

## Review

# Potential therapeutic effect of SO<sub>2</sub> on fibrosis

Xin-Bao Wang<sup>1</sup>, Hong Cui<sup>1</sup> and Jun-Bao Du<sup>2</sup>

<sup>1</sup>Department of Pediatrics, Beijing Friendship Hospital, Capital Medical University  
and <sup>2</sup>Department of Pediatrics, Peking University First Hospital, Beijing, PR China

**Summary.** Fibrosis is a pathological feature of most chronic diseases and leads to the dysfunction of various organs. However, there is currently no effective method for treating fibrosis. In recent years, a small gas, sulfur dioxide (SO<sub>2</sub>), which can be generated endogenously in mammals, has been found to have vasorelaxation activity, improve cardiac function and decrease myocardial injury. Endogenous SO<sub>2</sub> also mediates the process of fibrosis. Inhibition of endogenous SO<sub>2</sub> can aggravate small pulmonary artery remodeling and abnormal collagen accumulation. SO<sub>2</sub> treatment significantly improves pulmonary fibrosis and pulmonary arterial remodeling. Overexpression of the key enzymes associated with endogenous SO<sub>2</sub> generation, aspartate aminotransferase (AAT) 1 and AAT2, mimics the effect of SO<sub>2</sub> on the down-regulation of collagen synthesis, while AAT1 or AAT2 knockdown aggravates abnormal collagen accumulation in vascular smooth muscle cells (VSMCs). SO<sub>2</sub> also improves myocardial fibrosis induced by myocardial infarction or diabetes in rats, and inhibits myocardial fibroblast proliferation and migration by the extracellular signal-regulated protein kinase pathway. The mechanisms underlying the inhibition of fibrosis by SO<sub>2</sub> are related to its antioxidant effect, anti-inflammation effect,

improvement in cardiac function, and cell proliferation inhibition. Therefore, SO<sub>2</sub> has a potential therapeutic effect on fibrosis.

**Key words:** Sulfur dioxide, Fibrosis, Therapy

### Introduction

Tissue fibrosis is an important problem in clinical diseases and up to 45% of all deaths are due to fibrotic diseases in various organs (Wynn, 2008). When fibrosis occurs in organs, such as the lung, heart, liver and kidney, it will cause organ dysfunction and can even lead to death. The abnormal excessive and irregular deposition of extracellular matrices (ECM) is the key cause of fibrosis (Nogueira et al., 2017; Park et al., 2019; Weiskirchen et al., 2019). Fibroblasts are the principal cells involved in the production of ECM in fibrosis. ECM is composed of matrix proteins, such as collagen and elastin, structural glycoproteins (proteoglycans) and carbohydrates (hyaluronan) (Gressner and Weiskirchen, 2006; Tarbit et al., 2019). Collagens are the main component of ECM (Tarbit et al., 2019). Many pathological conditions (such as inflammation, hypoxia or hypertension, etc.) can induce the formation of pro-inflammatory cytokines, chemokines and angiogenic factors, which activate fibroblasts or mesenchymal cells to generate collagens and non-structural proteins in the ECM (Weiskirchen et al., 2019). However, no effective method for treating fibrosis has been developed.

A gasotransmitter is a small gaseous molecule,

*Offprint requests to:* Professor Hong Cui, Department of Pediatrics, Beijing Friendship Hospital, Capital Medical University, Yong'an Str. No. 95 West District, Beijing 100050, PR China. e-mail: cuihong100@sina.com or Professor Jun-Bao Du, Department of Pediatrics, Peking University First Hospital, Beijing, PR China. e-mail: junbaodu@126.com  
DOI: 10.14670/HH-18-169

endogenously generated, which exhibits special functionality at physiological concentrations, and has specific molecular or cell targets (Wang, 2002). Nitric oxide, carbon monoxide, hydrogen sulfide (H<sub>2</sub>S), and sulfur dioxide (SO<sub>2</sub>) are considered the four main gasotransmitters (Huang et al., 2016a; Yu et al., 2018). SO<sub>2</sub> is considered a toxic environmental gas. However, SO<sub>2</sub> is endogenously generated and plays an important role in mammal physiology. SO<sub>2</sub> can relax aortic rings and down-regulate blood pressure in rats (Du et al., 2008; Lu et al., 2012). SO<sub>2</sub> also improves cardiac function, reduces myocardial ischemia reperfusion, and decreases myocardial injury induced by isoproterenol (Liang et al., 2011; Wang et al., 2011). Additionally, SO<sub>2</sub> improves pulmonary vascular remodeling, and reduces pulmonary fibrosis and myocardial fibrosis (Yu et al., 2016; Wang et al., 2018). Further studies have shown that aspartate aminotransferase (AAT, a key enzyme in SO<sub>2</sub> generation) 1 or AAT2 overexpression can mimic the inhibitory effect of SO<sub>2</sub> on fibrosis, while AAT1 or AAT2 knock down aggravates fibrosis in pulmonary artery smooth muscle cells (Huang et al., 2016b). Therefore, SO<sub>2</sub> may be a potential therapy for fibrosis.

### Endogenous SO<sub>2</sub> and its associated enzymes in fibrosis pathogenesis

SO<sub>2</sub> is endogenously generated by L-cysteine, then catalyzed by cysteine dioxygenase and converted to L-cysteine sulfinate, then transaminated by AAT, converted to beta-sulfinylpyruvate, and finally spontaneously decomposed to pyruvate and SO<sub>2</sub> (Singer and Kearney, 1956; Stipanuk, 1986). The content of SO<sub>2</sub> is 15.54±1.68 μmol/L and its key generating enzyme (AAT) activity is 87±18 U/L in plasma (Du et al., 2008). SO<sub>2</sub> and AAT are extensively found in arteries, including the aorta, pulmonary, mesenteric, tail, and renal arteries. SO<sub>2</sub> and AAT are also extensively found in other tissues, such as heart, brain, pancreas, lung, kidney, spleen and liver tissues (Luo et al., 2011). In pulmonary hypertension induced by monocrotaline in rats, the content of SO<sub>2</sub> and activity of AAT were significantly increased in lung tissues (Jin et al., 2008). An inhibitor of SO<sub>2</sub> production, L-aspartate-beta-hydroxamate (HDX) reduced the generation of SO<sub>2</sub> and activity of AAT in monocrotaline-treated rats, but aggravated pulmonary artery remodeling, and further increased the relative medium thickness and relative media area of small pulmonary arteries. Inhibition of SO<sub>2</sub> by HDX significantly increased the percentage of pulmonary muscularized arteries, enhanced the content of collagen I and III, and up-regulated the mRNA expression of pre-collagen I and III in lung tissues of hypoxic rats (Sun et al., 2010). Inhibition of SO<sub>2</sub> by HDX also aggravated myocardial fibrosis in streptozotocin-induced diabetic rats (Liu et al., 2017). Therefore, the endogenous SO<sub>2</sub>/AAT pathway is involved in the process of fibrosis in pulmonary hypertension induced by hypoxia or monocrotaline in rats.

