization of HIF-1alpha requires mitochondrial reactive oxygen species. Am. J. Physiol. Lung Cell. Mol. Physiol. 283, L922-931.
- Sechi G. and Sechi M.M. (2023). New therapeutic paradigms in neonatal hypoxic-ischemic encephalopathy. ACS Chem. Neurosci. 14, 1004-1006.
- Sechi G.P., Bardanzellu F., Pintus M.C., Sechi M.M., Marcialis M.A. and Fanos V. (2022). Thiamine as a possible neuroprotective strategy in neonatal hypoxic-ischemic encephalopathy. Antioxidants (Basel). 11, 42.
- Sica V., Izzo V., Bravo-San Pedro J.M., Zamzami N. and Maiuri M.C. (2016). Chapter 3 - Mitophagy: Sensors, regulators, and effectors. In: Autophagy: Cancer, other pathologies, inflammation, immunity, infection, and aging. M. A. Hayat. Academic Press.
- Silachev D.N., Plotnikov E.Y., Pevzner I.B., Zorova L.D., Balakireva A.V., Gulyaev M.V., Pirogov Y.A., Skulachev V.P. and Zorov D.B. (2018). Neuroprotective effects of mitochondria-targeted plastoquinone in a rat model of neonatal hypoxic-lschemic brain injury. Molecules 23, 1871.
- Sosunov S., Bhutada A., Niatsetskaya Z., Starkov A. and Ten V. (2022). Mitochondrial calcium buffering depends upon temperature and is associated with hypothermic neuroprotection against hypoxiaischemia injury. PLoS One 17, e0273677.
- Starkov A.A. and Fiskum G. (2003). Regulation of brain mitochondrial H₂O₂ production by membrane potential and NAD(P)H redox state. J. Neurochem. 86, 1101-1107.
- Tarocco A., Morciano G., Perrone M., Cafolla C., Ferre C., Vacca T., Pistocchi G., Meneghin F., Cocchi I., Lista G., Cetin I., Greco P., Garani G., Stella M., Natile M., Ancora G., Savarese I., Campi F., Bersani I., Dotta A., Tiberi E., Vento G., Chiodin E., Staffler A., Maranella E., Di Fabio S., Wieckowski M.R., Giorgi C. and Pinton P.

(2022). Increase of Parkin and ATG5 plasmatic levels following perinatal hypoxic-ischemic encephalopathy. Sci. Rep. 12, 7795.

- Thornton C., Jones A., Nair S., Aabdien A., Mallard C. and Hagberg H. (2018). Mitochondrial dynamics, mitophagy and biogenesis in neonatal hypoxic-ischaemic brain injury. FEBS Lett. 592, 812-830.
- Torres J., Cooper C.E. and Wilson M.T. (1998). A common mechanism for the interaction of nitric oxide with the oxidized binuclear centre and oxygen intermediates of cytochrome c oxidase. J. Biol. Chem. 273, 8756-8766.
- Tretter L. and Adam-Vizi V. (2004). Generation of reactive oxygen species in the reaction catalyzed by alpha-ketoglutarate dehydrogenase. J. Neurosci. 24, 7771-7778.
- Turrens J.F. (2003). Mitochondrial formation of reactive oxygen species. J. Physiol. 552, 335-344.
- Vannucci R.C., Brucklacher R.M. and Vannucci S.J. (2005). Glycolysis and perinatal hypoxic-ischemic brain damage. Dev. Neurosci. 27, 185-190.
- Velaphi S. and Pattinson R. (2007). Avoidable factors and causes of neonatal deaths from perinatal asphyxia-hypoxia in South Africa: national perinatal survey. Ann. Trop. Paediatr. 27, 99-106.
- Ventura-Clapier R., Garnier A. and Veksler V. (2008). Transcriptional control of mitochondrial biogenesis: the central role of PGC-1alpha. Cardiovasc. Res. 79, 208-217.
- Verklan T. and Walden M. (2015). Hypoxic-ischaemic encephalopathy. In: Core curriculm for neonatal intensive care. Verklan M.T. and Walden M. (eds). Elsevier. Saint Louis.
- Vranic S., Rodrigues A.F., Buggio M., Newman L., White M.R.H., Spiller D.G., Bussy C. and Kostarelos K. (2018). Live imaging of label-free graphene oxide reveals critical factors causing oxidative-stressmediated cellular responses. ACS Nano 12, 1373-1389.
- Wang X., Carlsson Y., Basso E., Zhu C., Rousset C.I., Rasola A., Johansson B.R., Blomgren K., Mallard C., Bernardi P., Forte M.A. and Hagberg H. (2009). Developmental shift of cyclophilin D contribution to hypoxic-ischemic brain injury. J. Neurosci. 29, 2588-2596.
- Wang X., Han W., Du X., Zhu C., Carlsson Y., Mallard C., Jacotot E. and Hagberg H. (2010). Neuroprotective effect of Bax-inhibiting peptide on neonatal brain injury. Stroke 41, 2050-205.
- Wassink G., Davidson J.O., Dhillon S.K., Zhou K., Bennet L., Thoresen M. and Gunn A.J. (2019). Therapeutic hypothermia in neonatal hypoxicischemic encephalopathy. Curr. Neurol. Neurosci. Rep. 19, 2.
- Widgerow A.D. (2014). Ischemia-reperfusion injury: influencing the microcirculatory and cellular environment. Ann. Plast. Surg. 72, 253-260.
- Woodfield K., Ruck A., Brdiczka D. and Halestrap A.P. (1998). Direct demonstration of a specific interaction between cyclophilin-D and the adenine nucleotide translocase confirms their role in the mitochondrial permeability transition. Biochem J. 336, 287-290.
- Wu Y.W., Comstock B.A., Gonzalez F.F., Mayock D.E., Goodman A.M., Maitre N.L., Chang T., Van Meurs K.P., Lampland A.L., Bendel-Stenzel E., Mathur A.M., Wu T.W., Riley D., Mietzsch U., Chalak L., Flibotte J., Weitkamp J.H., Ahmad K.A., Yanowitz T.D., Baserga M., Poindexter B.B., Rogers E.E., Lowe J.R., Kuban K.C.K., O'Shea T.M., Wisnowski J.L., McKinstry R.C., Bluml S., Bonifacio S., Benninger K.L., Rao R., Smyser C.D., Sokol G.M., Merhar S., Schreiber M.D., Glass H.C., Heagerty P.J., Juul S.E. and Consortium H. (2022). Trial of erythropoietin for hypoxic-ischemic encephalopathy in newborns. N. Engl. J. Med. 387, 148-159.
- Yapa N.M.B., Lisnyak V., Reljic B. and Ryan M.T. (2021). Mitochondrial

