

Can the European abalone *Haliotis tuberculata* survive on an invasive algae? A comparison of the nutritional value of the introduced *Grateloupia turuturu* and the native *Palmaria palmata*, for the commercial European abalone industry

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Abstract The suitability of two red algae species, the introduced Grateloupia turuturu and the native Palmaria palmata, as feed for the culture of the European abalone Haliotis tuberculata, was compared over a 5-month period. Three experimental diets were tested: (1) P. palmata, (2) G. turuturu, and (3) a mixed diet of P. palmata and G. turuturu (1:1). Biochemical composition (proteins, carbohydrates, lipids, ashes) of algae was measured. No mortality was observed during the time of the experiment in any of the treatments. Growth in length and weight was higher for abalone fed with P. palmata and the mixed diet. Abalone exhibited a preference for P. palmata and showed the highest weight gain with this macroalgae (107.8 ± 7.2 %). Grateloupia turuturu disintegrated faster in abalone rearing conditions and was not suitable for significant growth of H. tuberculata in a monospecific diet. However, when it is provided in a mixed diet, abalone seem to show a higher lipid content than when fed a *P. palmata* monospecific diet. The invasive G. turuturu could be an interesting food supplement for European abalone.

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Introduction

Haliotis spp. (abalone) are of great commercial value (Shepherd and Steinberg 1992). Their high nutritional value, the flavor of their flesh, and the originality of their perforated shell lined with mother-of-pearl make of them a highly desirable seafood delicacies, particularly in Asia (Cook 2014). Due to the ever increasing demand for this product and the simultaneous overfishing of wild stocks, the farm production of abalone has increased over the past few years and reached a production of 100,352 t and 785.799 billion US dollars in 2012 (FAO 2014). The main obstacle for the production and the economic success of this product is its slow and variable growth rate (Shepherd and Steinberg 1992). Haliotis species need between 3 and 4 years to reach market size (Demetropoulos and Langdon 2004b; Shpigel et al. 1996). Abalone growth in the wild is very variable depending on season, food availability, population densities, temperature, and stress conditions (Huchette et al. 2003; Saunders et al. 2008). The growth of cultivated abalone, however, is influenced by factors linked to the culture system such as water quality, tanks design, abalone density, quantity, and quality of feed, of which feed is the most important economically in farms (Fleming and Hone 1996; Mgaya and Mercer 1995; Shpigel et al. 1999; Troell et al. 2006). After nearly 40 years of research on formulated and natural feeds, feeding of cultivated abalone is always a challenge (Fleming et al. 1996; Kirkendale et al. 2010; Mulvaney et al. 2013). This is the case in Europe where the annual production of the European

abalone, Haliotis tuberculata Linnaeus 1758, remains anecdotal (around 10 t for the last 3 years-FAO 2014), nutrition being among the main technical problems encountered by abalone growers. Macroalgae, the natural feed of abalone, which are known to deliver suitable growth rates and condition in abalone (e.g., Bansemer et al. 2014; Kirkendale et al. 2010), could contribute to a more sustainable aquaculture than formulated feed (e.g., Mulvaney et al. 2013). However, macroalgae harvesting is costly and quantitatively limited (Viera et al. 2014; Troell et al. 2006). The use of an introduced prolific macroalgae available most of the year in high quantities could be a cost-effective solution for the development of abalone aquaculture. This would also help with product diversification in some countries where shellfish industry is largely dominated by oyster production and is facing severe financial difficulties due to mass mortalities (e.g., in France-Petton et al. 2013; Segarra et al. 2010).

Grateloupia turuturu Yamada 1941 is an invasive red alga, native from Japan, which has shown considerable expansion along the Atlantic coasts of Europe and North America, in the Mediterranean Sea, as well as in Australian and New Zealand waters (Lafontaine et al. 2011; Janiak and Whitlatch 2012). This species constitutes a resource that is not yet exploited. A recent study has shown that this macroalga was rich in total proteins and had a low total lipids content, respectively 23.0 and 2.6 % dry weight (Denis et al. 2010), comparable to those of Palmaria palmata Linnaeus 1805. Palmaria palmata allows a higher growth rate of the European abalone H. tuberculata when compared to other macroalgae or artificial feeds (Basuyaux 1997; Mai et al. 1995b; Mercer et al. 1993). The biochemical composition of G. turuturu seems to match the food requirements of the European abalone which, in dry weight, are below 5 % lipids (Mai et al. 1995a), between 24 and 34.5 % proteins (Mai et al. 1995b), combined with a high level of carbohydrates from 40 to 60 % (Fleming and Hone 1996; Thongrod et al. 2003). Furthermore, its biochemical composition appears to be seasonally more stable than in P. palmata (Denis et al. 2010). Mulvaney et al. (2013) showed that G. turuturu provided good growth rates of juvenile abalone in Haliotis rubra Leach 1814 and Haliotis laevigata Donovan 1808.