### The effect of SO<sub>2</sub> on lung fibrosis

SO<sub>2</sub> improved lung fibrosis in monocrotaline-treated rats. The contents of collagen I and III, and hydroxyproline were increased in lung tissues of monocrotaline-treated rats, and increased following HDX treatment, but decreased after SO<sub>2</sub> treatment (Yu et al., 2016). The mRNA expression of procollagen I and III in lung tissues were significantly up-regulated in monocrotaline-treated rats, and were higher following HDX treatment, but decreased after SO<sub>2</sub> treatment. In addition, the protein expression of matrix metalloproteinase-13 (MMP-13) and tissue inhibitor of metalloproteinase-1 (TIMP-1) were also detected in lung tissues. In monocrotaline-treated rats, the expression of MMP-13 and TIMP-1 was significantly increased, and was higher following SO<sub>2</sub> treatment, but decreased after HDX treatment. The MMP-13/TIMP-1 ratio was also significantly increased in monocrotaline-treated rats, and was higher following SO<sub>2</sub> treatment, but decreased after HDX treatment (Yu et al., 2016). Therefore, endogenous SO<sub>2</sub> in lung fibrosis in monocrotaline-treated rats, as well as exogenous SO<sub>2</sub>, reduces abnormal collagen accumulation in lung tissue. In addition, SO<sub>2</sub> regulates the balance between MMP-13 and TIMP-1, increases collagen degradation, and limits pulmonary fibrosis.

### The effect of SO<sub>2</sub> on vascular remodeling

Vascular remodeling often occurs alongside hypertension. In pulmonary hypertension induced by monocrotaline in rats, the relative medium thickness and relative median area of the medial and small pulmonary arteries were significantly increased (Jin et al., 2008). However, they decreased following treatment with SO<sub>2</sub> (Na<sub>2</sub>SO<sub>3</sub>/NaHSO<sub>3</sub>, 72.3 mg/kg subcutaneously injected every day for three weeks). Therefore, SO<sub>2</sub> ameliorates vascular remodeling. A similar effect for SO<sub>2</sub> was also found in pulmonary hypertension induced by hypoxia in rats. SO<sub>2</sub> significantly decreased the high percentage of muscularized arteries induced by hypoxia (Sun et al., 2010). In hypoxia- or monocrotaline-treated rats, the content of collagen I and III, and the mRNA expressions of pre-collagen I and III were all significantly increased in lung tissues, but decreased after treatment with SO<sub>2</sub> (Sun et al., 2010; Yu et al., 2016). The mRNA expressions of MMP-13 and TIMP-1 were also significantly up-regulated in small and medium pulmonary arteries of hypoxia rats, and decreased following treatment with SO<sub>2</sub>. The ratio of MMP-13 to TIMP-1 mRNA obviously decreased in hypoxia rats, but increased following treatment with SO<sub>2</sub> (Sun et al., 2010). MMP-13 stimulated the degradation of collagen, and TIMP-1 inhibited this degradation. The imbalance between MMP-13 and TIMP-1 contributes to vascular remodeling. Therefore, the inhibitory effect of SO<sub>2</sub> on pulmonary artery remodeling is related to the inhibition of abnormal collagen synthesis and induction of collagen degradation.

In cultured VSMCs under the conditions of AAT1 or AAT2 overexpression, the level of endogenous SO<sub>2</sub> and SO<sub>2</sub> in the supernatant were both significantly increased (Huang et al., 2016b). The protein expressions of collagen I and III were highly increased by transforming growth factor (TGF)-β1 in VSMCs, and even increased following AAT1 or AAT2 knockdown, but decreased due to AAT1 or AAT2 overexpression. The mRNA expressions of procollagen I and III were also significantly increased by TGF-β1 in VSMCs, and further increased by AAT1 or AAT2 knockdown, but decreased by AAT1 or AAT2 overexpression (Huang et al., 2016b). Additionally, the mRNA and protein expressions of MMP-13 were significantly decreased by TGF-β1 in VSMCs, and further decreased by AAT1 or AAT2 knockdown, but increased by AAT1 or AAT2 overexpression. The mRNA and protein expression of TIMP-1 were significantly up-regulated by TGF-β1 in VSMCs, and increased by AAT1 or AAT2 knockdown, but decreased by AAT1 or AAT2 overexpression. Therefore, the beneficial effect of SO<sub>2</sub> on vascular remodeling is related to inhibition of collagen synthesis and the promotion of collagen degradation.

SO<sub>2</sub> can also inhibit VSMC proliferation induced by serum (Liu et al., 2014). The higher expression of proliferating cell nuclear antigen (PCNA) can be significantly down-regulated by AAT1 or AAT2 overexpression, but increased in serum-induced VSMCs. Exogenous and endogenous SO<sub>2</sub> repressed cell proliferation, but did not affect cell apoptosis of serum-induced VSMCs. In platelet-derived growth factor (PDGF)-BB treated VSMCs, the high expression of cyclin D1 induced by PDGF-BB was inhibited by AAT1 or AAT2 overexpression, and increased by AAT1 or AAT2 silencing (Liu et al., 2014). SO<sub>2</sub> inhibited the phosphorylation of extracellular regulated protein kinase (Erk) 1/2 (Thr202/Tyr204), mitogen-activated protein kinase (MAPK) kinase 1/2 (Ser217/221) and the phosphorylation of RAF proto-oncogene serine/threonine-protein kinase (c-Raf, Ser338) induced by PDGF-BB. SO<sub>2</sub> also promoted the cyclic adenosine monophosphate/protein kinase A (cAMP/PKA) pathway and inhibited activation of c-Raf. A selective inhibitor of PKA, H-89, reversed the inhibitory effect of SO<sub>2</sub> on proliferation. Therefore, SO<sub>2</sub> inhibits the proliferation of VSMCs by down-regulation via the cAMP/PKA pathway.

