

## Review

# Sulfur dioxide: a physiologic endothelium-derived relaxing factor

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**Summary.** The gasotransmitter nitric oxide was classified as the first endothelium-derived relaxant factor, and opened a new era in cardiovascular research. Another small gas, sulfur dioxide (SO<sub>2</sub>), can also be generated endogenously in mammals. Recent studies have shown that SO<sub>2</sub> may play important roles in the cardiovascular system. At low concentrations, the vasodilatory effect of SO<sub>2</sub> is endothelium-dependent. The vasodilation induced by an endothelium-derived relaxant factor is achieved by the opening of potassium channels, and hyperpolarization of the membranes of vascular smooth muscle cells. This feature is in accordance with that of SO<sub>2</sub>. The vasodilatory effect of SO<sub>2</sub> is related to the opening of adenosine triphosphate-sensitive potassium channels and high-conductance calcium-activated potassium channels. The 3'-5'-cyclic guanosine monophosphate pathway and activation of nitric oxide synthase are also involved in the endothelium-derived relaxant factor effect of SO<sub>2</sub>. The vasodilatory effect of gaseous SO<sub>2</sub> is much stronger than that of its derivatives (bisulfite and sulfite). It is suggested that SO<sub>2</sub> may be a candidate endothelium-derived relaxant factor, which could lead to a new era of research into cardiovascular disease in mammals.

**Key words:** Sulfur dioxide, Endothelium-derived relaxing factor, Gasotransmitter, Biology

### Introduction

Endothelial dysfunction plays important roles in the pathology of vascular diseases such as diabetes mellitus, atherosclerosis, and hypertension. Endothelial cells regulate basic vascular tone and reactivity by releasing a series of relaxing and contracting factors.

Nitric oxide (NO) and prostacyclin are believed to be endothelium-derived relaxing factors (EDRFs) (Radomski et al., 1987). However, abolition of the production of NO and PGI<sub>2</sub> does not prevent the endothelium-dependent relaxing effect (Scotland et al., 2005). Therefore, an unknown substance termed “endothelium-derived hyperpolarizing factor” (EDHF) might be present in blood vessels. Several candidates have been proposed to be EDHF: potassium (K<sup>+</sup>) channels, epoxyeicosatrienoic acids, carbon monoxide (CO), hydrogen sulfide (H<sub>2</sub>S), hydrogen peroxide, anandamide, citrulline, and ammonia (Feletou and Vanhoutte, 2007, 2009). However, the nature of EDHF is incompletely understood.

NO, CO and H<sub>2</sub>S have been considered to be EDRF or EDHF, and all are “gasotransmitters”. Gasotransmitters are small molecules that: can pass freely across cell membranes; are generated endogenously; can be regulated; have special biologic functions at physiologic concentrations; have specific biologic targets (Wang, 2002).

Besides NO, CO and H<sub>2</sub>S, another small gas

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molecule, sulfur dioxide (SO<sub>2</sub>) can also be generated endogenously in mammalian cells, and pass freely across cell membranes (Stipanuk, 1986, 2004). SO<sub>2</sub> can be generated endogenously from the metabolism of sulfur-containing amino acids in mammals (Stipanuk, 1986; 2004). Balazy et al. (2003) found that levels of carbonyl sulfide and SO<sub>2</sub> could be enhanced by acetylcholine and calcium ionophore (A23187). The vasodilatory effect of opening adenosine triphosphate-sensitive potassium (K<sub>ATP</sub>) channels might be another candidate for EDHF (Balazy et al., 2003).