Food preferences differ between abalone species and are linked to the available food in their natural habitats (Dunstan et al. 1996). Numerous studies on different abalone species have compared survival and growth rates obtained with various macroalgae, mainly in the genera *Laminaria*, *Ulva*, *Gracilaria*, *Sargassum*, *Ecklonia*, and *Palmaria* (Alcantara and Noro 2006; Demetropoulos and Langdon 2004a; Mai et al. 1996; Naidoo et al. 2006; Qi et al. 2010; Shpigel et al. 1999; Taylor and Tsvetnenko 2004; Troell et al. 2006; Viera et al. 2005). Different abalone species fed with identical diets have shown variable growth rates due to differences in nutritional requirements (Taylor and Tsvetnenko 2004). Mai et al. (1995a) also showed that responses of same species to different diets vary according to the physical properties and chemical composition of diets. This is particularly important given that chemical composition of algae varies with species, geographic area, season, or environmental conditions (Ito and Hori 1989).

The aim of this study was to estimate the suitability of *G. turuturu* as food for the abalone *H. tuberculata*. Consumption, growth, and mortality of *H. tuberculata* fed with *P. palmata* and *G. turuturu* were compared.

Materials and methods

Experimental culture conditions

The study was conducted between May 15th and October 15 2010, in a shellfish farm in Le Croisic ($47^{\circ}17'38'' \text{ N } 2^{\circ}30'33''$ W), a seaside town in the northwest of France. Four 300 L polygonal tanks designed for abalone aquaculture containing PEHD (high density polyethylene) plastic cages ($50 \times 50 \times 5$ cm) were used. Aerated tanks were provided with sand-filtered and UV-treated seawater. The experiment was conducted in the dark to not disrupt abalone; light was only used during feedings and measurements. Water temperature was measured daily and varied from 13.2 to 19.0 °C during the experimental period.

Experimental animals and diets

Two hundred and seventy abalone H. tuberculata (≈ 2 years old) with an initial shell length of 42.6 ± 1.7 mm (mean \pm standard deviation) and an initial body weight of 11.3 ± 1.5 g were purchased from the Labadie abalone hatchery/farm in Bouin (France). In the hatchery, abalone was fed with a mixture of fresh macroalgae (e.g., Ulva spp., Laminaria spp., Palmaria palmata). Thirty abalones were randomly assigned to each triplicate of three dietary treatments. They were fed one of three experimental diets consisting of the following: (a) P. palmata, (b) G. turuturu, and (c) a mixture of P. palmaria and G. turuturu with a mass ratio of 1:1. These seaweeds were harvested every 3 days at Batz-sur-mer (47°16'34.82" N-2°29' 36.64" E; Atlantic Coast, France). Immediately after each algal harvest, abalone were fed freshly harvested algae ad libitum to ensure feed availability and quality and as to not limit abalone feeding.

Algal consumption and growth of abalone

Seaweeds were weighed before being distributed. To estimate algae biomass loss, due to degradation, six plastic cages containing algae without abalone were used as controls. When uneaten algae remained at the next feeding time, it was removed, weighed

 Table 1
 Mean biochemical composition of the two macroalgae

 provided to abalone throughout experiment

	G. turuturu	P. palmata	P value
Proteins (% dw)	22.7 ± 2.7	16.2 ± 1.7	P<0.001
Carbohydrates (% dw)	51.3 ± 4.1	63.4 ± 7.4	P<0.001
Lipids (% dw)	2.6 ± 0.2	2.0 ± 0.3	P<0.01
Ash (% dw)	24.3 ± 0.8	27.2 ± 0.4	ns

Data are expressed as mean \pm SD (n = 12). *P* value obtained after a *t* test *ns* not significant, *dw* dry weight

again, and the difference enabled estimation of the amount that had been ingested by abalone. For the mixed diet, each species was weighed separately. During the experiment, degradation of *G. turuturu* was observed (cf. Results); therefore, only the consumption of *P. palmata* could be estimated.