### SO<sub>2</sub> reduces cardiac fibrosis

Recent studies also showed that SO<sub>2</sub> alleviates cardiac fibrosis in myocardial infarction rats or diabetic rats (Liu et al., 2017; Wang et al., 2018). In myocardial remodeling induced by myocardial infarction in rats, the internal diameter of the left ventricle was increased, and the left ventricle wall became thinner (Wang et al., 2018). The study found that SO<sub>2</sub> treatment significantly increased the thickness of the wall, and decreased the enlarged internal diameter of the left ventricle. SO<sub>2</sub>

reduced the area of cardiac fibrosis, and reduced the higher contents of myocardial collagen I and III induced by myocardial infarction. Additionally, in rats with myocardial infarction, the activity of MMP-9 was increased, but TIMP-1 was decreased. SO<sub>2</sub> treatment decreased the activity of MMP-9 and increased the level of TIMP-1. The MMP-9/TIMP-1 ratio was increased in rats with myocardial infarction, but decreased following SO<sub>2</sub> treatment (Wang et al., 2018). In addition, SO<sub>2</sub> significantly reduced myocardial fibrosis in streptozotocin-induced diabetic rats, while the SO<sub>2</sub> synthase inhibitor, HDX, aggravated myocardial fibrosis (Liu et al., 2017). SO<sub>2</sub> also decreased the expression of MMP9, MMP24 and TIMP1 in myocardial tissues, but increased these factors in the HDX-treated groups. Therefore, endogenous SO<sub>2</sub> mediates the process of myocardial fibrosis in streptozotocin-induced diabetic rats. SO<sub>2</sub> reduces cardiac fibrosis by reducing abnormal collagen accumulation and promoting collagen degradation.

### Mechanisms of SO<sub>2</sub> in fibrosis

Fibrosis is the abnormal accumulation and deposition of ECM, including collagen and elastin, structural glycoproteins and carbohydrates (Weiskirchen et al., 2019). Pathological injuries provoke inflammatory cell infiltration and activation, and generate many harmful molecules, such as reactive oxygen species and inflammatory and fibrogenic cytokines. These molecules stimulate fibroblast or mesenchymal cell activation and produce a large amount of ECM. However, SO<sub>2</sub> has been shown to reduce these injurious effects of fibrosis development (Fig. 1). Studies have demonstrated that SO<sub>2</sub> exerts antioxidant and anti-inflammatory activity, reduces collagen synthesis, induces collagen degradation, and inhibits cell apoptosis during fibrosis development (Sun et al., 2010; Jin et al., 2013b; Li et al., 2016; Yu et al., 2016; Yang et al., 2018). Signaling pathways are also involved in the inhibitory effect of SO<sub>2</sub> in fibrosis (Huang et al., 2016b).

### SO<sub>2</sub> and hypertension

Hypertension can cause mechanical stretching of vascular tissues and plays an important role in fibrosis. Blood pressure was significantly increased in spontaneously hypertensive rats, but decreased following SO<sub>2</sub> treatment (Lu et al., 2012). In addition, SO<sub>2</sub> administration significantly decreased the high systolic pulmonary artery pressure in pulmonary hypertension induced by abdominal aorta-inferior vena cava shunting (Luo et al., 2013). SO<sub>2</sub> also decreased the mean pulmonary artery pressure induced by monocrotaline in rats (Jin et al., 2008). Additionally, SO<sub>2</sub> treatment significantly decreased high mean arterial pressure in D-gal-induced aging rats (Dai et al., 2018). Inhibition of endogenous SO<sub>2</sub> by HDX caused even higher mean pulmonary artery pressure in pulmonary hypertension

induced by monocrotaline in rats (Jin et al., 2008). These data indicate that endogenous  $SO_2$  is involved in the development of pulmonary hypertension and that the decrease in hypertension by  $SO_2$  is related to its vasorelaxation effect.  $SO_2$  can dose-dependently (1–12 mmol/L) relax isolated aortic rings and may be a physiologic endothelium-derived relaxing factor (Du et al., 2008; Meng et al., 2012; Wang et al., 2017b).

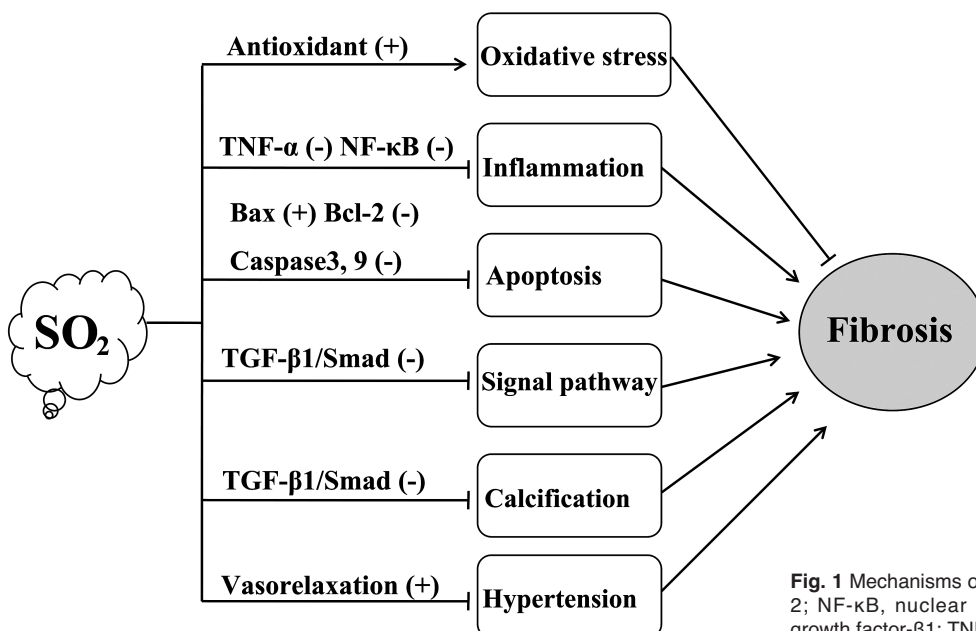
#### $SO_2$ and cardiac function

$SO_2$  improves cardiac function, which may contribute to an improvement in cardiac fibrosis. In diabetic rats with myocardial fibrosis,  $SO_2$  improved cardiac function, increased ejection fraction and fractional shortening, and decreased left ventricular end-diastolic dimension (LVEDD) and left ventricular end-systolic dimension (LVESD) (Liu et al., 2017). In rats with cardiac dysfunction induced by cecal ligation and puncture, mean arterial blood pressure, left ventricular systolic pressure and maximum contraction velocity were significantly decreased, but these parameters were increased after  $SO_2$  treatment (Yang et al., 2018). Left ventricular end-diastolic pressure was also increased in these rats, but decreased after  $SO_2$  treatment. Therefore,  $SO_2$  improves cardiac function in rats with sepsis-induced cardiac dysfunction. In rats with myocardial injury induced by isopropylarterenol,  $SO_2$  treatment reversed the inhibition of left ventricular ejection fraction and fraction shortening, and reduced the anterior wall thickness of the diastolic left ventricle (Jin et al., 2013a). In addition,  $SO_2$  reduced the high protein expression of GRP78, caspase-12 and CHOP

(endoplasmic reticulum stress markers) induced by isopropylarterenol in myocardial tissues (Chen et al., 2012). In isolated rat hearts with ischemia reperfusion injury, cardiac function was improved by  $SO_2$  preconditioning, but was reversed by 2-(2-Amino-3-methoxyphenyl)-4H-1-benzopyran-4-one (PD98059, an inhibitor of the ERK1/2 signaling pathway) (Huang et al., 2013). Therefore, the ERK1/2 signaling pathway mediates the beneficial effect of  $SO_2$  preconditioning on cardiac function in isolated rat hearts with ischemia reperfusion injury.

