- Density assessment and reporting for *Phlebotomus perniciosus* and other sand fly
 species in periurban residential estates in Spain
- 3

C. Muñoz^{1a*}, J. Risueño^{1*}, P. Pérez-Cutillas², L.J. Bernal³, J.M. Ortiz¹, R. Ruiz de Ybáñez¹, 4 P.F. Sánchez-López⁴, C. Martínez-Carrasco¹, L. Del Río¹, P. De la Rúa⁵, J.D. García-5 Martínez³, M. Gonzálvez¹, L. Murcia⁶, F. Collantes⁷, E. Goyena¹, T. Spitzova⁸, S. Elshanat⁹ 6 and E. Berriatua¹ 7 8 ¹Departamento de Sanidad Animal, Facultad de Veterinaria, Regional Campus of 9 10 International Excellence 'Campus Mare Nostrum', Universidad de Murcia, Murcia, Spain. ²Departamento de Geografía, Universidad de Murcia, Murcia, Spain. 11 ³Departamento de Medicina y Cirugía Animal, Facultad de Veterinaria, Regional Campus 12 of International Excellence 'Campus Mare Nostrum', Universidad de Murcia, Murcia, 13 Spain. 14 ⁴Servicio de Sanidad Ambiental, Dirección General de Salud Pública y Adicciones, 15 Consejería de Salud de la Región de Murcia, Murcia, Spain. 16 ⁵Área de Biología Animal, Departamento de Zoología y Antropología Física, Facultad de 17 Veterinaria, Regional Campus of International Excellence 'Campus Mare Nostrum', 18 Universidad de Murcia, Murcia, Spain. 19 ⁶Departamento de Genética y Microbiología, Facultad de Medicina, Regional Campus of 20 21 International Excellence "Campus Mare Nostrum", Universidad de Murcia, Murcia, Spain. 22 ⁷Departamento de Zoología y Antropología Física, Facultad de Biología, Regional Campus of International Excellence 'Campus Mare Nostrum', Universidad de Murcia, Murcia, 23 Spain. 24

25	⁸ Department of Parasitology, Faculty of Science, Charles University, Vinicna 7, 128 44
26	Prague 2, Czech Republic.
27	⁹ Department of Parasitology, Faculty of veterinary medicine, Alexandria University, Egypt.
28	
29	*The first two authors contributed equally to this work.
30	
31	^α Corresponding author: Clara Muñoz. Departamento de Sanidad Animal, Facultad de
32	Veterinaria, Campus de Espinardo, Universidad de Murcia, 30100 Murcia, Spain. Tel.:
33	0034 618972250. E-mail: clara.munoz1@um.es
34	
35	ORCID ID: Clara Muñoz (0000-0001-9847-8616), José Risueño (0000-0001-5006-7738),
36	Pedro Pérez-Cutillas (0000-0003-1271-3895), Luis Jesús Bernal (0000-0002-3648-7012),
37	Juana María Ortiz (0000-0002-0889-2985), Rocío Ruiz de Ybáñez (0000-0003-1402-8023),
38	Pedro F. Sánchez-López (0000-0003-1712-2576), Carlos Martínez-Carrasco (0000-0002-
39	8742-0109), Laura del Río (0000-0003-4719-8926), Pilar de la Rúa (0000-0002-0058-
40	1402), Juan Diego García-Martínez (0000-0003-4209-0107), Moisés Gonzálvez (0000-
41	0003-0423-3189), Laura Murcia (0000-0001-7768-0989), Francisco Collantes (0000-0002-
42	8085-6399), Elena Goyena (0000-0001-8993-2792), Tatiana Spitzova (0000-0003-2028-
43	3530), Sherif Elshanat (0000-0003-1649-3228), Eduardo Berriatua (0000-0002-0721-8091).
44	
45	Acknowledgements

We are grateful to house owners that participated in the study allowing researchers tomonitor their premises over a long time.

48 Abstract

Green periurban residential areas in Mediterranean countries have flourished in the last 49 decades and become foci for leishmaniasis. To remedy the absence of information on 50 vector ecology in these environments, we examined Phlebotomine sand fly distribution in 51 52 29 sites in Murcia City over a three year period, including the plots of 20 detached houses and nine non-urbanized sites nearby. We collected 5,066 specimens from five species using 53 "sticky" interception and light attraction traps. The relative frequency of the main 54 55 Leishmania infantum vector Phlebotomus perniciosus in these traps was 32% and 63%, respectively. Sand fly density was widely variable spatially and temporally, and greatest in 56 non-urbanized sites, particularly in caves and abandoned buildings close to domestic animal 57 58 holdings. Phlebotomus perniciosus density in house plots was positively correlated with those in non-urbanized sites, greatest in larger properties with extensive vegetation and 59 non-permanently lived, but not associated to dog presence or a history of canine 60 leishmaniasis. Within house plots, sand fly density was highest in traps closest to walls. 61 62 Furthermore, the study provides a guideline for insect-density assessment and reporting and is envisioned as a building block towards the development of a pan-European data-base for 63 robust investigation of environmental determinants of sand fly distribution. 64

65

66

Keywords: Phlebotomus, distribution, density, environment, residential, Leishmania

Phlebotomine sand flies (Diptera: Psychodidae) are endemic in tropical and subtropical 68 latitudes as well as the Mediterranean subregion, where they are vectors of life-threatening 69 Leishmania spp. (Kinetoplastida: Trypanosomatidae) and arboviruses (Phlebovirus, 70 Vesiculovirus and Orbivirus) (Akhoundi et al. 2016). Leishmania infantum is the only 71 endemic species in Spain and it causes zoonotic visceral leishmaniasis, a major disease of 72 dogs with a considerable public health impact (Herrador et al. 2015; Gálvez et al. 2020). 73 74 Phleboviruses identified in Spain include Toscana, Granada, Naples, Sicily, Arbia and Arrabida-like viruses, and the risk of infection is considered moderate for Toscana virus 75 and low for the other viruses (García San Miguel et al. 2020). Among the twelve sand fly 76 77 species described in Spain (Gil Collado et al. 1989; Martínez Ortega et al. 1992), Phlebotomus perniciosus and Phlebotomus ariasi are vectors of L. infantum and the former 78 79 is the predominant species in southeast Spain (Risueño et al. 2017). Sand flies are 80 considered to have low specificity for Phleboviruses (Ayhan and Charrel 2017) and six 81 viral isolates were detected in P. perniciosus in a human leishmaniasis outbreak in a residential area in the outskirts of Madrid (Arce et al. 2013; Remoli et al. 2016). This 82 unprecedented outbreak highlights the potential risks of leishmaniasis associated with 83 environmental changes in the natural environment of sand fly vectors. It was the result of 84 85 housing developments in former agricultural land leading to a massive buildup of P. perniciosus, coinciding with a demographic explosion of leporids (Lepus granatensis and 86 Oryctolagus cuniculus) on which they fed, and behaved as an unusual reservoir of L. 87 88 infantum (Molina et al. 2012; Arce et al. 2013). Human and canine leishmaniasis (CanL) are also emerging infections in modern residential estates built in the outskirts of cities, 89

90 consisting of detached and semidetached family houses with a garden and dogs (Pérez-91 Cutillas et al. 2015; Goyena et al. 2016), which are the main domestic reservoir of L. infantum (WHO 2010). It is logical to assume that these places provide ideal environments 92 for sand fly vectors including sites protected from desiccation and with abundant organic 93 material for sand flies to breed and rest, as well as a close-by source of blood required by 94 females for egg development (Alexander, 2000). However, the precise locations are not 95 96 well characterized and to the best of our knowledge, there are no studies describing vector 97 density and its relationship with environmental variables from these periurban settings.