Shell length (L) and body weight (W) of each abalone were measured twice a month. The following abalone growth variables were calculated for all treatments:

- $\Delta L \ (\mu m \ day^{-1}) = (L_t L_i)/day$
- SGR_W (% day⁻¹) = 100 [(ln W_t-ln W_i)/day]
- SGR_L (% day⁻¹) = 100 [(ln L_t-ln L_i)/day]
- Weight gain (%) = $100 [(W_t W_i/W_i]]$
- Survival (%) = $100 (n_t/n_i)$

where ΔL is the increase in length, L_i is the initial shell length (mm), L_t is the shell length (mm) at time t, (day) is duration of experiment in days, n_i and n_t are respectively initial and final numbers of abalone, SGR is the specific growth rate, W_i is the initial body weight (g), and W_t is the body weight (g) at time t.

Biochemical analysis

Once a month, certain algae thalli were set aside, cleaned of epibionts, rinsed successively with seawater, tap water, then distilled water and frozen at -20 °C. At the end of the

experiment, five abalone from each replicate of the three dietary treatments were also frozen at -20 °C. All seaweed and abalone samples (after shell removal) were freeze-dried and ground into a powder with liquid nitrogen before being homogenized with phosphate buffer (20 mM; pH 7.1). The suspensions were then centrifuged ($25,000 \times g, 20 \min, 4 \circ C$). The resulting supernatants were the aqueous crude extract (CE) containing all the water-soluble compounds. Each sample was analyzed in triplicate for the biochemical analysis. Water-soluble proteins in the CE were analyzed following the method of Bradford (1976), and total protein content was estimated by multiplying the nitrogen content quantified by the Kjeldhal method, by a factor of 6.25 (Miller and Miller 1948). Total lipids were extracted with a mixture of dichloromethane:methanol (2:1, v:v) and were determined by the gravimetric method as described by Bligh and Dyer (1959). Ash content was determined by incineration of 1 g of each sample at 450 °C for 24 h. Total water-soluble carbohydrates in the CE were analyzed using the modified colorimetric phenol-sulphuric acid method (White et al. 1986).

Statistical analyses

Statistical analyses were performed using SigmaStat software (3.1). Biochemical compositions of algae and abalone, biometrics, and growth parameters were compared with *t* test or oneway ANOVAs. When ANOVA showed a statistical difference, a Holm-Sidak procedure was used for pairwise comparisons. For each data sample, means and standard deviations are given.

Results

Algal biochemical composition

Grateloupia turuturu and *P. palmata* showed significant differences (p < 0.001) in their biochemical composition except for ash (Table 1). Protein content was significantly higher in

 Table 2
 Biometrics of the European abalone (*H. tuberculata*) fed with the three macroalgae diets for 5 months

Diet fed to abalone					
	G. turuturu	P. palmata	G. turuturu—P. palmata (1:1)		
Initial biomass (g)	$353.3 \pm 61.2^{\rm a}$	336.8 ± 26.9^{a}	356.1 ± 67.6^{a}		
Final biomass (g)	402.6 ± 49.8^{a}	658.0 ± 96.2^{b}	715.3 ± 66.2^{b}		
Initial length (mm)	42.9 ± 2.3^{a}	42.0 ± 0.8^{a}	42.9 ± 2.3^{a}		
Final length (mm)	47.3 ± 2.0^{a}	52.7 ± 1.7^{b}	54.2 ± 0.6^{b}		
Initial weight (g)	12.7 ± 2.2^{a}	10.3 ± 0.1^{a}	$11.8 \pm 1.8^{\rm a}$		
Final weight (g)	16.5 ± 1.8^{a}	27.3 ± 3.0^b	27.8 ± 0.7^b		

Data are expressed as mean \pm SD (n = 3 for biomass and n = 90 for length and weight). Statistical results were analyzed after a one-way ANOVA followed by a Holm-Sidak test

 a,b Values in the same row with different letters are significantly different (P<0.05)





G. turuturu (22.7 \pm 2.7 % dw) than in *P. palmata* (Table 1). However, *P. palmata* showed the highest carbohydrate content with 63.4 \pm 7.4 % dw. Both, *G. turuturu* and *P. palmata*, were low in lipid contents, 2.6 and 2.0 % dw, respectively.