#### $SO_2$ and fibroblasts

$SO_2$  is also involved in collagen synthesis by fibroblasts. In cultured pulmonary artery fibroblasts, mechanical stretching was induced by a computer-controlled cyclic strain unit. The results showed that mechanical stretching decreased the content of  $SO_2$  in the supernatant, and reduced the protein expression of AAT1 and activity of AAT in primary pulmonary artery fibroblasts (Liu et al., 2016). However, the expressions of collagen I and III were significantly up-regulated. AAT1 knockdown mimicked these effects of mechanical stretching. In contrast, AAT1 overexpression reversed the effects of mechanical stretching described above, increased the protein expression of AAT1 and activity of AAT, increased the concentration of  $SO_2$  in the supernatant, and down-regulated the expression of collagen I and III in primary pulmonary artery fibroblasts. Mechanical stretching also increased the expression of TGF- $\beta$ 1 and phosphorylation of Smad2/3, which were inhibited by AAT1 overexpression and



**Fig. 1** Mechanisms of  $SO_2$  in Fibrosis. Bcl-2, B-cell lymphoma-2; NF- $\kappa$ B, nuclear factor-kappa B; TGF- $\beta$ 1, transforming growth factor- $\beta$ 1; TNF- $\alpha$ , tumor necrosis factor- $\alpha$ .



## Therapeutic effect of SO<sub>2</sub> on fibrosis

enhanced by AAT1 knockdown. SB431542, an inhibitor of the TGF- $\beta$ 1/Smad2/3 pathway, reversed the effect of increased collagen I and III expression induced by AAT1 knockdown (Liu et al., 2016). Therefore, the effect of mechanical stretching on abnormal collagen accumulation is mediated by the SO<sub>2</sub>/AAT1 pathway via the TGF- $\beta$ 1/Smad2/3 signaling pathway.

Empty lentivirus or AAT knockdown lentivirus was used to determine the role of endogenous SO<sub>2</sub>/AAT in rat myocardial fibroblasts (Zhang et al., 2018b). The study found that the expression of AAT was reduced by 79% in AAT1 knockdown myocardial fibroblasts, and the production of SO<sub>2</sub> was reduced by 84.18% in the cell supernatant (Zhang, et al., 2018b). The expression of AAT was reduced by 77% in AAT2 knockdown myocardial fibroblasts, and the production of SO<sub>2</sub> was reduced by 82.49% in the cell supernatant. Therefore, an endogenous SO<sub>2</sub>/AAT pathway exists in rat myocardial fibroblasts. In AAT1 or AAT2 knockdown rat myocardial fibroblasts, the indicator of proliferation, PCNA expression and the activity of CCK-8 were significantly enhanced, and cell migration was also promoted. SO<sub>2</sub> treatment reduced the high expression of PCNA and CCK-8 activity, and inhibited cell migration in AAT1 or AAT2 knockdown fibroblasts. The phosphorylation of extracellular receptor kinase was highly expressed in AAT1 or AAT2 knockdown lentivirus transfected myocardial fibroblasts. An inhibitor of the MAPK pathway, PD98059, inhibited increased fibroblast proliferation and migration induced by AAT1 or AAT2 knockdown (Zhang et al., 2018b). Therefore, the effect of SO<sub>2</sub> on myocardial fibroblast proliferation inhibition is related to down-regulation of the ERK signal pathway.

### SO<sub>2</sub> and oxidative stress

Oxidative stress is involved in the process of fibrosis and is considered to be a therapeutic target for pulmonary fibrosis (Fois et al., 2018; Hosseinzadeh et al., 2018). In pulmonary hypertension induced by monocrotaline in rats, SO<sub>2</sub> increased antioxidative enzymes (such as plasma SOD and glutathione peroxidase [GSH-Px]) in lung tissues and plasma (Jin et al., 2008). The effect of SO<sub>2</sub> on the improvement in pulmonary vascular remodeling may be related to a reduction in oxidative stress. In acute lung injury induced by oleic acid, SO<sub>2</sub> significantly reduced the formation of oxygen radicals (OH<sup>-</sup> and H<sub>2</sub>O<sub>2</sub>), decreased lipid peroxidation (malondialdehyde, MDA), and enhanced the antioxidant content (total antioxidant capacity, catalase, SOD and GSH-Px) in rat lungs. The protein expression of SOD1 and SOD2 in lung tissues was also up-regulated by SO<sub>2</sub> treatment (Chen et al., 2015). SO<sub>2</sub> also increased the activities of myocardial SOD and GSH in isoproterenol-induced rat myocardial injury, up-regulated the mRNA expression of SOD2 and GSH-Px1, and decreased the formation of H<sub>2</sub>O<sub>2</sub> and O<sub>2</sub><sup>-</sup> (Liang et al., 2011). A similar effect of SO<sub>2</sub> was

also found in myocardial ischemia reperfusion injury. In rats with myocardial ischemia reperfusion injury, SO<sub>2</sub> preconditioning increased the levels of SOD, GSH and GSH-Px in plasma, and increased the expression of SOD1 in myocardium, but decreased the formation of MDA (Jin et al., 2013a). SO<sub>2</sub> also reduced the increased content of H<sub>2</sub>O<sub>2</sub> and MDA in the aorta of D-galactose (D-gal)-induced aging rats, and enhanced the activity of SOD (Dai et al., 2018). Therefore, SO<sub>2</sub> enhances the activities of antioxidative enzymes, decreases free radical formation, and reduces oxidative stress.