98 Field investigations aiming for a representative picture of sand fly density and diversity in a 99 particular area are expensive and difficult to perform. Their distribution is seasonal and highly heterogeneous at fine geographical scales, requiring a large sampling effort (Rioux 100 101 et al. 2013; Muñoz et al. 2018, 2019). Moreover, there is no universal methodological guideline for estimating and reporting sand fly density -i.e. sand fly numbers in relation to 102 sampling effort. In this sense, entomologists make use of a wide variety of trapping 103 methods and protocols; results are often difficult or impossible to compare across studies; 104 and published data may not be sufficient for other researchers to perform wider-scale 105 quantitative analysis. The first objective of the present study is to provide an insight into 106 107 the spatial distribution of *P. perniciosus* and other sand fly species in periurban residential 108 properties located in the outskirts of Murcia City (southeast Spain). Our second objective 109 includes a proposal about the type of data that should be reported in a scientific journal to 110 allow meta-analytic investigations of the environmental factors that affect sand fly temporal and spatial distribution. Such investigations are essential to improve our understanding and 111 capacity infections 112 to prevent and control vector-borne

113 (https://www.ecdc.europa.eu/en/about-us/partnerships-and-networks/disease-and-

114 laboratory-networks/vector-net).

115

116 **2. Materials and Methods**

117 **2.1. Study area and design**

118 Murcia City has 453,000 inhabitants and is located in a region endemic for sand flies 119 and sand fly-borne infections, both in dogs and humans (Martínez-García et al. 2007; 120 Pérez-Cutillas et al. 2015; Goyena et al. 2016; Muñoz et al. 2019). Sand fly sampling was 121 performed in 29 sites in 13 housing estates. They were located four to 14 km from the City 122 center, except one site which was 26 km away. Sampling was performed in the summers of 123 2013, 2014 and 2015 during 25 weeks (see below). Sites included the outside plots (mainly 124 gardens) of 20 detached houses and nine non-urbanized sites situated in the periphery of the housing estates (Fig. 1; Table 1). The latter were included to monitor the degree of sand fly 125 threat to which the housing estates were potentially exposed. Due to limited resources it 126 127 was not possible to sample non-urbanized sites in every estate, and those selected were a 128 representative example of the non-urbanized landscape in this part of Murcia. Houses were 129 conveniently selected as they belonged to the research team families and other trustworthy 130 people, in an attempt to ensure long-term adherence to the study. Eleven houses had one or more dogs, four had had CanL cases in the previous five years and all participants knew of 131 132 CanL cases in the neighborhood.

Sites were georeferenced using a submetric GPS (Trimble geoXH), using a differential
correction through RINEX files provided by GNSS services close to the study area.
Environmental temperature and relative humidity and wind speed were obtained to analyze

its relationship with sand fly density as described below. Other factors analyzed, considered 136 137 potentially associated with sand fly density were the number and age of human and animal occupants in the house, house and plot sizes (vegetated and paved areas) and the presence 138 of a swimming pool. Trap location features were also recorded, including orientation, sun, 139 140 wind exposure (as presumed by owners), ground type, roof cover and distance to the closest 141 wall, firewood and stone piles, plants, water tab, stationary water and irrigation point, and 142 presence of a dog house. The most common plants situated in the proximity of the trap were ivy, a variety of lawn grasses, citrus fruit trees, cypresses, Mediterranean pine and 143 bougainvillea. Shrubs were the predominant vegetation in non-urbanized sites including 144 145 large extensions of rosemary and thyme.

146 **2.2. Sampling protocol and trap types**

Sand flies were sampled for 25 weeks in four periods between September 2013 and July 2015. First period: from 3rd week of September to 2nd week of October 2013; second period: from 4th week of May to 2nd week of July 2014; third period: from 2nd week in September to 2nd week in October 2014; fourth period: from 4th week in May to 4th week in July 2015. The number of sampling weeks varied between sites (Table 1), mainly due to volunteers dropping out before the study ended for personal reasons.

Interception traps, made of half an A4 sheet of tracing paper measuring 210 mm x 148.5 mm (except on very few occasions when entire A4 sheets were used) impregnated with castor oil ("sticky traps"), were used throughout the study. The number of traps varied between 6 and 14 traps per site, with the aim of covering all potential main sand fly microhabitats in selected sites. Traps were individually identified, placed always in the same spot close to the ground, exposing both sides of the sheets to sand flies, and they were 159 kept untouched for an average of 7 days/week (range: 4-14 days/week) and 94% of traps
160 between 6 and 8 days/week.

Battery-operated, miniature Centers for Disease Control (CDC) light attraction traps (J. W. Hock Company, Gainesville, FL, U.S.A.) were also used fortnightly in 18 house plots (one trap per house plot) in June and July 2015, making a total of two to five nights (from 8 am to 8 pm) in each of those plots. The aim was to compare species diversity detected in both trap types. Traps were placed close to vegetation and a wall, at approximately 1.5 m from the ground to avoid dog and cat interference.

167 **2.3. Sand fly identification**

Sticky traps were collected and stored individually between two A4 paper sheets each 168 169 and kept at 4°C until sand flies were removed from the traps using a fine brush dipped in 70% ethanol. Collection cups from light traps were kept at -20° C for at least 2 h to kill the 170 171 insects. Specimens from sticky and light traps were then maintained in 70% ethanol until 172 morphologically identified based on the external genitalia in males, and the pharynx, 173 cibarium and spermatheca in females (Martínez-Ortega and Conesa-Gallego 1987; Gállego-Berenguer et al. 1992). Male and female specimen preparation consisted of the dissection 174 of the head and two terminal abdominal segments, clarification in Marc-Andre solution, 175 176 and mounting on a glass slide using Hoyer's medium. Slides were examined under the 177 microscope at x400 magnification.

178

2.4. Altitude and climatic data collection

Site altitude was obtained from the high resolution (1 meter per pixel) Digital Elevation
Model of the LIDAR project from the "Plan Nacional de Ortofotografía Aérea (PNOA)"
(https://pnoa.ign.es) and ranged from 23 to 287 m above sea level (a.s.l.). Climatic real-

182 time data was collected from ten close-range meteorological stations (http://siam.imida.es) 183 (Fig. 1). They included the mean temperature and relative humidity (RH %) and the mean and maximum wind speed. ArcGIS v.10 (ESRI, Redlands, USA) was used to produce 184 continuous map layers of these variables with values from the nightly periods (20.00 hrs. to 185 8.00 hrs.) when adult sand flies are most active and with a spatial resolution of 5 m/pixel. 186 187 Site 29, situated far from the other sites, was excluded from this analysis. Wind speed was 188 estimated using the inverse weighted distance interpolation method (Keskin et al. 2015). Temperature and RH layers were developed employing a linear regression model using 189 residual-corrected altitude as the independent, explanatory variable. The validity of the 190 191 estimated temperature and RH % was assessed by comparing the 2015 data with similar measurements taken at the same time from thermohygrometers (Digital Logtag Haxo-8 T, 192 193 Templyzer) placed in 18 sampling sites, using Pearson's correlation coefficient.

194 **2.5. Data description and statistical analysis**

195 Abundance referred to the absolute number of sand flies, species richness was the number of different species and species diversity considered both the number of different 196 species and their relative frequency. "Positive traps" were those with at least one sand fly. 197 "Sand fly density" was defined as the number of sand flies collected divided by the 198 sampling effort. The "sampling effort" was established as the number of trapping days 199 multiplied by the trap area (m^2) for sticky traps and by the number of traps in the case of 200 light traps. The proportion of positive traps and median sand fly densities across 201 202 environmental explanatory variables were compared using Yates-corrected chi-squared test 203 (or when necessary Fisher exact test) and Kruskal-Wallis test, respectively. Spearman'srank coefficient test for non-normally distributed data was employed to evaluate the 204

205 correlation between sand fly density in sticky and light traps, and between house plots and206 non-urbanized sites situated within 500 m.