Consumption, survival, and growth of abalone

In abalone aquaculture tanks, *G. turuturu* deteriorated rapidly, resulting in very small pieces which made the calculation of algae consumption difficult. No conservation problem was encountered with *P. palmata*. Daily average intake of *P. palmata* per abalone was 17.6 ± 2.6 g day⁻¹ during the entire feeding trial period.

During the entire experiment, abalone mortality was not observed; the survival rate was 100 % in all treatment groups. Initial biomass of all abalone triplicates was similar, whereas final biomass was significantly higher with *P. palmata* and the mixed diet (Table 2). The initial and final biomass in abalone fed *G. turuturu* showed a difference of only 49.25 g, whereas abalone fed *P. palmata* showed a difference of 359.17 g. Biometrical and growth parameters in *H. tuberculata* were higher with the *P. palmata* diet than with *G. turuturu* (Fig. 1 and Table 3). *Palmaria palmata* and the mixed diet induced a significantly higher SGR in abalone compared to animals fed with *G. turuturu* (Fig. 2). No significant differences were observed in SGR for length and weight between abalone fed with the mixed and those fed with *P. palmata* (Fig. 2).

Biochemical composition of abalone

Abalone fed with *G. turuturu* showed the lowest protein, carbohydrate, and lipid content with 0.18 ± 0.0 , 0.7 ± 0.1 , and 27.6 ± 1.8 % dw, respectively (Table 4). However, these abalones showed the highest ash content with 11.8 ± 1.5 % dw. The mixed diet and the *P. palmata* diet were not significantly different from each other but were significantly different to the *G. turuturu* diet (Table 4).

Discussion

The biochemical composition of the two red algae *G. turuturu* and *P. palmata* harvested on the French Atlantic coast was within the range of values reported for other species of seaweeds used as feed for abalone (Mercer et al. 1993; Shpigel

Table 3Growth and survival ofjuvenile European abalone(H. tuberculata) fed with the threemacroalgae diets for 5 months

Diet fed to abalone					
	G. turuturu	P. palmata	G. turuturu—P. palmata (1:1)		
DL ($\mu m \text{ day}^{-1}$)	0.01 ± 0.01^a	0.08 ± 0.01^{b}	0.06 ± 0.02^{b}		
SGRW (% day^{-1})	0.07 ± 0.02^{a}	0.64 ± 0.03^{b}	0.60 ± 0.10^{b}		
SGRL (% day ⁻¹)	0.21 ± 0.01^{a}	0.62 ± 0.03^{b}	0.62 ± 0.11^{b}		
Weight gain (%)	8.3 ± 2.3^a	107.8 ± 7.2^{b}	97.5 ± 21.6^{b}		
Survival (%)	100	100	100		

Data are expressed as mean \pm SD (n = 3). Statistical results were analyzed after a one-way ANOVA followed by a Holm-Sidak test

SGR shell growth rate, ΔL increase in length

 a,b Values in the same row with different letters are significantly different (P < 0.05)



Fig. 2 Specific growth rate (% day⁻¹), for length (*white bars*), and weight (*black bars*), in abalone *H. tuberculata* fed with three dietary treatments over 5 months (n=90). Data with *asterisk* are significantly different from the others (one-way ANOVA, *P* value <0.05)

et al. 1999). Mercer et al. (1993) showed that balanced levels of proteins (>15 %), lipids (3-5 %), and carbohydrates (20-30 %) in macroalgae were essential for optimal growth performance of abalone species. Protein content of G. turuturu and P. palmata (around 20 %) was within the range reported previously in red seaweeds (Fleurence 1999; Galland-Irmouli et al. 1999; Morgan et al. 1980) and for the protein requirements of *H. tuberculata* which have been estimated between 25 and 35 % (Mai et al. 1995b; Mercer et al. 1993). For P. palmata, the total carbohydrates were comparable to those found in the literature, which were from 38 to 45 % dw (Morgan et al. 1980; Morgan and Simpson 1981; Rosen et al. 2000) and were higher than those found in G. turuturu (51.3 %). To our knowledge, our paper reports the total carbohydrate content of G. turuturu for the first time. The carbohydrate contents of *P. palmata* is higher than the requirements for abalone (20-30 %-Mai et al. 1995b), and P. palmata exhibited the best growth of H. tuberculata. As previously reported for other species of red seaweeds (Mabeau and Fleurence 1993), lipid contents were low, 2.6 ± 0.2 % dw for G. turuturu and 2.0 ± 0.2 % dw for P. palmata. The low lipid contents are in agreement with the requirement of abalone (Mercer et al. 1993), where high levels of dietary lipids negatively affect abalone growth (Thongrod et al. 2003). As all the