### SO<sub>2</sub> and inflammation

Inflammation is also involved in pulmonary vascular structural remodeling in pulmonary hypertension induced by hyperoxia (Sun et al., 2010). The expression of nuclear factor-kappa B (NF- $\kappa$ B) and intercellular adhesion molecule 1 (ICAM1) were increased in the pulmonary arteries of hypoxic rats, but significantly decreased following SO<sub>2</sub> treatment (Sun et al., 2010). In EA.hy926 cells or primary endothelial cells, with cystathionine- $\gamma$ -lyase (CSE, a key enzyme in H<sub>2</sub>S generation) knockdown, the generation of SO<sub>2</sub> was increased and the activity of AAT was enhanced. An H<sub>2</sub>S donor (sodium hydrosulfide, NaHS) reversed these effects of CSE knockdown, and reduced the generation of SO<sub>2</sub> and AAT activity (Zhang et al., 2018a). Therefore, the generation of SO<sub>2</sub> can be inhibited by endogenous H<sub>2</sub>S. In CSE knockdown primary human umbilical vein endothelial cells, the expression of ICAM1 and the phosphorylation ratios of inhibitor of NF- $\kappa$ B $\alpha$  (I $\kappa$ B $\alpha$ ) and NF $\kappa$ B p65 were increased. Interleukin (IL) 6 and tumor necrosis factor (TNF)- $\alpha$  were also increased in the supernatant (Zhang et al., 2018a). HDX treatment increased the phosphorylation of NF $\kappa$ B p65 and the formation of inflammatory cytokines. A similar effect of SO<sub>2</sub> was also found in monocrotaline-treated rats. The SO<sub>2</sub> content and AAT activity were increased in lung tissues in monocrotaline-treated rats, but down-regulated in those rats treated with NaHS. Expression of the inflammatory factor, ICAM1, and the levels of IL6 and TNF- $\alpha$  were increased, but H<sub>2</sub>S production was decreased in lung tissues of monocrotaline-treated rats. HDX treatment increased the protein expression of ICAM1, IL6 and TNF- $\alpha$  in lung tissues of monocrotaline-treated rats. Therefore, increased SO<sub>2</sub> generation plays a compensatory role to inhibit inflammation induced by H<sub>2</sub>S/CSE deficiency.

SO<sub>2</sub> can also regulate inflammation in adipose tissues. SO<sub>2</sub> and AAT are extensively found in adipose tissues, including perivascular, aorta, heart, lung and liver adipose tissues (Zhang et al., 2016). In cultured 3T3-L1 adipocytes infected by the AAT1 adenovirus, the generation of SO<sub>2</sub> and the protein expression of AAT1 were significantly increased. AAT1 overexpression significantly reduced the high level of monocyte chemoattractant protein-1 (MCP-1) and IL-8 induced by TNF- $\alpha$  in the cell supernatant. AAT1 overexpression

also reduced the phosphorylation of NF- $\kappa$ B p65 and inhibited the degradation and phosphorylation of I $\kappa$ B $\alpha$  in 3T3-L1 adipocytes treated with TNF- $\alpha$ . This effect was reversed by lentivirus delivered shRNA, which caused AAT1 knockdown. AAT1 knockdown increased the phosphorylation of NF- $\kappa$ B p65, and aggravated the phosphorylation and degradation of I $\kappa$ B $\alpha$ . The generation of SO<sub>2</sub> and the protein expression of AAT1 were down-regulated by AAT1 knockdown. High levels of MCP-1 and IL-8 in the supernatant of 3T3-L1 adipocytes were promoted by TNF- $\alpha$ . Inhibitors of NF- $\kappa$ B activation (pyrrolidine dithiocarbamate or Bay 11-7082) successfully reversed this effect of AAT1 knockdown (Zhang et al., 2016; Chen et al., 2017). The NF- $\kappa$ B p65 pathway may mediate the effect of AAT1 deficiency on MCP-1 and IL-8 secretion. Therefore, SO<sub>2</sub> inhibits adipocyte inflammation induced by TNF- $\alpha$ .

### SO<sub>2</sub> and apoptosis

Cell apoptosis also plays important roles in the process of fibrosis. In spontaneously hypertensive rats, SO<sub>2</sub> treatment significantly increased the apoptosis index, down-regulated the expression of Bcl-2, and up-regulated the expressions of Fas and caspase-3 in aortic tissues (Zhao et al., 2008). In addition, the proliferation index of VSMCs was inhibited. Therefore, the effect of SO<sub>2</sub> on alleviating structural remodeling is related to the induction of apoptosis and inhibition of smooth muscle cell proliferation. On the other hand, in rats with myocardial injury induced by isopropylarterenol, SO<sub>2</sub> treatment significantly reduced the high percentage of TUNEL-positive cells, and decreased the activities of caspase-3 and caspase-9 in left ventricular tissues (Jin et al., 2013b). SO<sub>2</sub> also increased the inhibition of protein expression of B-cell lymphoma-2 (Bcl-2), and decreased the increased protein expression of Bax induced by isopropylarterenol in myocardial tissues. SO<sub>2</sub> preconditioning also reduced apoptotic cardiomyocytes and decreased the apoptotic index in rats with myocardial ischemia reperfusion injury (Wang et al., 2011). SO<sub>2</sub> preconditioning (1–10  $\mu$ mol/kg) reduced neuronal injury and decreased neuronal apoptosis in rats with febrile seizures (Han et al., 2014). SO<sub>2</sub> also reduced cardiomyocyte apoptosis, decreased the expression of myocardial Bax, caspase-3 and caspase-9, and increased the expression of Bcl 2 in streptozotocin-induced diabetic rats (Liu et al., 2017). SO<sub>2</sub> significantly reduced apoptotic cardiomyocytes, decreased the high Bax/Bcl-2 ratio and activity of caspase-3 in sepsis rats induced by cecal ligation and puncture (Yang et al., 2018). SO<sub>2</sub> also reduced hippocampal necrotic and apoptotic cells in cerebral ischemia reperfusion injury by increasing the antioxidant enzymes, SOD and glutathione (Zare Mehrjerdi et al., 2018). However, in epileptic rats, inhibition of SO<sub>2</sub> delayed the occurrence of apoptosis, but did not prevent apoptosis of neurons (Niu et al., 2018). The protein expression of PCNA was inhibited by

overexpression of AAT1, and up-regulated by AAT1 knockdown in cultured pulmonary artery smooth muscle cells or pulmonary artery fibroblasts (Yu et al., 2016). SO<sub>2</sub> stimulated the apoptosis of pulmonary artery smooth muscle cells, but did not affect pulmonary artery fibroblasts.

### Signaling pathways involved in SO<sub>2</sub> effects on fibrosis

The TGF- $\beta$ 1/Smad pathway is involved in the process of vascular fibrosis. The protein expression of TGF- $\beta$ 1 was significantly up-regulated in lung tissues of monocrotaline-treated rats, and increased following HDX treatment, but significantly decreased after SO<sub>2</sub> treatment (Yu et al., 2016). The expression of TGF- $\beta$ 1 was also significantly increased in pulmonary arteries of monocrotaline-treated rats as shown by immunohistochemical staining, but decreased following SO<sub>2</sub> treatment. The protein expressions of collagen I and III were significantly increased by TGF- $\beta$ 1 treatment in cultured primary rat pulmonary artery fibroblasts, but these effects were reversed by SO<sub>2</sub> treatment. SO<sub>2</sub> also inhibited the high phosphorylation of Smad2/3 and serine phosphorylation of TGF $\beta$  receptor I kinase (T $\beta$ RI) induced by TGF- $\beta$ 1 in pulmonary artery fibroblasts (Yu et al., 2016). AAT1 overexpression increased AAT activity and the content of SO<sub>2</sub> in the supernatant, inhibited the high phosphorylation of Smad2 and Smad3, and inhibited the high content of collagen I and III induced by TGF- $\beta$ 1 in cultured pulmonary artery fibroblasts. In contrast, AAT1 knockdown inhibited AAT activity and the content of SO<sub>2</sub> in the supernatant, aggravated the high phosphorylation of Smad2 and Smad3, and further increased the levels of collagen I and III induced by TGF- $\beta$ 1 in cultured pulmonary artery fibroblasts. Therefore, the TGF- $\beta$ 1/Smad pathway is involved in the alleviation of pulmonary vascular fibrosis by SO<sub>2</sub>.