Separate multilevel negative binomial regression for overdispersed count data, were 207 developed to investigate site- and trap-level factors associated with *P. perniciosus* density 208 209 in sticky traps, respectively. These models allowed an assessment of correlation in the 210 insect's density from repeated sampling of the same sites and trap locations over time (two-211 level hierarchical models with weeks as level-1 and sites or traps as level-2 random variables) (Snijders and Bosker 1999). To avoid potential bias resulting from analyzing 212 data from sites sampled during different periods, modeling was performed in two data 213 214 subsets. Subset "a" included the five weeks in September and October 2014 when all except three sites were continuously sampled, and subset "b" was all data from house plot 215 sites 2, 3, 4, 22 and 28, which were the only ones sampled almost every week throughout 216 the study and had a moderately high sand fly density. Subset "a" was used to investigate 217 site-specific explanatory variables (climatic and others) that were associated with P. 218 perniciosus density in the bivariate analysis, and included site and weeks as random effects. 219 With subset "b" we investigated trap-level variables and which together with climatic 220 variables were incorporated in the model as fixed effects, and trap and weeks as random 221 222 variables. In all cases, the decimal logarithm of the specie's density+1 was the outcome 223 variable and a backward model building strategy was used including all fixed explanatory 224 variables. Since some environmental variables were strongly correlated, for example the 225 highest altitude range included only small house-plots, and none of the largest house plots were permanently inhabited, Akaikes Information Criteria was used to select models with 226 different combinations of variables, choosing those with the lowest value (Kleinbaum et al. 227 1998). Models were estimated using the maximum likelihood method using the glmer.nb 228

function in the lme4 package in R (https://cran.r-project.org/web/packages/lme4/lme4.pdf)
(Bates et al. 2015). For all analysis significance was confirmed when α=5% (p<0.05) for a
two-tailed test.

232

233 **3. Results**

234 **3.1. Sampling effort and sand fly abundance and density**

A total of 3,498 sticky traps were placed for an average of 7 days/trap in 29 sites over 25 235 236 weeks between September 2013 and July 2015, and 3,328 (95%) traps were recovered, 237 totaling 208.7 m² of sticky trap surface. The resulting sampling effort was 1,471 m² x days. Since 4,586 sand flies were collected in the sticky traps, with 31% of them being positive, 238 the overall density was 3.1 sand flies/m²/day (smd) (Table 1). However, there were large 239 240 differences between sites in all of these parameters. Sampling efforts ranged from 6.6 $m^2 x$ days in site 23 to 124.5 m^2 x days in site 2, the percentage of positive traps went from 0% 241 242 in site 26 to 93% in site 1; and sand fly density in the 28 positive sites ranged from 0.1 smd 243 in sites 15 and 25, to 23.2 smd in site 1 (Table 1). Globally, the percentage of positive traps 244 and sand fly density was significantly higher in non-urbanized areas compared to house plots, and there was a moderate correlation between sand fly density in non-urbanized areas 245 246 and house plots situated within 500 m (rho=0.45, p<0.05).

247 CDC light traps placed in June and July 2015 in 18 sites represented 67 trap-nights and 248 collected 480 sand flies in 84% of the traps. Hence, sand fly density was 7.2 sand 249 flies/trap/night (stn) overall, and ranged between 0 stn and 29.5 stn in sites 25 and 18, 250 respectively (Table 1). There was a positive correlation between sand fly density measured 251 by sticky and light traps in all study sites (rho=0.71, p<0.05).

252 **3.2. Sand fly species frequency and abundance**

The number (relative frequency) of sand flies identified to species level from sticky traps 253 254 was 4,464 (97%), of which 65% were males and 35% were females. The relative frequencies (male/female ratio) of species included Sergentomyia minuta: 60 (46/54)%, P. 255 256 perniciosus: 32 (92/8)%, Phlebotomus papatasi: 5 (90/10)%, Phlebotomus sergenti: 3 (94/6)% and P. ariasi: 0.4 (88/12)% (Table 2). The remarkable difference in the sex ratio 257 between S. minuta and other species was observed in most places (Table 2). The relative 258 259 abundance of species varied between sites; for example, in sites where sand flies were most abundant, S. minuta dominated in sites 2, 22 and 28, whereas P. perniciosus was 260 comparatively more abundant in sites 4 and 17 (Table 2). Similarly, P. papatasi was 261 262 relatively more abundant in sites 1 and 18 and P. sergenti in 28, compared to other sites (Table 2). 263

264 CDC light trap specimens identified at species level included 474 sand flies (99%) with 265 55% males and 45% females. The same five species from sticky traps were detected in the 266 light trap, but their relative frequency and sex ratio was very different to the former. Species percentages (male/female) were, P. perniciosus: 63 (57/43)%, S. minuta: 24 267 (62/38)%, P. papatasi: 8 (23/78)%, P. ariasi: 5 (50/50)% and P. sergenti: 0.4 (50/50)%. 268 The relative abundance of light trap species also differed according to site. In the five sites 269 270 where sand flies were most abundant (sites 4, 5, 18, 20 and 28), species proportions ranged from 46-85% for P. perniciosus, 10-42% for S. minuta, 2-12% for P. papatasi, 0-17% for 271 272 *P. ariasi* and 0-2% for *P. sergenti* (Table 2).

273 **3.3. Sand fly temporal dynamics and relationship with climatic variables**

274 The proportion of positive sticky traps, and overall sand fly and S. minuta density in these traps peaked in September 2013 and in May and July 2014 (Table 3). Notably, 275 sampling was not possible during these three months in sites 1, 6 and 17 which were among 276 those with the highest sand fly density in the study (Table 1). In contrast, P. perniciosus 277 density peaked in September 2013 and 2014. Phlebotomus papatasi and P. sergenti were 278 found in low numbers most months although the majority of *P. sergenti* was captured in 279 280 June 2014. In contrast, P. ariasi was detected only in September and October 2014 and 281 May and July 2015 (Table 3). Sand fly density in CDC traps placed in June and July 2015 282 was also variable in time; total sand fly and *P. perniciosus* density peaked in the third week 283 of July, whilst S. minuta density was highest in the first week of July (data not shown).

The contrasting temporal dynamics between species, years and sites in sticky traps are reflected in the weekly variation of *P. perniciosus* and *S. minuta* densities in sites 2, 3, 4, situated in the same residential estate and sampled in the same weeks. In contrast to the overall pattern, *P. perniciosus* density peaked in the fourth week of October 2014 and May 2015, and was most abundant in site 4 (Fig. 2). *Sergentomyia minuta* predominated in sites 2 and 3 until the fourth week of July although it was similarly abundant in site 4 later, peaking in September 2014.

Regarding climatic variables, the nightly mean (range) RH (%), temperature and wind speed and the maximum (range) wind speed in all sites during the study period were 70 (45-95)%, 20 (14-25)°C, 0.7 (0.2-1.4)m/s and 2.0 (0.9-3.1)m/s, respectively. The overall sand fly density in sticky traps was negatively associated with the mean RH (%) and positively with the mean temperature and mean and maximum wind speed (p<0.05). 296

297

However, differences in the mean and maximum wind speed between high- and lowdensity sites were numerically very small: 0.11 m/s and 0.17 m/s, respectively.

298 **3.4.** Sand fly density in sticky traps and site and trap location environmental features

Ten of the thirteen trap locations with a median sand fly density >10 smd were in nonurbanized sites. They included three underground caves (29-44 smd), the ruins of an old pig farm (4 traps: 11-36 smd), an abandoned small, brick dog house (26 smd), an old pile of firewood (14 smd) and a 30 cm wide rock crevice (11 smd). The pig farm and one of the caves were part of site 1 and were 200-300 m away from two sheep farms and house plot sites 2 and 3. The other two caves and the crevice were 50-100 m away from house plot site 18 and other premises that had backyard chickens and sporting pigeons.

Bivariate analysis indicated that *P. perniciosus*- densities in house plots were significantly associated with larger properties situated in the middle altitude range, with extensive vegetated and non-vegetated earth areas and not permanently inhabited, but used mostly during weekends and summer periods and without a swimming pool (Table 4). Moreover, it was not associated with the permanent presence of dogs or to having a history of CanL. Similarly, *P. perniciosus* density within house plots was greatest in places situated at some distance from people's transit, protected from rains and near walls (Table 4).

In the site-specific multilevel negative binomial model type "a", none of the fixed explanatory variable were significantly associated with *P. perniciosus* density and there was large remaining, unexplained variability between sites (house-plot variance: 46.79) and very little between weeks (0.03) (not tabulated). Instead, trap-level type "b" model indicated that *P. perniciosus* density decreased with trap distance to the wall, and differed between sites, and there was some remaining unaccounted for variation between traps (trap
variance: 1.17) (Table 5).