Table 4Biochemicalcomposition of abalone fed withthree different diets throughoutexperiment

diets used in this study fulfilled requirements of abalone, the different growth performances could not be attributed to the biochemical composition of the algae. In particular, the *P. palmata* diet contained a lower protein content (16.1%) than the *G. turuturu* diet (22.7%), suggesting that protein levels in the other diets may not be a limiting factor for abalone growth.

Although survival percentages were not affected by any of the treatments assayed, the abalone fed with G. turuturu showed the lowest growth performance compared to those fed with P. palmata or the mixed diet. Previous growth trials already demonstrated that P. palmata was the best algal diet for H. tuberculata (Mai et al. 1996; Mercer et al. 1993), and the lower growth reported in our experiments with G. turuturu were certainly due to a lower feed intake due to algal breakup and possible difficulty in feeding. Indeed, consumption rate in abalone could be influenced by multiple factors other than the nutritional quality of algae. Secondary metabolites are factors modifying seaweed susceptibility to herbivores and can make algae unpalatable to abalone (Yates and Peckol 1993). The texture of algae also influences the food intake (Shepherd and Steinberg 1992); in the wild, abalone thus prefer soft texture seaweeds because their radula has a limited ability to exert force against the substrate (Steneck and Watling 1982). This explanation should be rejected as G. turuturu is the softer of the two algae used in the present study (D'Archino et al. 2007; Rosen et al. 2000). The preference for P. palmata could be explained by the rapid decomposition of G. turuturu in the culture tank making it unattractive to the animals.

Abalone fed with *G. turuturu* showed the lowest lipid and carbohydrate contents which can be explained by the fact that most of the energy absorbed sustains metabolic needs, and any deficit is drawn from the abalone's nutrient reserves. By the end of the experimental period, abalone fed on *P. palmaria* and the mixed diet had gained an average weight of 17 and 16 g and an average length of 10.7 and 11.3 mm, respectively. Whereas animals fed on *G. turuturu* had grown to an average of 3.8 and 4.4 mm under the same culture conditions. Specific growth rates of the weight were lower, 0.64 ± 0.03 % for abalone fed with *P. palmata* and 0.60 ± 0.10 % for abalone fed with the mixed diet, than those reported by Mai et al. (1996) of

	Abalone	Abalone	Abalone
	G. turuturu	P. palmata	<i>G. turuturu—P. palmata</i> (1:1)
Water-soluble proteins (% dw) Water-soluble carbohydrates (% dw) Lipids (% dw) Ash (% dw)	$\begin{array}{c} 0.18 \pm 0.00^{a} \\ 0.7 \pm 0.1^{a} \\ 27.6 \pm 1.8^{a} \\ 11.8 \pm 1.5^{a} \end{array}$	$\begin{array}{c} 0.21 \pm 0.00^{b} \\ 2.7 \pm 0.2^{b} \\ 36.9 \pm 11.3^{b} \\ 8.3 \pm 1.4^{b} \end{array}$	$\begin{array}{c} 0.20 \pm 0.00^{\rm b} \\ 2.4 \pm 0.2^{\rm b} \\ 42.4 \pm 5.7^{\rm b} \\ 8.0 \pm 0.5^{\rm b} \end{array}$

Data are expressed as mean \pm SD (n=3). P value obtained after a t test. Statistical results were analyzed after a one-way ANOVA followed by a Holm-Sidak test

ns not significant, dw dry weight

^{a,b} Values in the same row with different letters are significantly different (P < 0.05)

1.0–1.3 % for *H. tuberculata* fed with five species of macroalgae (*P. palmata, Alaria esculenta, Ulva lactuca, Laminaria digitata* and *Laminaria saccharina*). Nevertheless, the SGR of the length are among the highest compared to those obtained in 130 diets examined from the peer-reviewed literature (Kirkendale et al. 2010). When comparing the weight gain, a difference of 90–100 % was found between the *G. turuturu* diet and the mixed and *P. palmata* diet. However, when the animals were fed with *G. turuturu* in a mixed diet, they exhibited higher lipid content, although not significantly, than the ones fed only with *P. palmate*. This trend suggests that *G. turuturu* provided complementary nutrients to those found in *P. palmata*.