AAT1 or AAT2 overexpression inhibited the high phosphorylation of Smad2 and Smad3 induced by TGF- $\beta$ 1 in VSMCs, and reduced the mRNA expression of plasminogen activator inhibitor-1 (Huang et al., 2016a,b). In contrast, AAT1 or AAT2 knockdown aggravated the high phosphorylation of Smad2/3 induced by TGF- $\beta$ 1. Additionally, the high phosphorylation of T $\beta$ RI induced by TGF- $\beta$ 1 was inhibited. AAT1 or AAT2 overexpression was aggravated by AAT1 or AAT2 knockdown. These data indicate that endogenous SO<sub>2</sub> inhibited the phosphorylation of Smad2/3 and T $\beta$ RI in collagen remodeling of VSMCs induced by TGF- $\beta$ 1. Furthermore, SB431542, an inhibitor of the TGF- $\beta$ 1/Smad pathway, decreased the high mRNA and protein expression of procollagen I and III induced by AAT1 or AAT2 knockdown. Therefore, the effect of SO<sub>2</sub> on inhibiting collagen remodeling is mediated, to some extent, by the TGF- $\beta$ 1/Smad signaling pathway. The protein expression of TGF- $\beta$  and phosphorylation of Smad2/3 were significantly increased in rats with aortic

calcification, but decreased following SO<sub>2</sub> treatment (Li et al., 2016). SO<sub>2</sub> also inhibited the expression of Runx2 (osteochondrogenic marker), and decreased the high protein expression of TGF-β and phosphorylation of Smad2/3 induced by CaCl<sub>2</sub> in cultured VSMCs (A7r5 cells) (Li et al., 2016). Therefore, the effect of SO<sub>2</sub> on reducing rat vascular calcification is related to the TGF-β1/Smad pathway.

SO<sub>2</sub> treatment significantly improved aortic endothelium-dependent relaxation induced by acetylcholine (Dai et al., 2018). In addition, SO<sub>2</sub> treatment significantly decreased the high level of angiotensin I, and reduced the protein overexpression of angiotensin II type 1 receptor in thoracic aorta tissues in D-gal-induced aging rats. Therefore, the attenuating effect of SO<sub>2</sub> on endothelial dysfunction is related to inhibition of the angiotensin II/angiotensin II type 1 receptor pathway in D-gal-induced aging rats. In rats with pulmonary hypertension induced by high pulmonary blood flow, SO<sub>2</sub> significantly increased the content of H<sub>2</sub>S in lung tissues, and increased the expression of key enzymes involved in H<sub>2</sub>S generation, such as cystathionine-γ-lyase, mercaptopyruvate transsulfurase and cystathionine-β-synthase in small pulmonary arteries as shown by immunostaining (Luo et al., 2013). Therefore, SO<sub>2</sub> up-regulated the endogenous H<sub>2</sub>S pathway in rats with pulmonary hypertension induced by high pulmonary blood flow. SO<sub>2</sub> can also down-regulate increased protein expression of p-Akt1/2/3, up-regulate protein expression of caspase-3 and E-cadherin, and down-regulate mRNA expression of DKK1 in the Wnt pathway in the lung tissues of rats with hypoxia-induced pulmonary hypertension (Luo et al., 2018). These findings indicate that SO<sub>2</sub> can inhibit the proliferation of pulmonary artery smooth muscle cells and improve pulmonary arteriolar remodeling via the Dkk1/Wnt signaling pathway.

### Perpectives and challenges

Fibrosis is a pathological feature of most chronic diseases and causes organ dysfunction. It is characterized by excessive accumulation and deposition of ECM. The imbalance between collagen synthesis and degradation plays a key role in fibrosis. SO<sub>2</sub> is generated endogenously and endogenous SO<sub>2</sub> mediates fibrosis development. Inhibition of endogenous SO<sub>2</sub> by HDX aggravates small pulmonary artery remodeling and increases abnormal collagen accumulation. AAT1 or AAT2 knockdown aggravates abnormal collagen accumulation in VSMCs. On the other hand, SO<sub>2</sub> treatment significantly alleviates pulmonary fibrosis and pulmonary vascular remodeling. SO<sub>2</sub> decreases abnormal collagen accumulation and induces collagen degradation. The mechanism by which SO<sub>2</sub> attenuates fibrosis is related to its antioxidant effect, anti-inflammation effect, beneficial effect on cardiac function, and inhibitory effect on pulmonary artery smooth muscle cell proliferation.

However, the following limitations are associated with this study, (1) Fibrosis can occur in different organs and this study only demonstrated the effect of SO<sub>2</sub> in the lung, heart and vascular system. The effects of SO<sub>2</sub> on other organs (such as liver, kidney, and intestine) require further investigation. (2) The SO<sub>2</sub> donors used in this study were Na<sub>2</sub>SO<sub>3</sub>/NaHSO<sub>3</sub>, and other donors such as SO<sub>2</sub> water solution were not studied. (3) There is a network of gasotransmitters (such as nitric oxide, carbon monoxide and hydrogen sulfide) in mammals. SO<sub>2</sub> plays a compensatory role to inhibit inflammation induced by H<sub>2</sub>S/CSE deficiency (Zhang, et al., 2018a). It is unknown whether there is cross-talk and the relationship between SO<sub>2</sub> and other gasotransmitters in fibrosis is unclear.

SO<sub>2</sub> drugs can be activated to release SO<sub>2</sub> (Pardeshi et al., 2018; Wang and Wang, 2018), and SO<sub>2</sub> prodrugs have been shown to have an antimycobacterial effect and to inhibit methicillin-resistant *Staphylococcus aureus* (Malwal et al., 2012; Pardeshi et al., 2015; Wang et al., 2017a). The effectiveness and safety of SO<sub>2</sub> prodrugs require further study. In future studies, we will determine the effects and mechanisms of SO<sub>2</sub> on fibrosis in other organs. Suitable doses for SO<sub>2</sub> prodrugs also need further investigation. Thus, further experimentation is required to determine the potential therapeutic effect of SO<sub>2</sub> in fibrosis.

---

*Acknowledgements.* We thank International Science Editing for its linguistic assistance during the preparation of this manuscript.