320

321 **4. Discussion**

The present study assessed for the first time sand fly small-scale distribution and species 322 323 diversity in modern periurban residential estates in L. infantum endemic southern Europe. Sand flies, mostly S. minuta and P. perniciosus, were widespread in the area of study, but 324 325 their density varied greatly between and within sites. Moreover, the temporal dynamics 326 differed between species, years and sites situated close to each other. Also, as previously shown, the estimated species diversity and sex ratios of the five sand fly species here 327 identified strongly depended on the trap type used (Martínez-Ortega 1985a; Martínez-328 329 Ortega et al. 1991; Alexander 2000; Alten et al. 2015; Muñoz et al. 2018).

330 The predominance of S. minuta and P. perniciosus is in agreement with other studies in 331 Spain employing sticky traps (Gálvez et al. 2010; Ballart et al. 2014), and P. ariasi, P. papatasi and P. sergenti are the other most frequently reported in studies in southeast Spain 332 333 (Martínez-Ortega 1985b; Martínez-Ortega et al. 1991; Muñoz et al. 2018, 2019). Other species previously reported in southeast Spain in very small numbers and not detected in 334 335 the present include P. longicuspis, P. chabaudi, P. alexandri and P. langeroni (Martínez-Ortega 1985b; Martínez Ortega et al. 1992; Risueño et al. 2017; Díaz Sáez et al. 2018). 336 Phlebotomus longicuspis is morphologically very similar to P. perniciosus and specimens 337 338 from Spain were proposed to be the same species (Collantes and Martínez Ortega 1997; Martín-Sánchez et al. 2000). Phlebotomus langeroni is typically found associated to rabbit 339 burrows (Martínez Ortega et al. 1992; Díaz Sáez et al. 2018). Phlebotomus chabaudi is 340

mostly found in Northern Africa (Lehrter et al. 2017). In contrast, *P. alexandri* and other
closely related sister species, have a wider distribution ranging from Morocco in the west,
to China in the East and Ethiopia in the south (Depaquit et al. 2000).

Comparatively few premise-level environmental factors were associated with P. 344 345 perniciosus density in the bivariate analysis and none in the multilevel modeling which highlighted large, unexplained variation between sites. This may reflect low statistical 346 power, probably because relatively few sites from a fairly small and environmentally 347 348 similar area were examined for unequal periods of time in some cases. Clearly, the results from the present study indicate that a larger sample size would be required to identify other 349 features of periurban residencies that influence sand fly densities. They also highlight the 350 351 need for a combined effort to survey multiple similar residential areas across Mediterranean countries using standard sampling and reporting methodology and the need for meta-352 analysis. 353

354 The precise locations where sand flies breed have not been fully characterized, and eggs, 355 larvae and pupae are very difficult to find in soil samples. As a result, most entomological 356 surveys focus only on adult stages. In the natural environment they are typically found resting in large numbers in places protected from desiccation such as caves, uninhabited 357 buildings, rock crevices and undisturbed rock and log piles close to groups of domestic 358 359 animals (Alexander 2000). These were the precise habitats in the non-urbanized sites in the 360 present study where sand flies were most abundant. Sand fly density in non-urbanized sites 361 was positively correlated to that in close-by residencies, suggesting that the former could be a source of insects for the latter. This phenomenon is described in Israel, where residential 362 363 areas are continuously exposed to P. papatasi from surrounding agricultural land (Orshan et al. 2016). Alcover et al. (2014) in Majorca similarly found greater *P. perniciosus* density
at the edge compared to within human settlements. Further studies including a larger
number of sites are necessary for a better understanding on how non-urbanized areas
contribute to the vector population in nearby residential estates in Mediterranean countries.

368 Most likely, periurban residential estates also provided suitable breeding habitats for sand flies, and density was highly variable between traps in the same site. Sand fly 369 populations are typically spatially over-dispersed on a large and small geographical scale 370 371 (Rioux et al. 2013; Alten et al. 2016; Muñoz et al. 2019). Like in previous studies, the density of *P. perniciosus* was negatively associated to trap distance to a wall (Risueño et al. 372 373 2017; Muñoz et al. 2018). Walls have several advantages for sand flies (Alexander 2000). 374 They protect them from strong wind and they often have vegetation growing at the base. Their surfaces allow them to rest and move vertically in typical short hopping steps, and 375 376 holes and cracks provide suitable breeding places. However, other traps in the present study 377 that were not situated close to walls, also had high sand fly counts, but the multivariable 378 analysis revealed no association between density and variables potentially affecting the insect's life cycle such as being under cover or close to vegetation, surface water or the 379 dog's sleeping place. Sticky traps are interception traps collecting a random and 380 381 comprehensive selection of sand flies in the immediacy of the trap, and are ideal for 382 ecological studies investigating species diversity (Alexander 2000; Alten et al. 2015). Light trap captures are biased towards host-seeking phototrophic species present within a few 383 meters (<10 m) from the trap, including P. perniciosus and P. ariasi L. infantum vectors 384 385 (Alexander 2000). However, neither sticky nor light traps inform on whether the site is

suitable for breeding or not, and this constitutes an important limitation in sand fly andleishmaniasis control (Alexander 2000; Alten et al. 2015).

388 Climate determines the annual activity of sand flies, influencing the length of diapause during cold months, the number of life cycles and the resulting adult density peaks between 389 390 spring and autumn (Alten et al. 2016). Phlebotomus perniciosus seasonality in Murcia was found to be bimodal with maximum densities in July and September when using sticky 391 392 traps (Martínez-Ortega 1986; Muñoz et al. 2018), and unimodal with a single July peak 393 when sampling with CDC light traps (Muñoz et al. 2018). Here, the overall sand fly density differed between years and three peaks were detected, one in September 2013, one in May 394 2014 and one in July 2014, and there were substantial differences between sites and 395 396 species. All this reflects the complexity of the system regulating sand fly demographics at a small geographical scale, and that accurate estimation of species seasonality in a particular 397 area requires continuous longitudinal sampling of a large number of sites over several years 398 399 (Alten et al. 2016). In ideal laboratory conditions at 25-26 °C, P. perniciosus specimens 400 from Murcia may take 41 to 47 days to complete a life cycle (Volf and Volfova 2011). Feeding preferences and attraction to light vary between species and sexes (Alexander 401 2000; Alten et al. 2015); while female P. perniciosus are highly phototropic and tend to 402 403 concentrate closer to their blood source than males (Muñoz et al. 2018), S. minuta is less 404 attracted to light and in the rural environment both male and females may be similarly abundant relatively far away from groups of hot-blooded animal groups, with females 405 probably feeding on lizards (Muñoz et al. 2018). Such inherent biological diversity would 406 407 also explain the remarkable sex and trap-specific spatial and temporal distributional differences observed here and elsewhere (Martínez-Ortega 1985a; Martínez-Ortega et al. 408

409 1991; Muñoz et al. 2018). We can further conclude that a very large number of sticky traps
410 are required to attain a representative picture of sand fly distribution in a particular site and
411 gain statistical power to detect associations with environmental variables.

The small number of premises precluded a robust investigation of the relationship 412 413 between leishmaniasis incidence and sand fly density, and it was not an objective of the present study. This issue is a matter of debate. Outbreaks are typically associated with large 414 415 densities of infected vectors (Arce et al. 2013; Jiménez et al. 2013), but vectors may also be 416 very abundant and infection prevalence low, in areas with a high density of non-Leishmania competent hosts (Muñoz et al. 2019). The need for a more in depth understanding of this 417 418 highly relevant question is the of the VectorNet initiative at core 419 (https://www.ecdc.europa.eu/en/about-us/partnerships-and-networks/disease-and-

laboratory-networks/vector-net). The aim of this network of entomologists is to gather data
on vectors related to both animal and human health, to generate maps and investigate
environmental determinants of vector distributions
(https://www.ecdc.europa.eu/sites/default/files/documents/vector-abundance-and-

seasonality.pdf). Ideally, reports should convey quantitative sand fly density information at 424 the insect species and sex level for each of the places sampled. Essential trap-related 425 426 information includes: the type, number, dimensions (for sticky traps), operational time and 427 precise geographical location. The number of consecutive days that the same traps are in operation is also an important parameter to consider (Gálvez et al. 2010). Sand fly 428 population depletion and loss of viscosity in sticky traps over time may lead to an 429 430 underestimation of sand fly density. In this study relatively few traps per site were used and were placed in wide, open spaces so it is very unlikely that sand flies were depleted from 431