At present, very little information has been reported on the suitability of G. turuturu as a promoter of growth and survival in abalone species. Previous studies have shown that growth was improved when abalone were fed with a combination of different algae compared with those fed a monospecific or formulated feed (Mulvaney et al. 2013; Qi et al. 2010; Viera et al. 2005). In the present study, the mixed diet sustained a slightly lower growth rate than with *P. palmata* alone. But at the same time and as shown above, abalone fed with the mixed diet presented a higher lipid content, suggesting a qualitative improvement of the diet in mixing these two red algae. Therefore, G. turuturu cannot be used alone in monospecific diets for the abalone H. tuberculata, in contrast to the study by Mulvaney et al. (2013), but could be used in a mixed diet as a dietary supplement. Moreover, various studies have shown that numerous molecules and/or extracts of G. turuturu present a biological activity such as antibacterial (Pang et al. 2006) or antiviral ones (Hudson et al. 1998). More specifically, G. turuturu has shown an antibacterial activity against the abalone pathogen Vibrio harveyi (García-Bueno et al. 2014).

To benefit from its biological activities and as a complementary source of lipids in abalone aquaculture, further investigations are required to test the use of *G. turuturu* (1) as dried fronds, (2) as a component of a formulated food that must be further developed, or (3) as fresh fronds in an integrated coculture system to allow this source of algae to be changed every day. Considering the prolific nature and the wide distribution of *G. turuturu*, this could improve the development of the commercial European abalone industry.

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References

- Alcantara L, Noro T (2006) Growth of the abalone *Haliotis diversicolor* (reeve) fed with macroalgae in floating net cage and plastic tank. Aquac Res 37:708–717
- Bansemer MS, Qin JG, Harris JO, Howarth GS, Stone DA (2014) Nutritional requirements and use of macroalgae as ingredients in abalone feed. Rev Aquac. doi:10.1111/raq.12085
- Basuyaux (1997) Study and modelisation of the physico-chemical parameters influencing the growth of the ormer (*Haliotis tuberculata*) reared in a semi-closed system. Thesis, Université de Caen.
- Bligh EG, Dyer WJ (1959) A rapid method of total lipid extraction and purification. Can J Biochem Physiol 37:911–917
- Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem 72:248–254
- Cook PA (2014) The worldwide abalone industry. Mod Econ 5:1181
- D'Archino R, Nelson WA, Zuccarello GC (2007) Invasive marine red alga introduced to New Zealand waters: first record of *Grateloupia turuturu* (Halymeniaceae, Rhodophyta). N Z J Mar Freshw Res 41: 35–42
- Demetropoulos C, Langdon CJ (2004a) Pacific dulse (*Palmaria mollis*) as a food and biofilter in recirculated, land-based abalone culture systems. Aquac Eng 32:57–75
- Demetropoulos CL, Langdon CJ (2004b) Effects of nutrient enrichment and biochemical composition of diets of *Palmaria mollis* on growth and condition of Japanese abalone, *Haliotis discus hannai* and red abalone, *Haliotis rufescens*. J Exp Mar Biol Ecol 308:185–206
- Denis C, Morançais M, Li M, Deniaud E, Gaudin P, Wielgosz-Colin G, Barnathan G, Fleurence J (2010) Study of the chemical composition of edible red macroalgae *Grateloupia turuturu* from Brittany (France). Food Chem 119:913–917
- Dunstan GA, Baillie HJ, Barrett SM, Volkman JK (1996) Effect of diet on the lipid composition of wild and cultured abalone. Aquaculture 140:115–127
- FAO (2014) Global aquaculture production 1950–2012. http://www.fao. org/fishery/statistics/global-aquaculture-production/query/en, accessed on 26 July 2014.
- Fleming AE, Hone PW (1996) Abalone aquaculture. Aquaculture 140:1– 4
- Fleming AE, Van Barneveld RJ, Hone PW (1996) The development of artificial diets for abalone: a review and future directions. Aquaculture 140:5–53
- Fleurence J (1999) Seaweed proteins: biochemical, nutritional aspects and potential uses. Trends Food Sci Technol 10:25–28
- Galland-Irmouli AV, Fleurence J, Lamghari R, Luçon M, Rouxel C, Barbaroux O, Bronowicki JP, Villaume C, Guéant JL (1999) Nutritional value of proteins from edible seaweed *Palmaria palmata* (dulse). J Nutr Biochem 10:353–359
- García-Bueno N, Decottignies P, Turpin V, Dumay J, Paillard C, Stiger-Pouvreau V, Kervarec N, Pouchus YF, Marin-Atucha A, Fleurence J (2014) Seasonal antibacterial activity of two red seaweeds, *Palmaria palmata* and *Grateloupia turuturu*, on European abalone pathogen *Vibrio harveyi*. Aquat Living Resour 27:83–89
- Huchette SH, Koh CS, Day R (2003) The effects of density on the behaviour and growth of juvenile blacklip abalone (*Haliotis rubra*). Aquac Int 11:411–428
- Hudson JB, Kim JH, Lee MK, DeWreede RE, Hong YK (1998) Antiviral compounds in extracts of Korean seaweeds: evidence for multiple activities. J Appl Phycol 10:427–434
- Ito K, Hori K (1989) Seaweed: chemical composition and potential food uses. Food Rev Int 5:101–144
- Janiak DS, Whitlatch RB (2012) Epifaunal and algal assemblages associated with the native *Chondrus crispus* (Stackhouse) and the non-