---

### References

- Chen S., Du J., Liang Y., Ochs T., Liu D., Zhu L., Tang X., Tang C. and Jin H. (2012). Sulfur dioxide inhibits excessively activated endoplasmic reticulum stress in rats with myocardial injury. *Heart Vessels* 27, 505-516.
- Chen S., Zheng S., Liu Z., Tang C., Zhao B., Du J. and Jin H. (2015). Endogenous sulfur dioxide protects against oleic acid-induced acute lung injury in association with inhibition of oxidative stress in rats. *Lab. Invest.* 95, 142-156.
- Chen S., Huang Y., Liu Z., Yu W., Zhang H., Li K., Yu X., Tang C., Zhao B., Du J. and Jin H. (2017). Sulphur dioxide suppresses inflammatory response by sulphenylating nf-kappab p65 at cys(38) in a rat model of acute lung injury. *Clin. Sci.* 131, 2655-2670.
- Dai J., Liu R., Zhao J. and Zhang A. (2018). Sulfur dioxide improves endothelial dysfunction by downregulating the angiotensin ii/at1r pathway in d-galactose-induced aging rats. *J. Renin Angiotensin Aldosterone Syst.* 19, 1470320318778898.
- Du S.X., Jin H.F., Bu D.F., Zhao X., Geng B., Tang C.S. and Du J.B. (2008). Endogenously generated sulfur dioxide and its vasorelaxant effect in rats. *Acta Pharmacol. Sin.* 29, 923-930.
- Fois A.G., Paliogiannis P., Sotgia S., Mangoni A.A., Zinellu E., Pirina P., Carru C. and Zinellu A. (2018). Evaluation of oxidative stress biomarkers in idiopathic pulmonary fibrosis and therapeutic applications: A systematic review. *Respir. Res.* 19, 51.
- Gressner A.M. and Weiskirchen R. (2006). Modern pathogenetic concepts of liver fibrosis suggest stellate cells and TGF-beta as



- major players and therapeutic targets. *J. Cell. Mol. Med.* 10, 76-99.
- Han Y., Yi W., Qin J., Zhao Y., Zhang J. and Chang X. (2014). Dose-dependent effect of sulfur dioxide on brain damage induced by recurrent febrile seizures in rats. *Neurosci. Lett.* 563, 149-154.
- Hosseinzadeh A., Javad-Moosavi S.A., Reiter R.J., Yarahmadi R., Ghaznavi H. and Mehrzadi S. (2018). Oxidative/nitrosative stress, autophagy and apoptosis as therapeutic targets of melatonin in idiopathic pulmonary fibrosis. *Expert Opin. Ther. Targets.* 22, 1049-1061.
- Huang P., Sun Y., Yang J., Chen S., Liu A.D., Holmberg L., Huang X., Tang C., Du J. and Jin H. (2013). The ERK1/2 signaling pathway is involved in sulfur dioxide preconditioning-induced protection against cardiac dysfunction in isolated perfused rat heart subjected to myocardial ischemia/reperfusion. *Int. J. Mol. Sci.* 14, 22190-22201.
- Huang Y., Tang C., Du J. and Jin H. (2016a). Endogenous sulfur dioxide: A new member of gasotransmitter family in the cardiovascular system. *Oxid. Med. Cell. Longev.* 2016, 8961951.
- Huang Y., Shen Z., Chen Q., Huang P., Zhang H., Du S., Geng B., Zhang C., Li K., Tang C., Du J. and Jin H. (2016b). Endogenous sulfur dioxide alleviates collagen remodeling via inhibiting tgfbeta/smad pathway in vascular smooth muscle cells. *Sci. Rep.* 6, 19503.
- Jin H.F., Du S.X., Zhao X., Wei H.L., Wang Y.F., Liang Y.F., Tang C.S. and Du J.B. (2008). Effects of endogenous sulfur dioxide on monocrotaline-induced pulmonary hypertension in rats. *Acta Pharmacol. Sin.* 29, 1157-1166.
- Jin H., Wang Y., Wang X., Sun Y., Tang C. and Du J. (2013a). Sulfur dioxide preconditioning increases antioxidative capacity in rat with myocardial ischemia reperfusion (I/R) injury. *Nitric Oxide* 32, 56-61.
- Jin H., Liu A.D., Holmberg L., Zhao M., Chen S., Yang J., Sun Y., Chen S., Tang C. and Du J. (2013b). The role of sulfur dioxide in the regulation of mitochondrion-related cardiomyocyte apoptosis in rats with isopropylarterenol-induced myocardial injury. *Int. J. Mol. Sci.* 14, 10465-10482.
- Li Z., Huang Y., Du J., Liu A.D., Tang C., Qi Y. and Jin H. (2016). Endogenous sulfur dioxide inhibits vascular calcification in association with the tgfbeta/smad signaling pathway. *Int. J. Mol. Sci.* 17, 266.
- Liang Y., Liu D., Ochs T., Tang C., Chen S., Zhang S., Geng B., Jin H. and Du J. (2011). Endogenous sulfur dioxide protects against isoproterenol-induced myocardial injury and increases myocardial antioxidant capacity in rats. *Lab. Invest.* 91, 12-23.
- Liu D., Huang Y., Bu D., Liu A.D., Holmberg L., Jia Y., Tang C., Du J. and Jin H. (2014). Sulfur dioxide inhibits vascular smooth muscle cell proliferation via suppressing the erk/map kinase pathway mediated by camp/pka signaling. *Cell Death Dis.* 5, e1251.
- Liu J., Yu W., Liu Y., Chen S., Huang Y., Li X., Liu C., Zhang Y., Li Z., Du J., Tang C., Du J. and Jin H. (2016). Mechanical stretching stimulates collagen synthesis via down-regulating SO<sub>2</sub>/AAT1 pathway. *Sci. Rep.* 6, 21112.
- Liu M., Liu S., Tan W., Tang F., Long J., Li Z., Liang B., Chu C. and Yang J. (2017). Gaseous signalling molecule SO<sub>2</sub> via hippomst pathway to improve myocardial fibrosis of diabetic rats. *Mol. Med. Rep.* 16, 8953-8963.
- Lu W., Sun Y., Tang C., Ochs T., Qi J., Du J. and Jin H. (2012). Sulfur dioxide derivatives improve the vasorelaxation in the spontaneously hypertensive rat by enhancing the vasorelaxant response to nitric oxide. *Exp. Biol. Med. (Maywood).* 237, 867-872.
- Luo L., Chen S., Jin H., Tang C. and Du J. (2011). Endogenous generation of sulfur dioxide in rat tissues. *Biochem. Biophys. Res. Commun.* 415, 61-67.
- Luo L., Liu D., Tang C., Du J., Liu A.D., Holmberg L. and Jin H. (2013). Sulfur dioxide upregulates the inhibited endogenous hydrogen sulfide pathway in rats with pulmonary hypertension induced by high pulmonary blood flow. *Biochem. Biophys. Res. Commun.* 433, 519-525.
- Luo L., Hong X., Diao B., Chen S. and Hei M. (2018). Sulfur dioxide attenuates hypoxia-induced pulmonary arteriolar remodeling via Dkk1/Wnt signaling pathway. *Biomed. Pharmacother.* 106, 692-698.
- Malwal S.R., Sriram D., Yogeewari P. and Chakrapani H. (2012). Synthesis and antimycobacterial activity of prodrugs of sulfur dioxide (SO<sub>2</sub>). *Bioorg. Med. Chem. Lett.* 22, 3603-3606.
- Meng Z., Yang Z., Li J. and Zhang Q. (2012). The vasorelaxant effect and its mechanisms of sodium bisulfite as a sulfur dioxide donor. *Chemosphere* 89, 579-584.
- Niu M., Han Y., Li Q. and Zhang J. (2018). Endogenous sulfur dioxide regulates hippocampal neuron apoptosis in developing epileptic rats and is associated with the perk signaling pathway. *Neurosci. Lett.* 665, 22-28.
- Nogueira A., Pires M.J. and Oliveira P.A. (2017). Pathophysiological mechanisms of renal fibrosis: A review of animal models and therapeutic strategies. *In Vivo* 31, 1-22.
- Pardeshi K.A., Ravikumar G. and Chakrapani H. (2018). Esterase sensitive self-immolative sulfur dioxide donors. *Org. Lett.* 20, 4-7.
- Pardeshi K.A., Malwal S.R., Banerjee A., Lahiri S., Rangarajan R. and Chakrapani H. (2015). Thiol activated prodrugs of sulfur dioxide (SO<sub>2</sub>) as mrsa inhibitors. *Bioorg. Med. Chem. Lett.* 25, 2694-2697.
- Park S., Nguyen N.B., Pezhouman A. and Ardehali R. (2019). Cardiac fibrosis: Potential therapeutic targets. *Transl. Res.* 209, 121-137.
- Singer T.P. and Kearney E.B. (1956). Intermediary metabolism of L-cysteinesulfinic acid in animal tissues. *Arch. Biochem. Biophys.* 61, 397-409.
- Stipanuk M.H. (1986). Metabolism of sulfur-containing amino acids. *Annu. Rev. Nutr.* 6, 179-209.
- Sun Y., Tian Y., Prabha M., Liu D., Chen S., Zhang R., Liu X., Tang C., Tang X., Jin H. and Du J. (2010). Effects of sulfur dioxide on hypoxic pulmonary vascular structural remodeling. *Lab. Invest.* 90, 68-82.
- Tarbit E., Singh I., Peart J.N. and Rose/Meyer R.B. (2019). Biomarkers for the identification of cardiac fibroblast and myofibroblast cells. *Heart Fail. Rev.* 24, 1-15.
- Wang R. (2002). Two's company, three's a crowd: can H<sub>2</sub>S be the third endogenous gaseous transmitter? *FASEB J.* 16, 1792-1798.
- Wang W. and Wang B. (2018). SO<sub>2</sub> donors and prodrugs, and their possible applications: A review. *Front. Chem.* 6, 559.
- Wang X.B., Huang X.M., Ochs T., Li X.Y., Jin H.F., Tang C.S. and Du J.B. (2011). Effect of sulfur dioxide preconditioning on rat myocardial ischemia/reperfusion injury by inducing endoplasmic reticulum stress. *Basic Res. Cardiol.* 106, 865-878.
- Wang W., Ji X., Du Z. and Wang B. (2017a). Sulfur dioxide prodrugs: Triggered release of SO<sub>2</sub> via a click reaction. *Chem. Commun.* 53, 1370-1373.
- Wang X.B., Cui H. and Du J.B. (2017b). Sulfur dioxide: A physiologic endothelium-derived relaxing factor. *Histol. Histopathol.* 32, 21-26.
- Wang X.B., Cui H. and Du J.B. (2018). Sulfur dioxide ameliorates rat myocardial fibrosis by inhibiting endoplasmic reticulum stress. *Histol. Histopathol.* 33, 1089-1097.
- Weiskirchen R., Weiskirchen S. and Tacke F. (2019). Organ and tissue fibrosis: Molecular signals, cellular mechanisms and translational