sites. Loss of trap adherence is particularly important in high humidity places (Alexander 432 433 2000), which is not the case of Murcia, and care was taken in the present study to impregnate the sheets thoroughly before using them. These issues require further 434 investigation. Other useful data to report is on variables associated to the trap 435 microenvironment including if the trap is protected from rain and wind and trap distance to 436 437 the ground, walls and to resident animal groups (farms, kennels and catteries). These data should be incorporated into multivariable models to adjust sand fly species density 438 estimations in a particular ecotope. In a recent review of published scientific studies 439 reporting sand fly distribution data in Europe and neighboring countries, less than half of 440 441 the articles provided the data needed to calculate the sampling effort and sand fly density (as here proposed), and this was particularly a problem when sticky traps were used 442 443 (Muñoz C. and Berriatua E., personal communication). Other limitations with those studies included not providing precise geographical locations and scarce details on trap position 444 relative to potential micro-environmental risk factors. We have been careful to provide all 445 the information needed to ensure these data can be used in continental scale analyses. It is 446 hoped that this paper will encourage other authors to provide such details when reporting 447 the results of entomological surveys. 448

In summary, here we show that periurban residential estates provide the right conditions for sand fly vectors to thrive. We also proved that sand fly distribution is highly spatially temporarily heterogeneous at a very small geographical scale. Detailed understanding of factors governing sand fly density requires further studies with a similar reporting approach, which will enable a meta-analytic methodology to be implemented.

455 Supporting Information

456 Additional supporting information may be found online in the Supporting Information457 section at the end of the article.

458 Photos 01–18: Photographs of places where sticky and light traps were placed. A study of
459 sand fly abundance in 29 periurban sites in Murcia City in southeast Spain.

460

461 **Declarations**

462 Funding

The study was funded by the Spanish Ministry of Science and Innovation (Grant number: 463 AGL2013-46981-R) and received support from VectorNet, a European network for sharing 464 data on the geographic distribution of arthropod vectors, transmitting human and animal 465 466 disease agents funded by the European Food Safety Authority (EFSA) and the European Centre for Disease prevention and Control (ECDC). CM held a PhD grant from University 467 of Murcia (Contrato predoctoral FPU). TS and SE were beneficiaries of a COST TD1303 468 short scientific mission grant. PDLR receives support from the Fundación Séneca (Grant 469 470 number: 19908/GERM/15).

471 **Conflicts of interest**

There are no potential sources of conflict of interest in this work and no disputes over the ownership of the data presented in the paper, and all contributions have been attributed appropriately.

475 Availability of data

476 The data that support the findings of this study are available from the corresponding author477 upon reasonable request.

478 Authors' contribution

Every author of this article participated in the trapping and collection of sand flies, procured environmental data of the trapping sites and contributed to the final written version of the manuscript. CM, JR and TS were responsible for identifying individual sand fly specimens. CM, JR, PPC and EB analyzed the data, prepared tables and figures, and the first written version of the manuscript.

484

485 **References**

Akhoundi M, Kuhls K, Cannet A, Votýpka J, Marty P, Delaunay P, Sereno D (2016) A
historical overview of the classification, evolution, and dispersion of *Leishmania*parasites and sandflies. PLoS Negl Trop Dis 10:e0004349.
https://doi.org/10.1371/journal.pntd.0004349

Alcover MM, Ballart C, Martín-Sánchez J, Serra T, Castillejo S, Portús M, Gállego M
(2014) Factors influencing the presence of sand flies in Majorca (Balearic Islands,
Spain) with special reference to *Phlebotomus pernicious*, vector of *Leishmania infantum*. Parasit Vectors 7:421. https://doi.org/10.1186/1756-3305-7-421

- Alexander B (2000) Sampling methods for phlebotomine sandflies. Med Vet Entomol
 14:109-122. https://doi.org/10.1046/j.1365-2915.2000.00237.x
- 496 Alten B, Maia C, Afonso MO, Campino L, Jiménez M, González E, Molina R, Bañuls AL,
- 497 Prudhomme J, Vergnes B, Toty C, Cassan C, Rahola N, Thierry M, Sereno D,

498	Bongiorno G, Bianchi R, Khoury C, Tsirigotakis N, Dokianakis E, Antoniou M,
499	Christodoulou V, Mazeris A, Karakus M, Ozbel Y, Arserim SK, Erisoz Kasap O,
500	Gunay F, Oguz G, Kaynas S, Tsertsvadze N, Tskhvaradze L, Giorgobiani E, Gramiccia
501	M, Volf P, Gradoni L (2016) Seasonal dynamics of phlebotomine sand fly species
502	proven vectors of Mediterranean leishmaniasis caused by Leishmania infantum. PLoS
503	Negl Trop Dis 10:e0004458. https://doi.org/10.1371/journal.pntd.0004458
504	Alten B, Ozbel Y, Ergunay K, Kasap OE, Cull B, Antoniou M, Velo E, Prudhomme J,
505	Molina R, Bañuls A-L, Schaffner F, Hendrickx G, Van Bortel W, Medlock JM (2015)

506 Sampling strategies for phlebotomine sand flies (Diptera: Psychodidae) in Europe. Bull

507 Entomol Res 105:664-678. https://doi.org/10.1017/S0007485315000127

Arce A, Estirado A, Ordobas M, Sevilla S, García N, Moratilla L, de la Fuente S, Martínez
AM, Pérez AM, Aránguez E, Iriso A, Sevillano O, Bernal J, Vilas F (2013) Re-

510 emergence of leishmaniasis in Spain: community outbreak in Madrid, Spain, 2009 to

- 511 2012. Euro Surveill 18:20546. https://doi.org/10.2807/1560-7917.es2013.18.30.20546
- Ayhan N, Charrel RN (2017) Of phlebotomines (sandflies) and viruses: a comprehensive
 perspective on a complex situation. Curr Opin Insect Sci 22:117-124.
 https://doi.org/10.1016/j.cois.2017.05.019
- Ballart C, Guerrero I, Castells X, Barón S, Castillejo S, Alcover MM, Portús M, Gállego M
 (2014) Importance of individual analysis of environmental and climatic factors
 affecting the density of *Leishmania* vectors living in the same geographical area: the
 example of *Phlebotomus ariasi* and *P. perniciosus* in northeast Spain. Geospat Health
 8:389-403. https://doi.org/10.4081/gh.2014.28

520	Bates D, Mächler M, Bolker B, Walker S (2015) Fitting linear mixed-effects models using
521	lme4. J Stat Softw 67:1-48. https://doi.org/10.18637/jss.v067.i01
522	Collantes F, Martínez Ortega E (1997) Sobre la validez taxonómica de Phlebotomus

- *longicuspis* (Nitzulescu, 1931) (Diptera: Psychodidae). Bol Asoc Esp Entomol 21:141146.
- Depaquit J, Ferté H, Léger N, Killick-Kendrick R, Rioux JA, Killick-Kendrick M, Hanafi
 HA, Gobert S (2000) Molecular systematics of the phlebotomine sandflies of the
 subgenus *Paraphlebotomus* (Diptera, Psychodidae, *Phlebotomus*) based on ITS2
 rDNA sequences. Hypotheses of dispersion and speciation. Insect Mol Biol 9:293-300.
 https://doi.org/10.1046/j.1365-2583.2000.00179.x
- Díaz Sáez V, Morillas-Márquez F, Merino-Espinosa G, Corpas-López V, Morales-Yuste
 M, Pesson B, Barón-López S, Lucientes-Curdi J, Martín-Sánchez J (2018) *Phlebotomus langeroni* Nitzulescu (Diptera, Psychodidae) a new vector for *Leishmania infantum* in Europe. Parasitol Res 117:1105-1113. https://doi.org/10.1007/s00436-0185788-8
- Gállego-Berenguer J. Botet-Fregola J, Gállego-Culleré M, Portús-Vinyeta M (1992) Los
 flebotomos de la España peninsular e Islas Baleares: identificación y corología:
 comentarios sobre los métodos de captura. In: Hernández S (ed) Memoriam al Profesor
 Dr. DF de P Martínez Gómez. Publicaciones de la Universidad de Córdoba, Córdoba,
 pp 581-600.
- Gálvez R, Descalzo MA, Miró G, Jiménez MI, Martín O, Dos Santos-Brandao F, Guerrero
 I, Cubero E, Molina R (2010) Seasonal trends and spatial relations between