native *Grateloupia turuturu* (Yamada) in eastern Long Island sound. J Exp Mar Biol Ecol 413:38–44

- Kirkendale L, Robertson-Andersson DV, Winberg PC (2010) Review on the use and production of algae and manufactured diets as feed for sea-based abalone aquaculture in Victoria. University of Wollongong, Australia, p 188
- Lafontaine N, Mussio I, Rusig AM (2011) Production and regeneration of protoplasts from *Grateloupia turuturu* Yamada (Rhodophyta). J Appl Phycol 23:17–24
- Mabeau S, Fleurence J (1993) Seaweed in food products: biochemical and nutritional aspects. Trends Food Sci Technol 4:103–107
- Mai K, Mercer JP, Donlon J (1995a) Comparative studies on the nutrition of two species of abalone. *Haliotis tuberculata* L and *Haliotis discus hannai* Ino III response of abalone to various levels of dietary lipid. Aquaculture 134:65–80
- Mai K, Mercer JP, Donlon J (1995b) Comparative studies on the nutrition of two species of abalone. *Haliotis tuberculata* L and *Haliotis discus hannai* Ino IV optimum dietary protein level for growth. Aquaculture 136:165–180
- Mai K, Mercer JP, Donlon J (1996) Comparative studies on the nutrition of two species of abalone. *Haliotis tuberculata* L and *Haliotis discus hannai* Ino V the role of polyunsaturated fatty acids of macroalgae in abalone nutrition. Aquaculture 139:77–89
- Mercer JP, Mai KS, Donlon J (1993) Comparative-studies on the nutrition of two species of abalone, *Haliotis tuberculata* Linnaeus and *Haliotis discus hannai* Ino. I. Effects of algal diets on growth and biochemical composition. Int J Invertebr Reprod Dev 23:75–88
- Mgaya YD, Mercer JP (1995) The effects of size grading and stocking density on growth performance of juvenile abalone, *Haliotis tuberculata* Linnaeus. Aquaculture 136:297–312
- Miller GL, Miller EE (1948) Determination of nitrogen in biological materials. Anal Chem 20:481–488
- Morgan KC, Simpson FJ (1981) Cultivation of *Palmaria (Rhodymenia) palmata*: effect of high concentrations of nitrate and ammonium on growth and nitrogen uptake. Aquat Bot 11:167–171
- Morgan K, Wright J, Simpson F (1980) Review of chemical constituents of the red alga *Palmaria palmata* (dulse). Econ Bot 34:27–50
- Mulvaney W, Winberg P, Adams L (2013) Comparison of macroalgal (*Ulva* and *Grateloupia* spp.) and formulated terrestrial feed on the growth and condition of juvenile abalone. J Appl Phycol 25:815–824
- Naidoo K, Maneveldt G, Ruck K, Bolton JJ (2006) A comparison of various seaweed-based diets and formulated feed on growth rate of abalone in a land-based aquaculture system. J Appl Phycol 18:437– 443
- Pang SJ, Xiao T, Shan TF, Wang ZF, Gao SQ (2006) Evidences of the intertidal red alga *Grateloupia turuturu* in turning *Vibrio parahaemolyticus* into non-culturable state in the presence of light. Aquaculture 260:369–374
- Petton B, Pernet F, Robert R, Boudry P (2013) Temperature influence on pathogen transmission and subsequent mortalities in juvenile pacific oysters *Crassostrea gigas*. Aquac Environ Interact 3:257–273
- Qi Z, Liu H, Li B, Mao Y, Jiang Z, Zhang J, Fang J (2010) Suitability of two seaweeds, *Gracilaria lemaneiformis* and *Sargassum pallidum*,