For a long time, SO<sub>2</sub> was considered to be a toxic gas and air pollutant. SO<sub>2</sub> is detrimental to many organs (Meng, 2003; Meng and Bai, 2004). Recently, the cardiovascular effects of SO<sub>2</sub> have attracted considerable interest. SO<sub>2</sub> is generated in different tissues (stomach, intestine, myocardium, brain, pancreas, lung, kidney, spleen, and liver) (Luo et al., 2011). SO<sub>2</sub> has physical effects on the cardiovascular system: vasorelaxation, regulation of cardiac function and inhibition of activity of L-type calcium channels (Zhang et al., 2011). In addition, the pathophysiologic effects of SO<sub>2</sub> have been documented: amelioration of pulmonary hypertension (Jin et al., 2008; Sun et al., 2010; Luo et al., 2013); reduction of the myocardial injury induced by isoproterenol (Liang et al., 2011); inhibition of the development of atherosclerotic lesions (Li et al., 2011); reduction of myocardial ischemia–reperfusion injury (Wang et al., 2011b; Huang et al., 2013; Luo et al., 2013; Zhao et al., 2013); reduction of lung injury (Chen et al., 2015; Zhao et al., 2015); inhibition of vascular smooth muscle cells (VSMCs) proliferation (Liu et al., 2014); protection of neurons from the toxicity caused by febrile seizures (Han et al., 2014). Therefore, SO<sub>2</sub> is considered to be another novel gasotransmitter in mammals (Wang et al., 2010, 2011a, 2014, 2015; Chen et al., 2011; Huang et al., 2016a,b). This review indicates that SO<sub>2</sub> may be an EDRF, which could lead to a new era of research into cardiovascular disease in mammals.

### Endothelial production of SO<sub>2</sub>

Endogenous SO<sub>2</sub> is generated from sulfur-containing amino acids such as L-cysteine and then oxidized to L-

cysteinesulfinate by cysteine dioxygenase. L-cysteinesulfinate is converted to β-sulfinylpyruvate due to transamination by aspartate aminotransferase, and decomposes spontaneously to pyruvate and SO<sub>2</sub> (Shapiro, 1977; Stipanuk et al., 1990). SO<sub>2</sub> dissociates to its derivatives (bisulfite and sulfite [NaHSO<sub>3</sub>/Na<sub>2</sub>SO<sub>3</sub>] at 1:3 m/m in plasma), and is oxidized to a sulfate, then is excreted in urine (Stipanuk, 1986) (Fig. 1).

As a key SO<sub>2</sub>-generating enzyme, the location of aspartate aminotransferase has been detected using *in situ* hybridization in samples of rat aorta. Expression of aspartate aminotransferase-1 mRNA and aspartate aminotransferase-2 mRNA in endothelial cells is much greater than those of VSMCs. The SO<sub>2</sub> concentration is different among different tissues and arteries (Du et al., 2008; Luo et al., 2011). The highest concentration of SO<sub>2</sub> is in the aorta (5.55±0.35 μmol/g protein), followed by the pulmonary, mesenteric, tail and renal arteries (3.27±0.21, 2.67±0.17, 2.50±0.20 and 2.23±0.19 μmol/g protein, respectively) (Du et al., 2008). The SO<sub>2</sub> concentration in the plasma and aortic tissues of rats is 16.77±8.24 μM and 127.76±31.34 μM, respectively (Meng et al., 2009).

In cultured vascular endothelial cells and smooth muscle cells (SMCs), SO<sub>2</sub> generation in endothelial cells is much higher than that in SMCs (Meng et al., 2009). SO<sub>2</sub> production can be stimulated by acetylcholine fourfold compared with that treated with ethanol (solvent for acetylcholine) in incubated porcine coronary artery rings. In addition, calcium ionophore can enhance SO<sub>2</sub> production by 1.5-fold (Balazy et al., 2003). In male Wistar rats, acetylcholine can increase the generation of endogenous SO<sub>2</sub>, whereas noradrenaline inhibits these effects in thoracic aortic rings (Meng et al., 2009).

### Endothelium-dependent vasodilation induced by SO<sub>2</sub>

SO<sub>2</sub> can dose-dependently relax endothelium-intact or endothelium-denuded aortic rings in rats. SO<sub>2</sub> and its derivatives have vasodilatory effects, but the relaxation effects and their mechanism of action are different (Meng et al., 2009). Median effective concentrations (EC<sub>50</sub>) that induce a half-maximal vasodilation response for SO<sub>2</sub> are (1247.38±98.32) μM in endothelium-intact

**Table 1.** Mechanisms of endothelium-dependent vasodilation induced by SO<sub>2</sub>.

	Concentration	Mechanisms	Models
Gaseous SO <sub>2</sub>	< 450 μM (Li and Meng, 2009, Zhang and Meng, 2009)	cGMP (Li and Meng, 2009)	Rat aortic rings
		BK <sub>Ca</sub> (Zhang and Meng, 2009)	Rat aortic rings
		K <sub>ATP</sub> (Zhang et al., 2016)	Rat aorta
		L-Ca <sup>2+</sup> (Zhang et al., 2016)	Rat aorta
		sGC/cGMP/PKG pathway (Yao et al., 2016)	Rat aortic rings
SO <sub>2</sub> derivative	<2 mM (Wang et al., 2009)	NOS (Wang et al., 2009)	Rat aortic rings
		cGMP (Meng et al., 2012)	Rat aortic rings
		BK <sub>Ca</sub> (Meng et al., 2012)	Rat aortic rings