dynamics in health and disease. FEBS Lett. 595, 1184-1204.

- Youle R.J. and van der Bliek A.M. (2012). Mitochondrial fission, fusion, and stress. Science 337, 1062-105.
- Yusupov M., Van der Paal J., Neyts E.C. and Bogaerts A. (2017a). Synergistic effect of electric field and lipid oxidation on the permeability of cell membranes. Biochim. Biophys. Acta Gen. Subj. 1861, 839-847.
- Yusupov M., Wende K., Kupsch S., Neyts E.C., Reuter S. and Bogaerts A. (2017b). Effect of head group and lipid tail oxidation in the cell membrane revealed through integrated simulations and experiments. Sci. Rep. 7, 5761.
- Zaidan E. and Sims N.R. (1994). The calcium content of mitochondria from brain subregions following short-term forebrain ischemia and recirculation in the rat. J. Neurochem. 63, 1812-1819.
- Zhao M., Zhu P., Fujino M., Zhuang J., Guo H., Sheikh I., Zhao L. and Li X.K. (2016). Oxidative stress in hypoxic-ischemic encephalopathy: Molecular mechanisms and therapeutic strategies. Int. J. Mol. Sci. 17, 2078.
- Zhou K., Chen J., Wu J., Wu Q., Jia C., Xu Y.X.Z., Chen L., Tu W., Yang G., Kong J., Kou J. and Jiang S. (2019). Atractylenolide III

ameliorates cerebral ischemic injury and neuroinflammation associated with inhibiting JAK2/STAT3/Drp1-dependent mitochondrial fission in microglia. Phytomedicine 59, 152922.

- Zhou T., Mo J., Xu W., Hu Q., Liu H., Fu Y. and Jiang J. (2022). Mild hypothermia alleviates oxygen-glucose deprivation/reperfusioninduced apoptosis by inhibiting ROS generation, improving mitochondrial dysfunction and regulating DNA damage repair pathway in PC12 cells. Apoptosis 28, 447-457.
- Zitta K., Peeters-Scholte C., Sommer L., Gruenewald M., Hummitzsch L., Parczany K., Steinfath M. and Albrecht M. (2017). 2-Iminobiotin superimposed on hypothermia protects human neuronal cells from hypoxia-induced cell damage: An *in Vitro* Study. Front. Pharmacol. 8, 971.
- Zorov D.B., Filburn C.R., Klotz L.O., Zweier J.L. and Sollott S.J. (2000). Reactive oxygen species (ROS)-induced ROS release: a new phenomenon accompanying induction of the mitochondrial permeability transition in cardiac myocytes. J. Exp. Med. 192, 1001-1014.

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