as feed for the abalone *Haliotis discus hannai* Ino. Aquaculture 300: 189–193

- Rosen G, Langdon CJ, Evans F (2000) The nutritional value of *Palmaria* mollis cultured under different light intensities and water exchange rates for juvenile red abalone *Haliotis rufescens*. Aquaculture 185: 121–136
- Saunders TM, Mayfield S, Hogg A (2008) A simple, cost-effective, morphometric marker for characterising abalone populations at multiple spatial scales. Mar Freshw Res 59:32–40
- Segarra A, Pépin JF, Arzul I, Morga B, Faury N, Renault T (2010) Detection and description of a particular Ostreid herpesvirus 1 genotype associated with massive mortality outbreaks of Pacific oysters, Crassostrea gigas, in France in 2008. Virus Res 153:92–99
- Shepherd SA, Steinberg PD (1992) Food preference of three Australian abalone species with the review of the algal food of abalone. In: Shepherd SA, Tegner MJ, Del Proo SAG (eds) Abalone of the world. Biology, fisheries and culture. Fishing News Books, Oxford, pp 169–181
- Shpigel M, Marshall A, Lupatsch I, Mercer JP, Neori A (1996) Acclimation and propagation of the abalone *Haliotis tuberculata* in a land-based culture system in Israel. J World Aquacult Soc 27: 435–442
- Shpigel M, Ragg NL, Lupatsch I, Neori A (1999) Protein content determines the nutritional value of the seaweed Ulva lactuca L for the abalone Haliotis tuberculata L. and H. Discus hannai Ino. J Shellfish Res 18:227–234
- Steneck RS, Watling L (1982) Feeding capabilities and limitation of herbivorous molluses: a functional group approach. Mar Biol 68: 299–319
- Taylor MH, Tsvetnenko E (2004) A growth assessment of juvenile abalone *Haliotis laevigata* fed enriched macroalgae Ulva rigida. Aquac Int 12:467–480
- Thongrod S, Tamtin M, Chairat C, Boonyaratpalin M (2003) Lipid to carbohydrate ratio in donkey's ear abalone (*Haliotis asinina*, linne) diets. Aquaculture 225:165–174
- Troell M, Robertson-Andersson D, Anderson RJ, Bolton JJ, Maneveldt G, Halling C, Probyn T (2006) Abalone farming in South Africa: an overview with perspectives on kelp resources, abalone feed, potential for on-farm seaweed production and socio-economic importance. Aquaculture 257:266–281
- Viera MP, Gómez Pinchetti JL, Courtois de Vicose G, Bilbao A, Suárez S, Haroun RJ, Izquierdo MS (2005) Suitability of three red macroalgae as a feed for the abalone *Haliotis tuberculata coccinea* Reeve. Aquaculture 248:75–82
- Viera MP, Courtois de Vicose G, Fernández-Palacios H, Izquierdo M (2014) Grow-out culture of abalone *Haliotis tuberculata coccinea* Reeve, fed land-based IMTA produced macroalgae, in a combined fish/abalone offshore mariculture system: effect of stocking density. Aquac Res. doi:10.1111/are.12467
- White C, Kennedy J, Chaplin M (1986) Carbohydrate analysis: a practical approach, second ed. The practical approach series.
- Yates J, Peckol P (1993) Effects of nutrient availability and herbivory on polyphenolics in the seaweed *Fucus versiculosus*. Ecology 74: 1757–1766

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