BK<sub>Ca</sub> channel, big calcium activated potassium channel; cGMP, 3′-5′-cyclic guanosine monophosphate; K<sub>ATP</sub>, ATP-sensitive potassium channel; NOS, nitric oxide synthase; PKA, protein kinase A; and L-Ca<sup>2+</sup>, L type of calcium channel; sGC, guanylate cyclase; PKG, protein kinase G

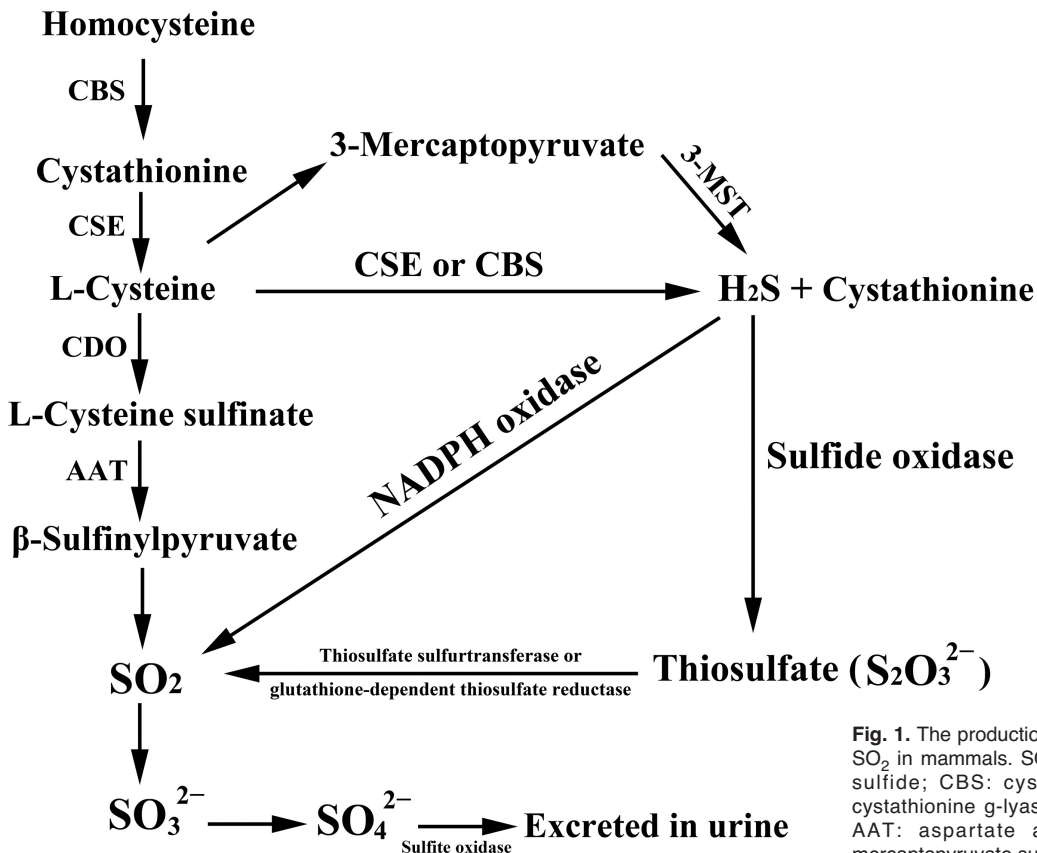
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and (1321.89±89.67) μM in endothelium-denuded rat aortic rings (Zhang and Meng, 2009). EC<sub>50</sub> of SO<sub>2</sub> derivatives is 7.28±0.12 mM (Zhang and Meng, 2009). Therefore, the vasodilatory effect of gaseous SO<sub>2</sub> is much stronger than that of its derivatives.