*Therapeutic effect of SO<sub>2</sub> on fibrosis*

- implications. *Mol. Aspects Med.* 65, 2-15.
- Wynn T.A. (2008). Cellular and molecular mechanisms of fibrosis. *J. Pathol.* 214, 199-210.
- Yang L., Zhang H. and Chen P. (2018). Sulfur dioxide attenuates sepsis-induced cardiac dysfunction via inhibition of NLRP3 inflammasome activation in rats. *Nitric Oxide.* 81, 11-20.
- Yu W., Liu D., Liang C., Ochs T., Chen S., Chen S., Du S., Tang C., Huang Y., Du J. and Jin H. (2016). Sulfur dioxide protects against collagen accumulation in pulmonary artery in association with downregulation of the transforming growth factor beta1/smad pathway in pulmonary hypertensive rats. *J. Am. Heart Assoc.* 5, e003910.
- Yu W., Jin H., Tang C., Du J. and Zhang Z. (2018). Sulfur-containing gaseous signal molecules, ion channels and cardiovascular diseases. *Br. J. Pharmacol.* 175, 1114-1125.
- Zare Mehrjerdi F., Shoshtari A., Mohseni F., Khastar H., Norozi P., Asadi Y., Dadkhah M. and Khaksari M. (2018). Sulfur dioxide reduces hippocampal cell death and improves learning and memory deficits in a rat model of transient global ischemia/reperfusion. *Iran J. Basic Med. Sci.* 21, 998-1003.
- Zhang H., Huang Y., Bu D., Chen S., Tang C., Wang G., Du J. and Jin H. (2016). Endogenous sulfur dioxide is a novel adipocyte-derived inflammatory inhibitor. *Sci. Rep.* 6, 27026.
- Zhang D., Wang X., Tian X., Zhang L., Yang G., Tao Y., Liang C., Li K., Yu X., Tang X., Tang C., Zhou J., Kong W., Du J., Huang Y. and Jin H. (2018a). The increased endogenous sulfur dioxide acts as a compensatory mechanism for the downregulated endogenous hydrogen sulfide pathway in the endothelial cell inflammation. *Front. Immunol.* 9, 882.
- Zhang L.L., Du J.B., Tang C.S., Jin H.F. and Huang Y.Q. (2018b). Inhibitory effects of sulfur dioxide on rat myocardial fibroblast proliferation and migration. *Chinese Med. J.* 131, 1715-1723.
- Zhao X., Jin H.F., Tang C.S. and Du J.B. (2008). Effects of sulfur dioxide, on the proliferation and apoptosis of aorta smooth muscle cells in hypertension: Experiments with rats. *Zhonghua yi xue za zhi* 88, 1279-1283.

Accepted October 11, 2019