- 542 environmental/meteorological factors and leishmaniosis sand fly vector abundances in
- 543 Central Spain. Acta Trop 115:95-102. https://doi.org/10.1016/j.actatropica.2010.02.009
- 544 Gálvez R, Montoya A, Cruz I, Fernández C, Martín O, Checa R, Chicharro C, Migueláñez
- 545 S, Marino V, Miró G (2020) Latest trends in Leishmania infantum infection in dogs in
- 546 Spain, Part I: Mapped seroprevalence and sand fly distributions. Parasit Vectors
- 547 13:204. https://doi.org/10.1186/s13071-020-04081-7
- 548 García San Miguel L, Sierra MJ, Vazquez A, Fernandez-Martínez B, Molina R, Sanchez-
- 549 Seco MP, Lucientes J, Figuerola J, de Ory F, Monge S, Suarez B, Simón F (2020)
- 550 Phlebovirus-associated diseases transmitted by phlebotominae in Spain: Are we at
- risk? Enferm Infecc Microbiol Clin. https://doi.org/10.1016/j.eimc.2020.02.026
- Gil Collado J, Morillas Márquez F, Sanchís Marín MC (1989) Los flebotomos en España.
 Rev San Hig Púb 63:15-34.
- 554 Goyena E, Pérez-Cutillas P, Chitimia L, Risueño J, García-Martínez JD, Bernal LJ,
- 555 Berriatua E (2016) A cross-sectional study of the impact of regular use of insecticides
- in dogs on Canine Leishmaniosis seroprevalence in southeast Spain. Prev Vet Med
- 557 124:78-84. https://doi.org/10.1016/j.prevetmed.2015.12.009
- 558 Herrador Z, Gherasim A, Jimenez BC, Granados M, San Martín JV, Aparicio P (2015)
- 559 Epidemiological changes in leishmaniasis in Spain according to hospitalization-based
- records, 1997–2011: raising awareness towards leishmaniasis in non-HIV patients.
- 561 PLoS Negl Trop Dis 9:e0003594. https://doi.org/10.1371/journal.pntd.0003594
- Jiménez M, González E, Iriso A, Marco E, Alegret A, Fúster F, Molina R (2013) Detection
- 563 of *Leishmania infantum* and identification of blood meals in *Phlebotomus perniciosus*

- from a focus of human leishmaniasis in Madrid, Spain. Parasitol Res 112:2453-2459.
 https://doi.org/10.1007/s00436-013-3406-3
- 566 Keskin M, Dogru AO, Balcik FB, Goksel C, Ulugtekin N, Sozen S (2015) Comparing
- 567 spatial interpolation methods for mapping meteorological data in Turkey. In: Bilge A,
- Toy A, Günay M (eds) Energy Systems and Management. Springer International
 Publishing, Cham, pp 33-42.
- Kleinbaum DG, Kupper LL, Muller KE, Nizam A (1998) Applied regression analysis and
 other multivariable methods. Duxbury Press, Pacific Grove, CA.
- 572 Lehrter V, Bañuls A-L, Léger N, Rioux J-A, Depaquit J (2017) *Phlebotomus*573 (*Paraphlebotomus*) *chabaudi* and *Phlebotomus riouxi*: closely related species or
 574 synonyms? Parasite 24:47. https://doi.org/10.1051/parasite/2017050
- 575 Martínez Ortega E, Conesa Gállego E, Goyena Salgado M, Romera Lozano E (1992)
 576 Presencia de *Phlebotomus (Larroussius) langeroni* Nitzulescu, 1930 (Diptera:
 577 Psychodidae) en la Península Ibérica. Bolm Soc Port Ent 139:196.
- 578 Martínez-García FA, Moreno-Docón A, López-López M, Albert-Lacal L, Martínez-Toldos
- 579 MC, Segovia-Hernández M, Fernández-Barreiro A (2007) [A case of meningitis due to
 580 Toscana virus in Murcia]. Rev Neurol 45:317-318.
- 581 Martínez-Ortega E (1985a) Los flebotomos ibéricos (Diptera: Psychodidae). II. El sureste.
 582 An Biol 3:113-119.
- 583 Martínez-Ortega E (1985b) Los flebotomos Ibéricos (Diptera: Psychodidae). I. Almería. An
 584 Biol 3:107-111.

- 585 Martínez-Ortega E (1986) Biología de los flebotomos ibéricos (Diptera: Psychodidae) en
 586 condiciones naturales. Ann Ist Super Sanita 22:73-78.
- 587 Martínez-Ortega E, Conesa-Gallego E (1987) Caracteres morfológicos de interés
 588 taxonómico de los flebotomos (Diptera, Psychodidae) de la Península Ibérica. An Biol
 589 11:43-53.
- 590 Martínez-Ortega E, Romera E, Conesa-Gallego E, Goyena M (1991) Estudio comparado de
 591 la antropofilia y el fototropismo de los flebotomos en un foco de leishmaniasis del
 592 sureste de la Península Ibérica. Parassitologia 33:413-419.
- Martín-Sánchez J, Gramiccia M, Pesson B, Morillas-Marquez F (2000) Genetic
 polymorphism in sympatric species of the genus *Phlebotomus*, with special reference
 to *Phlebotomus perniciosus* and *Phlebotomus longicuspis* (Diptera, Phlebotomidae).
 Parasite 7:247-254. https://doi.org/10.1051/parasite/2000074247
- Molina R, Jiménez MI, Cruz I, Iriso A, Martín-Martín I, Sevillano O, Melero S, Bernal J
 (2012) The hare (*Lepus granatensis*) as potential sylvatic reservoir of *Leishmania infantum* in Spain. Vet Parasitol 190:268-271.
 https://doi.org/10.1016/j.vetpar.2012.05.006
- 601 Muñoz C, Martínez-de la Puente J, Figuerola J, Pérez-Cutillas P, Navarro R, Ortuño M,
- 602 Bernal LJ, Ortiz J, Soriguer R, Berriatua E (2019) Molecular xenomonitoring and host
- 603 identification of *Leishmania* sand fly vectors in a Mediterranean periurban wildlife
- 604 park. Transbound Emerg Dis 66:2546-2561. https://doi.org/10.1111/tbed.13319
- 605 Muñoz C, Risueño J, Yilmaz A, Pérez-Cutillas P, Goyena E, Ortuño M, Bernal LJ, Ortiz J,
- Alten B, Berriatua E (2018) Investigations of *Phlebotomus perniciosus* sand flies in

rural Spain reveal strongly aggregated and gender-specific spatial distributions and
advocate use of light-attraction traps. Med Vet Entomol 32:186-196.
https://doi.org/10.1111/mve.12275

610 Orshan L, Elbaz S, Ben-Ari Y, Akad F, Afik O, Ben-Avi I, Dias D, Ish-Shalom D,

611 Studentsky L, Zonstein I (2016) Distribution and dispersal of *Phlebotomus papatasi*

(Diptera: Psychodidae) in a zoonotic cutaneous leishmaniasis focus, the Northern
Negev, Israel. PLoS Negl Trop Dis 10:e0004819.
https://doi.org/10.1371/journal.pntd.0004819

Pérez-Cutillas P, Goyena E, Chitimia L, De la Rúa P, Bernal LJ, Fisa R, Riera C, Iborra A, 615 Murcia L, Segovia M, Berriatua E (2015) Spatial distribution of human asymptomatic 616 Leishmania infantum infection in southeast Spain: a study of environmental, 617 618 demographic and social risk factors. Acta Trop 146 :127-134. https://doi.org/10.1016/j.actatropica.2015.03.017 619

620 Remoli ME, Jiménez M, Fortuna C, Benedetti E, Marchi A, Genovese D, Gramiccia M,

621 Molina R, Ciufolini MG (2016) Phleboviruses detection in *Phlebotomus perniciosus*

from a human leishmaniasis focus in South-West Madrid region, Spain. Parasit Vectors
9:205. https://doi.org/10.1186/s13071-016-1488-3