Gaseous SO<sub>2</sub> exerts a vasodilatory effect in an endothelium-dependent manner at low concentrations (<450 μM, Table 1), and in an endothelium-independent manner at high concentrations (>500 μM) (Meng et al., 2009; Zhang and Meng, 2009). The vasodilatory effect of SO<sub>2</sub> gas or gas solution is similar. SO<sub>2</sub> dissolved in water is present mainly as SO<sub>2</sub> molecules (Akhmetov et al., 1983; Feletou and Vanhoutte, 2006; Zhang and Meng, 2009).

In male Wistar rats treated with SO<sub>2</sub> (3.5, 7, 14 mg/m<sup>3</sup>) 4 h every day for 30 consecutive days, the SO<sub>2</sub> concentrations described above could increase the expression of high-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels (BK<sub>Ca</sub>) subunits alpha and beta-1 in rat aortas in vivo. Exposure of rats to a concentration of 14 mg/m<sup>3</sup> of SO<sub>2</sub> can up-regulate expression of K<sub>ATP</sub> channel subunits Kir6.1, Kir6.2, and the sulfonylurea 2B receptor in rat aortas, with no effects on levels of the sulfonylurea 2A receptor (Zhang et al., 2016). Simultaneously, SO<sub>2</sub> downregulates expression of L-type calcium channel

subunits Cav1.2 and Cav1.3 (Zhang et al., 2016). However, vasodilation induced by SO<sub>2</sub> (30 or 300 μM) can be inhibited by iberiotoxin (selectively inhibits current through BK<sub>Ca</sub>) in endothelium-intact aortic rings, but this phenomenon is not affected by apamin (selectively inhibits current through low-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels) (Zhang and Meng, 2009). These data suggest that the vasodilation caused by SO<sub>2</sub> at low concentrations might be mediated by BK<sub>Ca</sub> but not by low-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels. A recent study also showed that soluble guanylate cyclase (sGC), cyclic guanosine monophosphate (cGMP), and protein kinase G (PKG) pathways were also involved in the vasorelaxation of SO<sub>2</sub> (1–300 μM). Thiol reductants dithiothreitol could reverse the dimerization of sGC and PKG and vasodilation induced by SO<sub>2</sub>. These data indicate that dimerization of sGC and PKG was related to the vasorelaxation effect of SO<sub>2</sub> (Yao et al., 2016). The vasorelaxation induced by SO<sub>2</sub> (1500 μM) could be partially inhibited by nifedipine (L-type calcium channel blocker). Additionally, the vasorelaxation could also be inhibited by tetraethylammonium or glibenclamide. These data indicate the vasodilation of SO<sub>2</sub> was related to the K<sub>ATP</sub> channel, L-type calcium channel and calcium-influx and release pathway (Zhang and Meng,



**Fig. 1.** The production and metabolism of endogenous SO<sub>2</sub> in mammals. SO<sub>2</sub>: sulfur dioxide; H<sub>2</sub>S: hydrogen sulfide; CBS: cystathionine β-synthase; CSE: cystathionine γ-lyase; CDO: cysteine dioxygenase; AAT: aspartate aminotransferase; 3MST: 3-mercaptopyruvate sulfurtransferase.

2009).

The relaxation wrought by SO<sub>2</sub> derivatives (0.5 and 1 mM) in endothelium-intact rings is stronger than that in endothelium-denuded rings. The NO synthase (NOS) inhibitor NG-nitro-L-arginine methyl ester (100 μM) can abolish the effects of SO<sub>2</sub> derivatives described above, but has no effect at higher doses (2–8 mM) (Wang et al., 2009). These data suggest that the NOS pathway is involved in the endothelium-dependent vasodilation caused by SO<sub>2</sub> derivatives. In addition, the vasodilatory effect of SO<sub>2</sub> derivatives is mediated by the 3'-5'-cyclic guanosine monophosphate (cGMP) pathway (Li and Meng, 2009). Interestingly, there are some differences in the SO<sub>2</sub> derivatives Na<sub>2</sub>SO<sub>3</sub> and NaHSO<sub>3</sub> in terms of dilation of aortic rings (Meng et al., 2012). NaHSO<sub>3</sub> can cause dilation of rat aortic rings in a concentration-dependent manner (100–4000 μM), whereas Na<sub>2</sub>SO<sub>3</sub> exerts a contractive effect at 500–1000 μM and relaxant effect at high concentrations (2000–4000 μM). The EC<sub>50</sub> of NaHSO<sub>3</sub> and Na<sub>2</sub>SO<sub>3</sub> on aortic rings is 2326±78.56 μM and 4100±87.68 μM, respectively. Vasodilation caused by NaHSO<sub>3</sub> is endothelium-dependent at low concentrations (≤500 μM) but endothelium-independent at high concentrations (≥1000 μM). At low concentrations, the vasodilatory effect of NaHSO<sub>3</sub> is mediated (at least in part) by the cGMP pathway and is related to BK<sub>Ca</sub> channels, but not to the actions of prostaglandins, protein kinase C or the 3'-5'-cyclic adenosine monophosphate pathway (Meng et al., 2012). These data suggest that the vasorelaxant effect of sodium bisulfite was much stronger than that of sodium sulfite. The endothelium-dependent vasorelaxant effect of sodium bisulfite was related to the cGMP pathway and BK<sub>Ca</sub> channels at low concentrations.

### Resemblance of SO<sub>2</sub> to EDHF

The vasodilation caused by EDHF is through open potassium channels, and involves hyperpolarization of the membranes VSMCs by close voltage-dependent calcium channels (Shimokawa and Morikawa, 2005). The effect of EDHF is mainly caused by small-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channel and partially by intermediate-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channel. SO<sub>2</sub> and EDHF have some common characteristics.

(1) Studies have shown that SO<sub>2</sub> induces vasodilation by opening K<sub>ATP</sub> channels and might be EDHF (Balazy et al., 2003). In rat aortas, SO<sub>2</sub> can increase the expression of K<sub>ATP</sub> channel subunits Kir6.1, Kir6.2, and the sulfonylurea 2B receptor (Zhang et al., 2016). These data indicate the relaxation effect of SO<sub>2</sub> was related to the opening K<sub>ATP</sub> channel.

(2) In health, Ca<sup>2+</sup> influx contributes to the contraction of VSMCs. At high concentration, SO<sub>2</sub> can inhibit expression of the subunits of the L-type calcium channels Cav1.2 and Cav1.3, which inhibit the contraction mediated by Ca<sup>2+</sup> during depolarization of sarcolemmal membranes (Zhang et al., 2016). SO<sub>2</sub> can also elicit vasodilation by opening BK<sub>Ca</sub> channels.

Expression of BK<sub>Ca</sub> channel subunits alpha and beta-1 can be up-regulated by SO<sub>2</sub> in rat aortas *in vivo* (Zhang et al., 2016).

### Interaction of SO<sub>2</sub> and other EDRFs

The gasotransmitters NO and H<sub>2</sub>S were considered as EDRFs in mammals (Fleming and Busse, 1999; Wang, 2009). Both of them play important roles in vascular tone regulation. Recent studies showed that there is some crosstalk between SO<sub>2</sub>, NO and H<sub>2</sub>S.

The vasodilatory effect of SO<sub>2</sub> can be enhanced under the presence of the NO donor sodium nitroprusside (3 or 5 nM), the EC<sub>50</sub> of which are 598 μM and 217 μM SO<sub>2</sub>, respectively (Li and Meng, 2009). In healthy rat, the EC<sub>50</sub> of the relaxing effect induced by SO<sub>2</sub> is 1247.38±98.32 μM, so the vasodilatory effect of SO<sub>2</sub> is enhanced by NO by nearly six-fold. Interestingly, the vasodilatory effect of NO can also be enhanced by low concentrations of SO<sub>2</sub>. In the presence of 3 μM SO<sub>2</sub>, the relaxant effect of NO is enhanced at various concentrations. The EC<sub>50</sub> of the vasodilatory effect induced by NO is 210 nM in the absence of SO<sub>2</sub>, whereas it is 34 nM in the presence of 3 μM SO<sub>2</sub>. The NOS inhibitor NG-nitro-L-arginine methyl ester (100 μM) can reverse the relaxant effect induced by SO<sub>2</sub> derivatives (0.5 and 1 mM) in endothelium-intact rings (Wang et al., 2009). In spontaneously hypertensive rats, SO<sub>2</sub> affects the NO level in aortic tissues. The vasodilatory effect of SO<sub>2</sub> can be enhanced by the presence of NO in isolated aortic rings (Lu et al., 2012). These data suggest that the actions of NO mediate the vasodilatory effect of SO<sub>2</sub> to some extent.