624 Rioux J-A, Carron S, Dereure J, Périères J, Zeraia L, Franquet E, Babinot M, Gállego M,

625 Prudhomme J (2013) Ecology of leishmaniasis in the South of France. 22. Reliability

and representativeness of 12 Phlebotomus ariasi, P. perniciosus and Sergentomyia

627 *minuta* (Diptera: Psychodidae) sampling stations in Vallespir (eastern French Pyrenees

628 region). Parasite 20:34. https://doi.org/10.1051/parasite/2013035

629	Risueño J, Muñoz C, Pérez-Cutillas P, Goyena E, Gonzálvez M, Ortuño M, Bernal LJ,
630	Ortiz J, Alten B, Berriatua E (2017) Understanding Phlebotomus perniciosus
631	abundance in south-east Spain: assessing the role of environmental and anthropic
632	factors. Parasit Vectors 10:189. https://doi.org/10.1186/s13071-017-2135-3
633	Snijders TAB, Bosker RJ (1999) Multilevel analysis: an introduction to basic and advanced

- 634 multilevel modeling. SAGE Publications, London.
- 635 Volf P, Volfova V (2011) Establishment and maintenance of sand fly colonies. J Vector

636 Ecol 36:S1-S9. https://doi.org/10.1111/j.1948-7134.2011.00106.x

- 637 WHO (2010) Control of the leishmaniases: report of a meeting of the WHO Expert
- 638 Committee on the Control of Leishmaniases. World Health Organization.

Estate	Environment	Latitude/	Sticky traps								CDC light traps			
/Site		Longitude (zone 30S)	No. weeks	No. traps	% positive traps	No. sand flies	Trap area (m²)	Sampling effort ^a	Sand fly density ^b	No. traps ^c	% positive traps	No. sand flies	Sand fly density ²	
1/1	non-urban. ^d	653667.7/4208231.8	12	74	93	751	4.6	32.3	23.2	-	-	-	-	
1/2	house plot	653914.4/4208225.3	20	285	40	335	17.8	124.5	2.7	4	100	25	6.3	
1/3	house plot	653967.9/4208363.3	20	245	42	307	15.3	104.0	3.0	4	100	15	3.8	
1/4	house plot	655073.0/4208773.0	20	239	26	208	14.9	103.8	2.0	4	100	49	12.3	
1/5	house plot	654948.2/4208851.9	14	149	19	49	9.3	66.3	0.7	3	100	47	15.7	
2/6	non-urban.	659529.9/4219233.3	12	142	42	483	8.9	67.3	7.2	4	100	12	3.0	
3/7	house plot	655131.2/4214164.0	12	104	32	56	6.6	50.2	1.1	4	100	15	3.8	
4/8	non-urban.	661587.4/4216176.4	15	162	7	15	10.1	69.1	0.2	-	-	-	-	
4/9	house plot	661533.9/4216140.4	16	108	18	26	6.7	46.1	0.6	5	100	13	2.6	
4/10	house plot	661581.3/4215885.7	12	84	12	10	5.2	35.9	0.3	4	100	14	3.5	
4/11	non-urban.	661917.0/4217693.0	11	72	7	7	4.5	31.4	0.2	-	-	-	-	
4/12	house plot	660487.9/4218302.0	11	58	21	21	3.6	25.4	0.8	3	100	11	3.7	
5/13	house plot	660762.8/4206747.6	4	32	28	64	2.0	14.0	4.6	-	-	-	-	
5/14	non-urban.	658596.3/4207038.2	11	96	38	78	6.0	41.1	1.9	-	-	-	-	
5/15	house plot	659167.9/4206919.6	19	140	3	4	8.7	59.5	0.1	4	25	1	0.3	
5/16	house plot	658946.7/4206592.0	8	53	19	21	3.3	22.0	1.0	-	-	-	-	
6/17	non-urban.	667080.8/4213124.3	5	48	44	297	3.2	22.7	13.1	-	-	-	-	
6/18	house plot	666660.6/4213356.0	5	50	42	115	3.7	26.2	4.4	2	100	59	29.5	
9/19	house plot	666589.4/4212170.4	10	110	12	18	6.9	48.0	0.4	3	67	4	1.3	
7/20	house plot	670594.1/4214073.1	12	155	43	146	9.7	68.3	2.1	4	100	115	28.8	

Table 1. Percentage of sticky and light traps with sand flies and sand fly density in 29 periurban sites in Murcia City in 2013-15.

8/21	non-urban.	664857.6/4209411.4	10	86	17	28	5.4	42.1	0.7	-	-	-	-
8/22	house plot	664752.1/4209476.1	17	217	62	835	13.5	101.0	8.3	4	100	16	4.0
10/23	house plot	663540.0/4211065.0	3	15	7	1	0.9	6.6	0.2	-	-	-	-
11/24	non-urban.	661913.0/4201210.0	12	75	20	22	4.7	32.8	0.7	-	-	-	-
11/25	house plot	661809.0/4201088.0	12	78	4	4	4.9	34.1	0.1	4	0	0	0.0
11/26	house plot	661777.0/4201090.0	12	74	0	0	4.6	32.3	0.0	4	50	2	0.5
12/27	non-urban.	671824.9/4200912.1	12	68	15	20	4.2	29.8	0.7	-	-	-	-
12/28	house plot	668160.0/4203927.0	18	234	58	656	14.6	100.9	6.5	4	100	75	18.8
13/29	house plot	687342.0/4218749.0	12	75	9	9	4.7	33.5	0.3	3	67	7	2.3
All				3328	31	4586	208.7	1471	3.1	67	84	480	7.2

 a Sampling effort: number of trapping days multiplied by the trap area (m²) for sticky traps and by the number of traps in the case of light traps.

642 ^b Sand fly density: the number of sand flies collected divided by the sampling effort.

643 ^c In this case the number of traps equals the sampling effort.

^d Non urbanized areas situated in the perimeter of the residential estates where house-plots were located.

645

Site	No. sand flies		P. ariasi		S. minuta	S. minuta		P. papatasi		P. perniciosus		P. sergenti		
5100	Sticky	Light	Sticky	Light	Sticky	Light	Sticky	Light	Sticky	Light	Sticky	Light		
1	745	0	0.8 (100/0)	-	51 (44/56)	-	14 (95/5)	-	32 (96/4)	-	2 (89/11)	-		
2	323	24	0	0	78 (41/59)	8 (50/50)	3 (89/11)	25 (0/100)	19 (93/7)	67 (12/88)	0	0		
3	297	14	0	0	56 (44/56)	43 (50/50)	3 (80/20)	14 (50/50)	41 (91/9)	36 (40/60)	0	7 (0/100)		
4	205	48	0.5 (100/0)	2 (0/100)	34 (64/36)	21 (60/40)	0.5 (100/0)	2 (0/100)	65 (99/1)	75 (58/42)	0.5 (100/0)	0		
5	43	46	0	0	26 (36/64)	11 (100/0)	0	4 (0/100)	67 (93/7)	85 (62/38)	7 (100/0)	0		
6	480	12	0	0	58 (51/49)	58 (71/29)	1 (100/0)	0	41 (87/13)	42 (20/80)	0	0		
7	55	15	0	0	69 (58/42)	20 (100/0)	13 (57/43)	27 (0/100)	18 (90/10)	53 (25/75)	0	0		
8	14	0	0	-	86 (33/67)	-	0	-	14 (100/0)	-	0	-		
9	25	13	0	0	72 (83/17)	15 (50/50)	0	8 (0/100)	28 (86/14)	77 (70/30)	0	0		
10	10	13	0	0	80 (38/63)	8 (100/0)	0	0	20 (100/0)	92 (42/58)	0	0		
11	7	0	0	-	57 (50/50)	-	0	-	43 (100/0)	-	0	-		
12	20	11	0	0	75 (40/60)	9 (100/0)	0	0	25 (60/40)	91 (30/70)	0	0		
13	61	0	0	-	80 (47/53)	-	0	-	20 (92/8)	-	0	-		
14	78	0	0	-	81 (56/44)	-	4 (100/0)	-	14 (82/18)	-	1 (0/100)	-		
15	4	1	0	0	75 (67/33)	100 (100/0)	0	0	25 (100/0)	0	0	0		
16	19	0	0	-	11 (50/50)	-	0	-	84 (100/0)	-	5 (100/0)	-		
17	296	0	0.7 (100/0)	-	24 (26/74)	-	0.4 (100/0)	-	75 (95/5)	-	0	-		
18	115	59	2 (50/50)	5 (33/66)	0	10 (50/50)	24 (86/14)	12 (29/71)	70 (90/10)	71 (67/33)	3 (100/0)	2 (100/0)		
19	18	4	0	0	67 (42/58)	25 (0/100)	6 (100/0)	0	28 (80/20)	75 (33/67)	0	0		
20	145	115	4 (83/17)	17 (55/45)	39 (54/46)	25 (62/38)	10 (73/27)	6 (57/43)	45 (75/25)	51 (81/19)	2 (100/0)	0		

Table 2. Absolute number of sand flies identified and relative frequency (male/female ratio) of species in sticky and CDC light traps in

study sites in 29 periurban sites in Murcia's metropolitan area in 2013-15.

21	28	0	0	-	89 (76/24)	-	4 (0/100)	-	7 (50/50)	-	0	-
22	794	16	0	0	88 (35/65)	38 (50/50)	1 (80/20)	6 (0/100)	11 (93/7)	56 (67/33)	0.3 (50/50)	0
23	1	0	0	-	100 (0/100)	-	0	-	0	-	0	-
24	22	0	0	-	68 (87/13)	-	9 (100/0)	-	23 (100/0)	-	0	-
25	4	0	0	0	100 (100/0)	0	0	0	0	0	0	0
26	0	2	0	0	0 (0/9)	0	0	0	0	100 (50/50)	0	0
27	20	0	0	-	45 (100/0)	-	0	-	55 (100/0)	-	0	-
28	628	74	0	0	67 (56/44)	42 (58/42)	2 (100/0)	12 (22/78)	17 (94/6)	46 (53/47)	14 (97/3)	0
29	7	7	0	0	43 (33/67)	0	29 (50/50)	0	29 (50/50)	100 (14/86)	0	0
All	4464	474	0.4 (88/12)	5 (50/50)	60 (46/54)	24 (62/38)	5 (90/10)	8 (23/78)	32 (92/8)	63 (57/43)	3 (94/6)	0.4(50/50)

Table 3. Percentage of sticky traps with sand flies (positive traps) and sand fly density (No. sand flies/m²/day) according to month and

653	year in 29	periurban	sites	in Murc	ia City.
-----	------------	-----------	-------	---------	----------

Year-	No.	% positive	Sampling	No. sand flies	Sand fly density ^b							
month	traps	traps	effort ^a		All	S. minuta	P. perniciosus	P. papatasi	P. sergenti	P. ariasi		
2013												
September	186	42	81	386	4.8	3.2	1.5	0.09	0.01	0.00		
October	125	26	52	79	1.5	1.0	0.4	0.06	0.02	0.00		
2014												
May	94	47	37	180	4.8	3.6	1.1	0.05	0.05	0.00		
June	255	34	111	441	4.0	2.9	0.6	0.04	0.48	0.00		
July	110	36	46	227	4.9	4.2	0.4	0.02	0.28	0.00		
September	654	27	290	952	3.3	1.4	1.6	0.18	0.03	0.02		
October	432	25	190	432	2.3	0.9	1.2	0.14	0.03	0.04		
2015												
May	364	31	166	400	2.4	1.3	0.8	0.17	0.04	0.01		
June	741	32	331	956	2.9	1.9	0.7	0.18	0.08	0.00		
July	367	32	165	411	2.5	1.8	0.5	0.15	0.02	0.01		

^a Sampling effort: number of trapping days multiplied by the trap area (m²).

^b Sand fly density: the number of sand flies collected divided by the sampling effort.

Table 4. Percentage (95% CI) of sticky traps with *P. perniciosus* (positive traps) and
median (range) density (No. specimens/m²/day) in positive traps according to house plot

and trap location variables. A study of sand fly abundance in 29 periurban sites in Murcia

659 City in southeast Spain.

	No	Phlebotomus perniciosus				
Variable	traps	% positive traps (95% CI)	Median (range) sand fly density ^a			
(A) House plot						
Altitude						
23-90	1934	14 (12-16)	2 (1-71)			
112-180	968	23 (21-26)*	2 (1-96)			
248-287	426	3 (1-4)	2 (2-5)			
Vegetated/soil area (m ²)						
40-100	780	8 (6-10)	2(2-16)			
140-320	713	14 (11-17)	2 (1-50)			
450-999	519	20 (16-23)*	2 (1-23)			
1824-9096	493	19 (16-23)	2 (2-23)			
Permanently lived						
No	710	20 (17-23)*	2 (1-32)			
Yes	1795	12 (10-13)	2 (1-50)			
Swimming pool						
No	775	18 (16-21)*	2 (1-32)			
Yes	1730	12 (11-14)	2 (2-50)			
(B) Trap location						
Transit area						
No	1132	16 (14-18)*	2 (1-50)			
Yes	1192	12 (10-20)	2 (1-32)			
Wind exposure ^b						
No	545	16 (13-19)	2 (1-16)			
Low	1100	15 (13-17)	2 (1-50)			
Moderate-strong	834	13 (10-15)	2 (1-23)			
Undercover						
No	1724	12 (11-14)	2 (1-50)			
Yes	755	20 (17-23)*	2 (1-16)			
Distance to wall (m)						

0.0-0.1	833	22 (19-25)*	2 (1-50)*					
0.2-0.5	659	12 (9-14)	2 (2-18)					
0.8-2.0	499	9 (7-12)	2 (1-14)					
2.5-10	432	11 (8-14)	2 (2-23)					
>10	56	4 (0-8)	2 (2-2)					
Distance to soil/plants (m)								
0-0.5	1244	15 (13-17)	2 (1-50)*					
0.75-2	517	13 (10-16)	2 (2-16)					
3-10	527	17 (14-21)	2 (1-23)					
15-40	191	10 (6-15)	2 (2-7)					
Distance to dog sleeping area (m)								
0-5	346	19 (15-23)	2 (2-21)					
6-20	607	17 (14-20)	3 (2-50)*					
>25	540	14 (11-17)	2 (2-23)					

660 * p<0.05. Asterisk placed in the highest median or maximum.

^a Sand fly density: the number of sand flies collected divided by the sampling effort, which is No. of trapping
 days multiplied by the trap area (m²).

663 ^b Wind exposure as presumed by owners.

- **Table 5.** Estimates from multilevel negative binomial regression models investigating trap-
- level factors associated with *P. perniciosus* density (log₁₀-transformed) in sticky traps. Five

Variable	Levels	Estimate	Se ^a	P value
Fixed effects				
Intercept		-1.19	0.38	0.0017
Distance to a wall (m)	0	0.00		
	0.1-0.3	-0.34	0.49	0.4936
	0.5-1	-1.06	0.49	0.0289
	1.5-5	-1.30	0.52	0.0125
	6-20	-2.44	0.77	0.0015
Site	2	0.00		
	3	0.91	0.49	0.0618
	4	0.92	0.61	0.1342
	22	1.78	0.55	0.0012
	28	0.93	0.54	0.0826
Random effects variance				
Trap		1.17		
Week		0.001		

sites sampled for 21 weeks in 2013-15.

^a Standard error.

669

671 **Figure captions**

Fig. 1 Location of Phlebotomine sand fly sampling sites in periurban areas of Murcia City,

and meteorological stations from which climatic data was obtained

- **Fig. 2** Temporal dynamics in *P. perniciosus* density (sand flies/m²/day) in sticky traps from
- week 3 in September 2013 (3S.3) to week 2 in July 2015 (5J.3) in the plot of three detached
- 676 homes (sites 2, 3 and 4) in residential estate number 1, in the outskirts of Murcia City
- 677 (southeast Spain). ND1-4 denote periods between two sampling weeks when no data was
- 678 collected