In the pulmonary hypertension induced by high pulmonary blood flow, H<sub>2</sub>S production as well as the mRNA and protein expression of cystathionine-γ-lyase (CSE) is increased in pulmonary tissues in SO<sub>2</sub>-exposed rats. These data suggest that SO<sub>2</sub> can up-regulate an endogenous H<sub>2</sub>S pathway (Luo et al., 2013). In a model of atherosclerosis in rats, SO<sub>2</sub> treatment can significantly increase H<sub>2</sub>S levels in plasma and aortic tissues and is associated with a reduction in the number of atherosclerotic lesions. Therefore, SO<sub>2</sub> can enhance H<sub>2</sub>S production in rats with atherosclerosis (Li et al., 2011). These data suggest that SO<sub>2</sub> has some crosstalk with other EDRFs.

### Perspectives and challenges

Accumulating data of the vascular effect of SO<sub>2</sub> suggest that SO<sub>2</sub> may be a new EDRF or EDHF. This hypothesis is based on five major pieces of evidence. Firstly, SO<sub>2</sub> can be generated in cultured normal endothelial cells and SMCs, and the level of SO<sub>2</sub> in endothelial cells is much higher than that in SMCs (Du et al., 2008). Secondly, at physiologic concentrations, SO<sub>2</sub> has an endothelium-dependent vasodilatory effect. The vasodilatory effect of SO<sub>2</sub>, like that of NO, is mediated by a cGMP pathway and related to high-



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conductance Ca<sup>2+</sup>-activated channels (Li and Meng, 2009; Zhang and Meng, 2009). Thirdly, SO<sub>2</sub> production can be evoked by acetylcholine in endothelial cells and vascular tissues, which conforms to an EDHF being released by acetylcholine and other compounds from arteries (Feletou et al., 2003; Meng et al., 2009). Finally, SO<sub>2</sub> derivatives increase the intracellular concentration of free Ca<sup>2+</sup> in rat ventricular myocytes (Nie and Meng, 2006), which might activate big conductance Ca<sup>2+</sup>-activated channels, induce hyperpolarization of cells, and cause relaxation.

In conclusion, SO<sub>2</sub> could be endogenously generated in vascular tissues. It could be enhanced by acetylcholine and calcium ionophore. At physiological, or low, concentrations of SO<sub>2</sub>, the vasodilation effect was endothelium dependent. The mechanism was related to BK<sub>Ca</sub>, L-type calcium channel, K<sub>ATP</sub> channel and cGMP pathway. At the same time, K<sub>ATP</sub> channel subunits Kir6.1, Kir6.2, and the sulfonylurea 2B receptor, and L-type calcium channel subunits Cav1.2 and Cav1.3 may contribute to the vasorelaxation effect of SO<sub>2</sub> (Zhang et al., 2016). Additionally, the sGC/cGMP/PKG pathway, in association with sulfhydryl-dependent dimerization, was also involved in the vasodilatory effect of SO<sub>2</sub> (Yao et al., 2016). Further study showed that endogenous SO<sub>2</sub> was involved in vascular remodeling (Sun et al., 2010). Endogenous SO<sub>2</sub> could alleviate collagen remodeling and vascular calcification by inhibiting the TGF-β/Smad pathway (Huang et al., 2016a,b; Li et al., 2016). These data indicate that SO<sub>2</sub> plays important roles in the regulation of vascular activities.

As an EDRF, SO<sub>2</sub> plays an important part in the regulation of endothelium-dependent vasoactive activities. Qualification of SO<sub>2</sub> as an EDHF will hinge on electrophysiologic changes in membrane potential in vascular endothelial cells and SMCs by patch-clamp studies. A deficiency in SO<sub>2</sub> will also probably affect the pathologies of vascular-related diseases, such as atherosclerosis, coronary vascular diseases and diabetes mellitus. More extensive studies are expected to further clarify SO<sub>2</sub> as a new EDRF/EDHF. In this way, the importance of SO<sub>2</sub> in the regulation of cardiovascular function will be better appreciated. Full understanding of the vasodilator effect of SO<sub>2</sub> carries considerable importance for further study on its pharmacologic effects.

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